# QUALITATIVE PHYTOCHEMICAL SCREENING OF SOME MEDICINALLY IMPORTANT ALPINE PLANTS FROM MASTUJ VALLEY, HINDU KUSH RANGE PAKISTAN

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### Abstract

Qualitative phytochemical screening of 24 medicinally important alpine plants from Mastuj Valley, Hindu Kush Range, Pakistan was carried out for the detection of alkaloids, saponins, tannins, and oil contents at three phenological stages: Prereproductive (young vegetative stage), Reproductive (flowering/fruiting stage) and Post-reproductive stages (fruit/seed maturation and dispersal stage). The selected plants were distributed among, 5 tree, 8 shrub, 7 herbaceous and 4 grassy species belonging to 16 families. Phytochemical screening indicated that 15 species contained all the 4 chemicals, 5 species had 3, 3 species had 2 and one species had only oil. Alkaloids and saponins were present in 21 species. Tannins and oils were detected in 20 species. Alkaloids were recorded in 12, 8 and 20 species in pre-reproductive, reproductive and post reproductive stages, respectively. Saponins were recorded in 21, 19 and 16 species respectively at pre-reproductive, reproductive and post reproductive stages. Pre-reproductive, reproductive and post reproductive stages respectively had tannins in 18, 15 and 16 species. Oils were recorded in 14, 15 and 18 species in the three successive phenological stages. This preliminary study suggests further quantitative analysis for the screened and other phytochemicals in different plants, their various parts to exactly pin-point the quantity of these useful chemicals under varied environmental conditions by using different solvents.

Key words: Phytochemicals, Phenology, Alpine plants, Mastuj-Hindu Kush Range.

### Introduction

Phytochemicals are naturally occurring biologically active chemical compounds found in various parts of plants. These phytochemicals are distinguished into primary and secondary metabolites. Primary metabolites including chlorophyll, proteins sugar and amino acids are directly involved in the plant growth and metabolic functions. Secondary metabolites are derived from the primary metabolic pathways. These metabolites have antifungal, antimicrobial, anti-inflammatory, anticancer, anti-tumor, anti-hypertensive and anti-proliferative nature. Secondary metabolites are also used as agrochemicals, pharmaceuticals, flavor, fragrance, food additives and pesticides (Fazili et al., 2022; Twaiji et al., 2022). These chemicals also protect plants from diseases, pollution hazards, grazing and other environmental damages. Their nature and concentration vary among plants and plant parts under a set of climatic and phenological conditions. Plants are used for medicinal purposes since time immemorial. Some 80% of the populations in developing world still rely on the traditional herbal healthcare system (Balamurugan et al., 2019). Khan et al., (2021) hinted upon the possibility of potent role of secondary chemicals in fighting COVID-19. Afzal et al., (2022) reported that total phenolics, flavonoids, tannins, alkaloids, saponins and terpenoids contents varied in different plant extracts of *Pseudocaryopteris foetida*. Lydia *et al.*, (2022) after phytochemical screening of 53 Philippine plants stated that tannins were present in 50 plants, alkaloids in 43 plants, glycosides in 33 plants, flavonoids in 31 species and saponins in 8 species. Otari et al., (2022)

