

AN INVESTIGATION OF PHYLOGENETIC RELATIONSHIPS OF POPULATIONS OF *ARTEMISIA TAURICA* Willd. (ASTERACEAE) FROM TURKEY USING cpDNA “*trn*” REGIONS

PELIN YILMAZ SANCAR^{1*}, SUEDA DELIPOYRAZ¹, SEMSETTIN CIVELEK¹ AND MURAT KURSAT²

¹Firat University, Faculty of Sciences, Department of Biology, 23119 Elazığ, Turkey

²Bitlis Eren University, Faculty of Arts and Sciences, Department of Biology, 13000 Bitlis, Turkey

*Corresponding author's email: peyilmaz@firat.edu.tr

Abstract

Artemisia L. is one of the biggest genera in the family Asteraceae, with around 500-600 taxa at specific and sub-specific levels divided in five subgenera. The majority of the members of this genus have a high economic value. Due to the high number of taxa, lot of taxonomists are tried to solve the problem of its classification and phylogeny but its natural classification still has not been achieved.

In order to find the phylogenetic relations and kinships between individual samples, the present study investigated the base polymorphisms in sequences of non - coding *trnT-trnL3'* section of chloroplast DNA (cpDNA) of 54 individual samples of 22 different populations belonging to *A. taurica* Willd. in Turkey.

DNA was isolated from the leaves of the studied individual samples by the CTAB method, and isolated genomic DNA was multiplied in PCR by using *trna* and *trnd* primaries of the *trnT-trnL3'* section. The obtained data were evaluated by the Mega X program, and phylogenetic trees were prepared by using the “Maximum Likelihood” method.

There are no molecular significant differences between populations of the species *A. taurica* in Turkey depending on absence of genetic isolation and still ongoing gene. The view that the morphological differences observed on the samples of the Zerne dam lake (Van) population of the species *A. taurica* were found to be related to the ecological conditions and ploidy level.

Key words: Asteraceae, *Artemisia taurica*, cpDNA *trnT-trnL3'*, Phylogeny, PCR.

Introduction

Artemisia L. is a member of the Asteraceae family, which is a polymorphic genus and is very significant from a therapeutic and economic point of view. This genus is mostly distributed in the temperate sectors of northern hemisphere. A limited number of *Artemisia* species also occur in the southern hemisphere of the globe (Oberprieler *et al.*, 2009). This genus contains five hundred plants including both shrubs and herbs (Valles & McArthur, 2001) and is designated as a prevalent and diverse genus of the Anthemideae community from Asteraceae (Martin *et al.*, 2003).

A great number of the *Artemisia* species possess economic importance because they are utilized as fodder, therapeutics, aesthetics, and soil binder, whilst a few taxa have been shown to cause allergy and few are just noxious weeds damaging number of crops because of their poisonous properties (Hayat *et al.*, 2009a; Hussain *et al.*, 2019).

Twenty two species of *Artemisia* without any infraspecific taxa are reported in the 5th volume of the Flora of Turkey (Cullen, 1975; Davis, 1975; Davis *et al.*, 1988). Later, *Artemisia verlotiorum* Lamotte was added to the 10th volume of the Flora of Turkey as a new record for Turkey, so the number of the species in *Artemisia* in Turkey grew to 23 (Davis *et al.*, 1988).

The genus *Artemisia* is divided into 4 subgenera as subgenus *Artemisia* Lessing, *Dracunculus* (Besser) Rydberg, *Seriphidium* Besser ex Lessing, and *Tridentatae* (Rydberg) McArthur (McArthur *et al.*, 1981; Civelek *et al.*, 2010; Kursat, 2010; Kursat *et al.*, 2015).

The general distribution areas of the species *Artemisia taurica* worldwide are Europe, Southern Russia (Caucasus), Crimea and Turkey. *Artemisia taurica* is widely distributed in the steppes of Central, Eastern, and Southeastern Anatolia in Turkey. It is one of the four species that have a wide distribution in Turkey. The other three species with a wide distribution in Turkey are *Artemisia absinthium* L., *Artemisia vulgaris* L., and *Artemisia campestris* L. (Civelek *et al.*, 2010; Kursat, 2010).