investigated phenolics, glycosides, terpenes, steroids and fatty acids in Barleria terminalis and Calacanthus grandiflorus. The extracts from leaves of Neoglaziovia variegatea (Gomes et al., 2022) collected monthly round contained varying concentrations the year of anthocyanins, anthraquinones, anthracene derivatives, flavonoids, tannins, terpenes and steroids. Sharifi-Rad et al., (2022) identified 49 compounds in Teucrium polium having differences in the phytochemicals based on the growth and the phenological stages of plants. Maximum phenolics and sapnins contents were recorded at flowering stage. Aljaiyash et al., (2022) while evaluating the yield, chemical composition and biological activities of essential oil from Thymus maroccanus at different phenological stages observed that carvacrol and pcymene gradually increased from the lowest amount at the pre-flowering stage to the highest level at postflowering stage. Pereira et al., (2022) reported that the alkaloids, organic acids, flavonoids, phenols and tannins in Montrichardia linifera leaf extracts varied according to the place of collection and solvent used. Rubio et al., (2022) reported similar phytochemical compositions from naturally growing plants and micro propagated explants. Akinmoladun et al., (2022) identified various phytochemicals from Chrysophyllum albidum. Phytochemical screening of methanolic extract from leaves of Chapta lianutans revealed the presence of alkaloids, coumarins, tannins, phenolics, flavonoids, and free steroids (deSouza et al., 2022). Abbas et al., (2021) reported biologically active phenolics and flavonoids from Seriphidium oliverianum. Abeysinghe et al., (2021) identified alkaloids, flavonoids, saponins, tannins, terpenes and steroids with antioxidant and antimicrobial

activities from Murrava koenigii and Micromelum minutum. Alkaloids, saponins, flavonoids, steroids, tannins, phenolics, glycosides, oil and fats were detected in Plumbago indica (Bashir & Kumar, 2021), Ocimum sanctum (Gwari et al., 2021) and Euphorbia helioscopia and Rumex dentatus with antimicrobial, analgesic and antioxidant properties (Mehmood et al., 2021). Alkaloids were also reported in Acorus calamus, Senna alata, Solanum torvum and S. trilobatum (Mohan et al., 2021). Mokua et al., (2021) identified alkaloids, saponins, tannins, phenols and flavonoids from 14 plants locally used for treating snakebite in Kenya. Yu et al., (2021) reported several phytochemicals in fruit plants with varying concentrations among the tested species. Phytochemical screening of Echinops amplexicaulis, Ruta chalepensis and Salix subserrata (Marami et al., 2021); and Prosopis africana, Curcuma longa and Psorospermum febrifugum (Elaigwu et al., 2020) revealed alkaloids, saponins, phenols, tannins, flavonoids, phytosterols, steroids, terpenoids, and cardiac glycosides. Iheagwam et al., (2019) observed that natural ripened mangoes had higher carotenoids, phenols and terpenes compared to alkaloids, tannins and flavonoids. Many recent contemporary studies on phytochemical screening revealed the presence of various secondary plant metabolites varying in occurrence and concentration based on the plant part, species, climate, growth, phenological stage and type of solvent used for extraction (Afzal et al., 2022; Twaij et al., 2022; Sreenivasulu et al., 2022; Wawrosch & Zotchev, 2021; Onefeliet al., 2021; Feudjio et al., 2020; Choukri et al., 2020; Reaisi et al., 2019).

The survey of recent literature strongly favours the importance of phytochemical screening of medicinal plants as a primary source for commercial exploitation. It is also obvious that no such work is available on the alpine plants of Mastuj, Hindu Kush Range, Pakistan. The present endeavour was, therefore, aimed to chemically evaluate 24 common alpine plants being locally used in the traditional health care system (Hussain et al., 2007; Shah & Hussain, 2012) in the Mastuj Valley, Pakistan. These preliminary findings will pave future way for phytochemists, biotechnologists, herbalists and pharmaceutical industries for exploring new drugs for combating various diseases.

## **Materials and Methods**

Healthy shoots/leaves of 24 alpine plant species were collected at three phenological stages, *i.e.* pre-reproductive stage (young vegetative stage), reproductive stage (flowering/fruiting stage) and post-reproductive stage (fruit/seed maturation and dispersal stage) during summer from Mastuj Valley. The plant material was shade dried for 21 days and powdered. These were qualitatively screened for the presence of alkaloids, saponins, tannins and oils following the standard methods (Banu & Cathrine, 2015; Balamurugan *et al.*, 2019; Trease & Evans, 2009). There were, 5 tree, 8 shrub, 7 herbaceous and 4 grassy species of 16 families. For alkaloids, 2 drops of Mayer's reagent was added to extract; a creamy or white precipitate indicated the presence of alkaloids. It

was confirmed by adding 2ml HCl to 5 ml extract, followed by 1 ml of Dragendroff's reagent. The appearance of orange or red precipitate indicated the presence of alkaloids. Saponins were detected by vigorous shaking of 5 ml of extract with few ml distilled water. The formation of froth indicated saponins. The resultant froth with few drops of olive oil was vigorously shaken that formed emulsion, which further confirmed the presence of saponins. Tannins were deducted by adding few drops of neutral ferric chloride solution to 5 ml of extract. The appearance of dark green colour showed the presence of tannins. The presence of oils was checked by spot test. Small amount of extract was pressed between 2 filter papers. The appearance of spot showed the presence of oils. Further testing was done by saponification. Few drops of 0.5N alcoholic KOH and few drops of phenolphthalein were added and heated for 2 hours. The formation of soap or partial neutralization of alkali indicated the presence of fixed oils or fats.

## **Results and Discussion**

The qualitative screening of four secondary metabolites in 24 medicinally important alpine plants of Mastuj Valley indicated that alkaloids and saponins were present in 21 species. Tannins and oils were detected in 20 species. Fifteen species contained all the 4 phytochemicals, 5 species had 3 phytochemicals and 3 species had 2 phytochemicals. Ibrahim et al., (2021) observed that all the tested plants had varied amounts of alkaloids, saponins and tannins. In the present case 24 species showed variations in types of phytochemicals among different species and at various phenological stages. The results agree with Lydia et al., (2022), who reported that tannins were present in 50 plant species, alkaloids in 43 species, glycosides in 33 species and saponins in 8 species. Based on the phenological stages alkaloids were respectively recorded in 12, 8 and 20 species at pre-reproductive, reproductive and post reproductive stages. Saponins were accordingly present in 21, 19 and 16 species in pre-reproductive, reproductive and post reproductive stages. The prereproductive, reproductive and post-reproductive stages sequentially had tannins in 18, 15 and 16 species. Oils were recorded in 14, 15 and 18 species in the three successive phenological stages. Based on the age and plant part, leaves of Cola millenii had none of the tested secondary compounds including tannins, saponins, steroids and alkaloids etc (Onefeli et al., 2021), but its fruit had the highest flavonoid contents. Verdecia et al., (2021) also observed that primary compounds decreased with the advancing age; while secondary compounds enhanced with maturity. This also supported the present findings as the presence/absence of various phytochemicals depended on phenological stage. Feudjio et al. (2020) also reported that phenology affected quantitative and qualitative values of phytochemicals in plants. They further stated a decrease of phytochemicals during flowering and fruiting stage. Adegbaju et al., (2020) is also of the view that developmental stages influence the physiology and chemistry of plants. Phenological stage is the growth

stage that varies with season. The Pre-reproductive stage is the young seedling stage in annuals and pre-flowering stage in perennial plants. Reproductive stage is the flowering and fruiting stage in annuals and perennial species; while Post-reproductive phase represents postflowering, fruiting and dispersal of seeds and fruits period. It is the completion of life cycle in annual plants; while in perennial species it represents dormant stage. These stages are correlated to the seasonal changes in each locality under a set of climatic conditions. Keeping this view in mind, individual phytochemicals are discussed below.

1. Alkaloids: Alkaloids are important in traditional human healthcare system due to their antibacterial, antifungal, anti-parasite, antimalarial, analgesic, antifibrillation and anti-tumor activities (Fazili et al., 2022; Twaij et al., 2022). Alkaloids were present in 21 species, which were distributed respectively in 12, 8 and 20 species at pre-reproductive, reproductive and post reproductive stages. The present study highlights the presence of alkaloids in Betula utilis at all the three phonological stages (Table 1). It was present at reproductive stage in Tamarix sp. and at postreproductive stage in Hippophae rhamnoides. It was not detected at pre-reproductive stage in Elaeagnus angustifolia and Juniperus excelsa. Among the shrubs (Table 2), alkaloids were present at all the phenological Acantholimon longiscapum, stages in Ephedra gerardiana, Sophora mollis and Berberis lyceum. However, it was completely absent in Capparis spinosa. It remained below the deductible level in Myrtama elegans and Rosa webbiana at pre-reproductive stages. Datura stramonium, Glycyrrhiza glabra, Mentha