Civelek *et al.*, (2010) revised the genus *Artemisia* in Turkey. During this study of the genus *Artemisia*, significant morphological differences were observed in the of *Artemisia taurica*, especially in the eastern Anatolia (Civelek *et al.*, 2010; Kursat, 2010; Ayaz *et al.*, 2019). A molecular study was needed to determine if these differences had some genetic basis or just morphological polymorphisms. For this reason, the purpose of this study was to evaluate the genetic diversity and taxonomic position of the species *Artemisia taurica* in Turkey using some molecular techniques. No molecular systematic studies were conducted using the populations of *Artemisia* genus of Turkey. This study is important in that it is the first molecular-based study using *Artemisia taurica* taxa of Turkey. In addition, the first molecular data for the *trnT-trnL3'* gene region from Turkey were submitted to the international NCBI GenBank databases.

Material and Methods

Plant material: A total of 54 different individuals belonging to the taxa of *Artemisia taurica* species were gathered M. Kursat from the populations from the natural

habitat in 22 different regions in Turkey. These plants were kept in the Herbarium belonging to the Faculty of Sciences at Bitlis Eren University. The collection was made during the vegetative and flowering periods in the months of April-September in years of 2007-2010. The locations of the collections areas were given in (Table 1) and (Fig. 1).

DNA isolation: The leaf tissue of the samples were used for DNA isolation which was carried out manually using the CTAB (Cetyl Trimethyl Ammonium Bromide) method (Doyle & Doyle, 1987). By measuring the concentrations of isolated DNAs using a nanodrop spectrophotometer, they were adjusted to 25 ng/ul. Stock DNA was preserved at -20 C.

PCR amplification: In the PCR studies conducted using *trnT* and *trnL* primers, the *trnT* - *trnL3'* region multiplied with the 54 samples had a nucleotide length of 750-800. The sequence of primers used to amplify the *trnT* - *trnL3'* region is given in Table 2 (Taberlet *et al.*, 2007). Aiming for the final concentration for the PCR studies to be 50 μ L; 10 μ L buffer, 3 μ L MgCl₂, 1 μ L dNTPs, 0,50 μ L from each primer (forward and reverse), 0,50 μ L taq polymerase, and nearly 6 ng (2,70 μ L) template DNA were mixed. Amplifications were carried out in a Biorad T100 Gradient Thermal Cycler. The PCR device was repeated for 34X cycles, consisting of 2 minutes at 95°C initial denaturation, 1 minute at 95°C denaturation, 40 seconds at 57°C annealing, 1 minute at 72°C extension, and 5 minutes at 72°C final extension. The PCR products were monitored in agarose gel containing 2% ethidium bromide (0.5 μ g/ml) and viewed with the UV transilluminator.

Sequence analysis: Two-way reading was applied to the amplification products. The PCR purification process was carried out before the sequence analysis. The purification

and sequencing processes were done by the Macrogen Company. Finch TV Version 1.4 was used to evaluate the data from the chromatogram. In order to specify the phylogenetic relations among *Artemisia taurica* taxa that grew in different geographical regions, Mega program was used and the molecular variability parameters among the taxa were determined. The sequence alignments were recorded in the NCBI data bank and accession numbers were obtained (Table 3). The DNA sequence alignments for 55 individuals (one of them being outgroup) were evaluated using the Mega Program X Version. The DNA sequence alignments for all the individuals were subjected to statistical analysis.

Results

In this research, 55 individuals taken from 22 different populations of *A. taurica* were examined. All the examined individuals from the same and different populations of *Artemisia taurica*, were used to determine the sequences of “*trn*” regions. The peak results of the bi-directional sequences sent to us from MacroGene were evaluated using Version 1.4 of the Finch TV program. Using the “Multiple Alignment Blast System” of the automatic sequencing systems, the sequences were aligned. The noticeable differences were manually corrected.

As a result of the scans performed in the NCBI (National Center for Biotechnology Information) database for *Artemisia taurica*, no reference regions were obtained. For a more accurate visualization of the results of the alignment, we did not evaluate about 50-100 bases from the beginning and the end. For this reason, approximately 779 base pairs for the cpDNA *trn* regions were used. In the phylogenetic tree drawing, the DNA sequences of the *trnT-trnL3'* region in the chloroplast genome were evaluated using Version X of the Mega program.