longifolia, Rheum emodi and Salvia nubicola among the herbs possessed alkaloids at all the three phenological stages (Table 3). Alkaloids were also noticed in the postreproductive stages of Nepeta cataria and Verbascum thapsus. Among the grasses (Table 4), alkaloids were present in Cynodon dactylon and Phragmites karka at all the three phenological stages; but it tested negative in Calamogrostris pseudophragmites and Saccharum spontaneum at the three phenological stages. Alkaloids were detected in 21 species. Of them, it was recorded 12, 18 and 20 times respectively at pre-reproductive, reproductive and post-reproductive stages. Our results agreed with many workers who reported the presence of alkaloids along with other phytochemicals that differed from species to species and collection site (Pereira et al., 2022), and season (deSouza et al., 2022; Gomes et al., 2022; Fazili et al., 2022). The present findings also agree with those of Sharifi-Rad et al., (2022), who stated that alkaloid contents of Teucrium polium differed in shoots and roots at various phenological stages. The maximum alkaloids contents were achieved at flowering stage. Mohan et al., (2021) also spotted alkaloids in all the tested medicinal plants, which agreed with the present findings. Similarly, Mehmood et al., (2021) also observed the presence of alkaloids in Euphorbia helioscopia and Rumex dentatus. Marami et al., (2021) concluded that all the analyzed phytochemicals were present in all the plants tested by them. This also strengthened the present findings as we recorded the presence of four tested phytochemicals in 21 investigated species. Many available studies indicate the presence of alkaloids in various plants or their parts (Gwari et al., 2021; Bashir & Kumar, 2021; Shukla et al., 2021) that are in line with the present study.

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Plant species	Phenological Stage		Alkaloids	Saponins	Tannins	Oils
1. Betula utilis D. Don	•		•			
	i.	Pre-reproductive stage	+	+	-	-
	ii.	Reproductive stage	+	+	+	+
	iii.	Post-reproductive stage	+	+	+	+
2. Elaeagnus angustifolia L.						
	i.	Pre-reproductive stage	-	+	+	-
	ii.	Reproductive stage	+	+	+	+
	iii.	Post-reproductive stage	+	+	+	+
3. Hippophae rhamnoides L.						
	i.	Pre-reproductive stage	-	+	+	+
	ii.	Reproductive stage	-	+	+	+
	iii.	Post-reproductive stage	+	-	+	+
4. Juniperus excelsa M. Bieb.						
	i.	Pre-reproductive stage	-	+	+	+
	ii.	Reproductive stage	+	+	+	+
	iii.	Post-reproductive stage	+	+	+	+
5. Tamarix sp.						
	iv.	Pre-reproductive stage	-	+	+	+
	v.	Reproductive stage	+	+	-	-
	vi.	Post-reproductive stage	-	+	-	+

Table 1. Some phytochemicals in alpine medicinal tree species of Mastuj Valley, Hindu Kush Range, Pakistan.