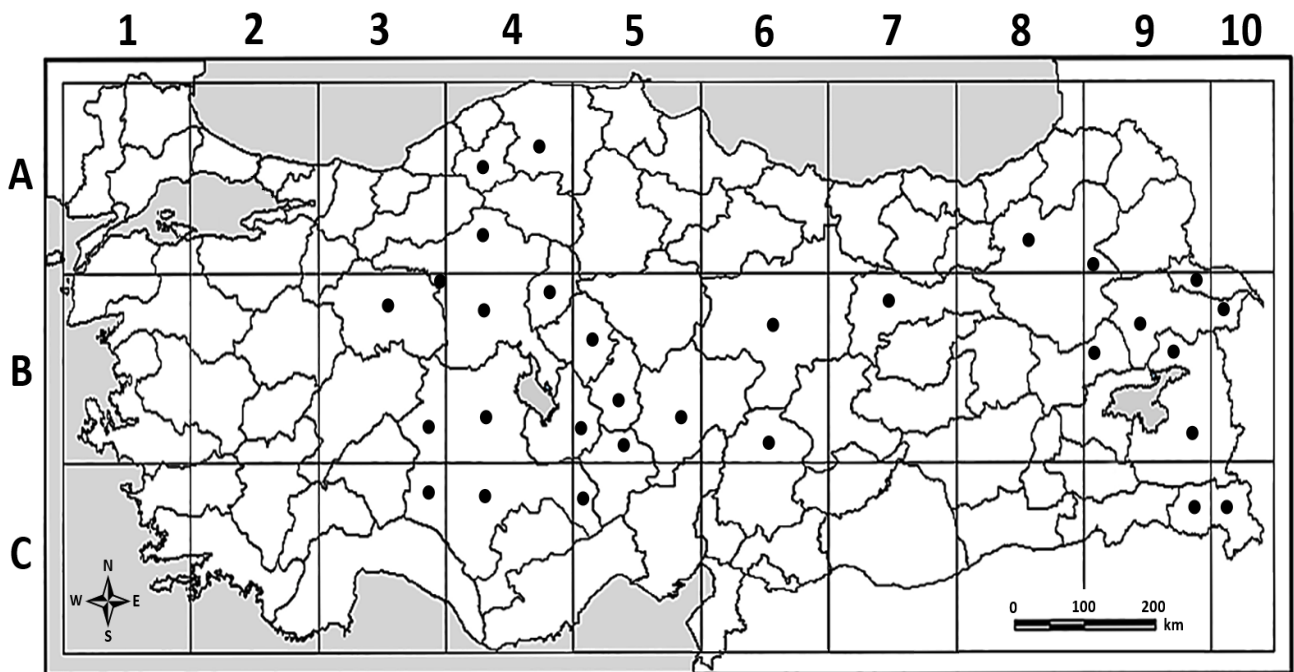


Fig. 1. Geographic distribution of species *A. taurica* (●) in Turkey (Civelek *et al.*, 2010; Kursat, 2010).

Table 1. Information of *Artemisia taurica* location in field.