Plant species Phenological Stage Alkaloids Saponins Tannins								
Plant species		Phenological Stage	Alkaloids	Saponins	Tannins	Oils		
1. Acantholimon longiscapum Bokhari								
	i.	Pre-reproductive stage	+	+	-	-		
	ii.	Reproductive stage	+	+	-	-		
	iii.	Post-reproductive stage	+	+	+	+		
2. Berberis lyceum Royle								
	i.	Pre-reproductive stage	+	-	+	+		
	ii.	Reproductive stage	+	-	+	+		
	iii.	Post-reproductive stage	+	-	+	+		
3. Capparis spinosa L								
	i.	Pre-reproductive stage	-	+	+	-		
	ii.	Reproductive stage	-	+	+	-		
	iii.	Post-reproductive stage	-	+	+	-		
4. Ephedra gerardiana L.		1 0						
I	i.	Pre-reproductive stage	+	+	_	+		
	ii.	Reproductive stage	+	+	+	+		
	iii.	Post-reproductive stage	+	+	+	+		
5. Myrtama elegans (Royle) Ovcz. & K		i ost reproductive stage						
	i.	Pre-reproductive stage	-	+	+	+		
	ii.	Reproductive stage	+	+	+	_		
	iii.	Post-reproductive stage	+	_	+	+		
6. <i>Rosa webbiana</i> Wall ex Royle		i ost reproductive stage						
0. Rosa webbiana wan ex Royie	i.	Pre-reproductive stage	-	+	+			
	ii.	Reproductive stage	+	+	+	-		
						-		
7. C	iii.	Post-reproductive stage	+	+	+	+		
7. Sophora mollis (Royle) Baker		<b>D</b>						
	i.	Pre-reproductive stage	+	+	+	+		
	ii.	Reproductive stage	+	+	+	+		
	iii.	Post-reproductive stage	+	+	+	+		
8. Artemisia parviflora Roxb.								
	i.	Pre-reproductive stage	-	+	+	-		
	ii.	Reproductive stage	+	-	-	-		
	iii.	Post-reproductive stage	+	-	-	-		

# Table 2. Some phytochemicals in alpine medicinal shrubby species of Mastuj Valley,Hindu Kush Range, Pakistan.

2. Saponins: Saponins are therapeutically important due to their hypolipidemic and anticancer activities. Cardiac glycosides and steroidal saponins are used commercially for the production of sex hormones. Saponins, present in 21 species, were accordingly recorded in 21, 19 and 16 species in pre-reproductive, reproductive and post reproductive stages. Saponins were present at all the phenological stages in the tree species (Table 1). Saponins were also detected in Hippophae rhamnoides at pre- and reproductive stages. Among the shrubs (Table 2), saponins were present at all the three phenological stages in Acantholimon longiscapum, Capparis spinosa, Ephedra gerardiana, Rosa webbiana and Sophora mollis. Berberis lyceum tested negative for saponins at all the three phenological stages. Myrtama elegans was positive for saponins at pre-reproductive and productive stages. Saponins were recorded in Artemisia parviflora at prereproductive stage. Among the herbs (Table 3), saponin was present in the three phenological stages in Datura stramonium, Glycrhiza glabra, Mentha longifolia, Rheum emodi and Verbascum thapsus. Salvia nubicola had saponins at pre- and reproductive stages. Saponins were noticed in Nepeta cateria only at pre-reproductive stages. absent Saponins were in Calamogrostris and pseudophragmites Phragmiteskarkaat all the phenological stages; but on the contrary, Cynodon dactylon and Saccharum spontaneum were positive for saponins at all the three growth stages (Table 4). Akinmoladun et al., (2022) reported the absence of saponins in Chrysophyllum albidum, which supports the present findings. Sharifi-Rad et al., (2022) recorded the highest and lowest saponin contents in the aerial parts of Teucrium polium respectively at flowering and at immature stages. Similar to the present findings, Mehmood et al., (2021) also observed tannins in Euphorbia helioscopia. Marami et al., (2021) concluded that saponins were present in all the plants analyzed by them. In our case, 87.5% plants contained saponins. Many recent studies support the presence of saponins in various plants (Gwari et al., 2021; Bashir & Kumar, 2021; Mansoori et al., 2020; Elaigwu et al., 2020; Sulaiman et al., 2020) that are in line with the present findings.

Plant species		Phenological Stage	Alkaloids	Saponins	Tannins	Oils	
1. Datura stramonium L.							
	i.	Pre-reproductive stage	+	+	+	+	
	ii.	Reproductive stage	+	+	+	+	
	iii.	Post-reproductive stage	+	+	+	+	
2. Glycyrrhiza glabra L.							
	i.	Pre-reproductive stage	+	+	+	-	
	ii.	Reproductive stage	+	+	+	-	
	iii.	Post-reproductive stage	+	+	+	-	
3. Mentha longifolia (L.) Huds.							
	i.	Pre-reproductive stage	+	+	+	-	
	ii.	Reproductive stage	+	+	+	+	
	iii.	Post-reproductive stage.	+	+	+	+	
4. Nepeta cataria L.							
	i.	Pre-reproductive stage	-	+	+	+	
	ii.	Reproductive stage	-	-	+	+	
	iii.	Post-reproductive stage	+	-	+	+	
5. Rheum emodi Wall ex Meissn.							
	i.	Pre-reproductive stage	+	+	+	-	
	ii.	Reproductive stage	+	+	-	-	
	iii.	Post-reproductive stage	+	+	-	-	
6. Salvia nubicola Wall ex Sweet							
	i.	Pre-reproductive stage	+	+	+	-	
	ii.	Reproductive stage	+	+	+	+	
	iii.	Post-reproductive stage	+	-	+	+	
7. Verbascum thapsus L.							
	i.	Pre-reproductive stage	-	+	+	+	
	ii.	Reproductive stage	-	+	-	-	
	iii.	Post-reproductive stage	+	+	-	-	

## Table 3. Some phytochemicals in alpine medicinal herbaceous species of Mastuj Valley, Hindu Kush Range, Pakistan.

## Table 4. Some phytochemicals in alpine grasses of Mastuj Valley, Hindu Kush Range, Pakistan.

Plant species		Phenological Stage	Alkaloids	Saponins	Tannins	Oils
1. Calamogrostris pseudophragmites H	all. f. Ko	eler				
	i.	Pre-reproductive stage	-	-	-	+
	ii.	Reproductive stage	-	-	-	+
	iii.	Post-reproductive stage	-	-	-	+
2. Cynodon dactylon (L.) Pers.						
	i.	Pre-reproductive stage	+	+	-	+
	ii.	Reproductive stage	+	+	-	+
	iii.	Post-reproductive stage	+	+	-	+
3. Phragmites karka (Retz.) Trin ex Ste	eud.					
	i.	Pre-reproductive stage	+	-	-	+
	ii.	Reproductive stage	+	-	-	+
	iii.	Post-reproductive stage	+	-	-	+
4. Saccharum spontaneum L.						
	i.	Pre-reproductive stage	-	+	-	+
	ii.	Reproductive stage	-	+	-	+
	iii.	Post-reproductive stage	-	+	-	+

3. Tannins: Tannins are water and alcohol soluble phenolic compounds of high molecular weight, with common distribution in various parts of plants. Tannins were detected in 20 species. The pre-reproductive, reproductive and post reproductive stages sequentially had tannins in 18, 15 and 16 species. The present study reported tannins at three phenological stages of tree species (Table 1) including Betula utilis, Elaeagnus angustifolia, Hipophae rhamnoides and Juniperus excelsa Tannins were absent in Tamarix sp. at reproductive and post-reproductive stages. Amongst shrubs (Table 2), Berberis lyceum, Capparis spinosa, Myrtama elegans, Rosa webbiana and Sophora mollis contained tannins at all the three phenological stages. It was present at post-reproductive stage in Acantholimon longiscapum; and at reproductive and post-reproductive stages in Ephedera gerardiana. Artemisia parviflora contained tannins at pre-reproductive phase. Among the herbs (Table 3), tannins were recorded in Datura stramonium, Glycyrrhiza glabra, Mentha longifolia, Nepeta cataria and Salvia nubicola at all the phenological stages. Whereas, Rheum emodi and Verbascum thapsus had tannins at pre-reproductive stages only. Tannins were completely absent in all the 4 tested grasses at all the growth stages (Table 4). Like the present study, tannins were present in Montrichardia (Pereira et al., 2022) and Chrysophyllum albidum (Akinmoladum et al., 2022) that varied in concentration due to locality. The findings of Aljaiyash et al., (2022) are in line with the present findings as they also stated that phytochemicals present in Thymus marocannus increased from pre-flowering to post-flowering stages. Similarly, Yu et al., (2021) also achieved highest tannin contents in pomegranate among various food and fruit plants. They recorded that tannins varied from absent to various amounts among the tested alpine plants at various phenological stages. This also strengthened our findings regarding the role of phenology in the absence/presence of a particular secondary chemical. Various studies also reported tannins in various plants (Gwari et al., 2021; Nagano & Batalini, 2021; Mehmood et al., 2021; Bashir & Kumar, 2021; Mansoori et al., 2020; Farag et al., 2020; Elaigwa et al., 2020), which support the present findings.