Taxa of the <i>Artemisia taurica</i>	Number of populations and collector	Collection date	Detailed localities of specimens populations	Lat- Lon
<i>A. taurica</i> 1 (4 individual)	1007 M. KURŞAT	06.07.2007	B5 Niğde: Kayseri highway, after 5 km, the area between the railway and the highway, 1250-1300m.	38.15 N 34.17 E
<i>A. taurica</i> 2 (3 individual)	1033 M. KURŞAT	10.09.2007	B4 Ankara: Between Lalahan and Elmadağ, around garrison and slopes, field edges, 1225 m	39.22 N 31.01 E
<i>A. taurica</i> 3 (4 individual)	1031 M. KURŞAT	06.12.2007	B4 Ankara: Polatlı highway, 10 km before Temelli, roadsides, 843 m	40.43 N 32.20 E
<i>A. taurica</i> 4 (2 individual)	1098 M. KURŞAT	23.10.2007	B3 Eskişehir: Polatlı - Sivrihisar, 35 km before Sivrihisar, roadsides, stepper, 837 m.	40.33 N 29.31 E
<i>A. taurica</i> 5 (1 individual)	1114 M. KURŞAT	26.11.2007	B10 Ağrı: Between Habur-Tutak, 4 km after Habur, roadsides, stepper, 1605 m.	38.31 N 30.32 E
<i>A. taurica</i> 6 (3 individual)	1032 M. KURŞAT	10.09.2007	B4 Ankara: Between Gölbaşı and Bayındır Dam, stepper 994 m.	41.16 N 27.59 E
<i>A. taurica</i> 7 (3 individual)	1034 M. KURŞAT	10.09.2007	B4 Ankara: Between Elmadag and Kirikkale, 1000 m.	38.08 N 30.56 E
<i>A. taurica</i> 8 (2 individual)	1035 M. KURŞAT	11.09.2007	B5 Kırşehir: Kaman - Hirfanlı dam road, roadsides, 10 km after Kaman, 975 m.	40.31 N 29.32 E
<i>A. taurica</i> 9 (3 individual)	1011 M. KURŞAT	07.07.2007	B4 Ankara: Şereflikoçhisar, South slopes of Akinler village, stepper, 989 m.	38.27 N 30.16 E
<i>A. taurica</i> (var. nova) 10 (3 individual)	1056 M. KURŞAT	20.09.2007	B9 Van: Van - Hakkari highway, Northern slopes of Zerneke dam, roadside, mountain steppe, 1960m.	41.37 N 26.38 E
<i>A. taurica</i> 11 (2 individual)	1185 M. KURŞAT	02.11.2008	B9 Van: Erçiş, Zernaki Mountain, Irşat region, slopes, steppe, 1712 m.	39.03 N 43.20 E
<i>A. taurica</i> 12 (1 individual)	1119 M. KURŞAT	26.11.2007	B8 Muş: Malazgirt, between Aktuzla-Karıncalı villages, 3 km before Karıncalı village, slopes, 1550 m.	39.21 N 42.15 E
<i>A. taurica</i> 13 (3 individual)	1071 M. KURŞAT	23.09.2007	B9 Van: Highway between Erçiş-Adilcevaz, 21. km, roadsides, slopes, 1750m.	38.57 N 43.13 E
<i>A. taurica</i> 14 (1 individual)	1089 M. KURŞAT	22.10.2007	B5 Kayseri: Between Kayseri and Avanos, 39 km after Kayseri, roadsides, slopes, steppes, 1121 m	38.43 N 35.04 E
<i>A. taurica</i> 15 (1 individual)	1036 M. KURŞAT	11.09.2007	B5 Kırşehir: Between Hirfanlı Dam and Şereflikoçhisar, from Hirfanlı Dam to the West, hills and roadsides, 972m.	39.16 N 33.30 E
<i>A. taurica</i> 16 (4 individual)	1028 M. KURŞAT	10.09.2007	B4 Ankara: Polatlı highway, 37 km before Polatlı, roadsides, 843m.	39.45 N 32.28 E
<i>A. taurica</i> 17 (3 individual)	1005 M. KURŞAT	04.07.2007	B6 Kahramanmaraş: Göksun, Upper parts of Fındıklıkoyak village, forest area, summit, open area, 640 m.	37.60 N 36.32 E
<i>A. taurica</i> 18 (3 individual)	1090 M. KURŞAT	22.10.2007	B6 Kahramanmaraş: Göksun, Fındıklıkoyak village entrance and surroundings, 1450 m	37.60 N 36.32 E
<i>A. taurica</i> 19 (1 individual)	1097 M. KURŞAT	23.10.2007	B4 Ankara: Polatlı highway, roadsides, 843m.	39.43 N 32.23 E
<i>A. taurica</i> 20 (2 individual)	1009 M. KURŞAT	07.07.2007	B4 Ankara: Şereflikoçhisar, Hamzalı village, Kayacık (Mutlucan) salt factory area, 933 m.	38.50 N 33.26 E
<i>A. taurica</i> 21 (3 individual)	1172 M. KURŞAT	26.08.2008	B10 Ağrı: Doğubeyazıt, Bottom of İshakpaşa Palace, 1935 m	39.31 N 44.07 E
<i>A. taurica</i> 22 (2 individual)	1058 M. KURŞAT	20.09.2007	C10 Hakkari: Hakkari highway, 37 km before Hakkari, roadsides, slopes, steppes, 1496 m.	37.41 N 43.58 E