4. Oils: Oils were detected in 20 species, which successively appeared in 14, 15 and 18 species in prereproductive, reproductive and post-reproductive phenological stages. Amongst the tree species (Table 1), oil was detected in Hipophae rhamnoides and Juniperus excelsa at all the three phenological stages. It was also present at reproductive and postreproductive stages in Betula utilis and Elaeagnus angustifolia, While, Tamarix sp. contained oils in preand post-reproductive stages. Within the shrubs (Table 2), oil was present at all the three phenological stages in Berberis lyceum, Ephedera gerardiana and Sophora mollis; but oil was absent at all the three growth stages in Artemisia. Oil was detected at post-reproductive stage in Acantholimon longiscapum and Rosa webbiana and at pre- and post-reproductive stages in Myrtama elegans. Among the herbs (Table 3), oils

were spotted at all the three phenological stages of Datura stramonium, and Nepeta cataria. On the other hand, Glycyrrhiza glabra and Rheum emodi tested negative at all the three phenological stages for oil contents. Mentha longifolia and Salvia nubicola had oils at reproductive and post-reproductive stages. Verbascum thapsus had oil only at the pre-reproductive stage. All the three phenological stages of the four analyzed grasses (Table 4) contained oils. Aljaiyash et al., (2022) observed that oil contents increased from pre-flowering to flowering stage; but declined in the post-flowering stage. Mollaei et al., (2020) demonstrated that the maximum yield of Zosima oil was achieved at the late-mature/ripe seeds. The findings also agree with those of Reaisi et al., (2019), who achieved highest yield of oil at seed filling stage and demonstrated a correlation between altitude, phenology and oil contents and their composition.

## Conclusions

Plants exhibit phytochemical diversity due to climatic, edaphic and phenologic variation. Same was true in the present study. Phytochemical screening is desirable for the discovery of therapeutic agents and new sources of medicinal material. Various secondary compounds including tannins, oils, gums, flavonoids, saponins and essential oils are medicinally important that act as precursors for the synthesis of complex chemical substances. The chemical composition of plants varies according with growth stage due to physiological demands. The pre-reproductive phase is vegetative seedling stage, which is sequentially followed by reproductive (flowering & fruiting) and postreproductive (maturation of fruits and their dispersal) stages. The annual plants complete their life cycle after shedding their seeds and fruits; while for perennial species this stage means advancing towards dormant stage. The results may help in finding the suitable stage for harvesting of medicinal plant for a particular chemical. The findings can also be helpful in regulating the time of grazing as Mastuj Valley is a high altitude grazing land. Qualitative screening sometimes fails to mark the presence or absence of particular phytochemical due undetectable trace amount of material. We used one solvent and collected plants from one locality due to time, physical and financial limitations. This preliminary qualitative effort, therefore, can be advanced further towards quantitative analysis using more plant species from different locations, seasons and extraction with various solvents of different polarity to exactly quantify the phytochemicals at different growth stages in different parts of the plant.

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