The nucleotide composition and other features of the individuals were determined as a result of the statistical analyses performed by cutting the excess parts at the head and end of the DNA sequences. In the DNA sequences of the *trnT-trnL3'* region in the chloroplast genome of the examined individuals, some of the parameters of the molecular diversity such as the conserved regions (C), variation regions (V), parsimony informative regions (Pi), single parts (S), homologous base pairs (ii), transitional base pairs (si), transversional base pairs (sv), and R-value (si / sv) were calculated, and the obtained values were given in (Table 4).

Using the Best DNA / Protein step in the Models menu of the MEGA program, the methods that best expressed the phylogenetic relationship between the individuals were determined.

In the list of the methods given, the lowest value of the BIC (Bayesian Information Criterion) was found in the T92 + G (Tamura-3-parameter) method (Kumar *et al.*, 2019). The Maximum Likelihood, Neighbor-Joining, UPGMA, and Maximum Parsimony methods were applied separately. The method that illustrated best the evolutionary and phylogenetic relationship between the examined individuals was the Maximum Likelihood method.

Many of the phylogenetic trees were drawn by trying many methods, and the most useful tree was chosen. In the Maximum Likelihood method, by entering the bootstrap value as 100, a single phylogenetic tree for a total of 55 individuals, 54 of which were examined and one of which was an outgroup, was obtained (Fig. 2). The species *Haplocarpa scaposa* L. as an outgroup was used (Sancar, 2017, Kursat *et al.*, 2018, Sancar *et al.*, 2019).

Table 2. The base sequences of the primers used.

Primers	Base sequences (5' – 3')
<i>trna</i> (Forward):	5' CAT TAC AAA TGC GAT GCT CT 3'
<i>trnd</i> (Reverse):	5' GGG GAT AGA GGA CTT GAA C 3'

Table 3. Genbank accession numbers for the *trnT-trnL3'* regions of the studied samples.

Specimens	GenBank Accession Numbers
<i>Artemisia taurica</i> 1	MT637781
<i>Artemisia taurica</i> 2	MT637782
<i>Artemisia taurica</i> 3	MT637783
<i>Artemisia taurica</i> 4	MT637784
<i>Artemisia taurica</i> 5	MT637785
<i>Artemisia taurica</i> 6	MT637786
<i>Artemisia taurica</i> 7	MT637787
<i>Artemisia taurica</i> 8	MT637788
<i>Artemisia taurica</i> 9	MT637789
<i>Artemisia taurica</i> 10	MT637790
<i>Artemisia taurica</i> 11	MT637791
<i>Artemisia taurica</i> 12	MT637792
<i>Artemisia taurica</i> 13	MT637793
<i>Artemisia taurica</i> 14	MT637794
<i>Artemisia taurica</i> 15	MT637795
<i>Artemisia taurica</i> 16	MT637796
<i>Artemisia taurica</i> 17	MT637797
<i>Artemisia taurica</i> 18	MT637798
<i>Artemisia taurica</i> 19	MT637799
<i>Artemisia taurica</i> 20	MT637800
<i>Artemisia taurica</i> 21	MT637801
<i>Artemisia taurica</i> 22	MT637802

Table 4. PCR amplified summary statistics of the cpDNA *trn* region of the genus *A. taurica*.

Parameters of molecular diversity	<i>trnT - trnL3'</i> region
Total population number	22
Total individuals	54
Total band Length	779
The ratio of G-C base pair (%)	35,5
Conserved regions (C)	719
Variation regions (V)	37
Single parts (S)	32
Parsimony informative regions (Pi)	5
Homologous base pairs (ii)	752
Transitional base pairs (si)	1.00
Transversional base pairs (sv)	1.00
R value (si/sv)	1.00

Discussion

In this research, a phylogenetic systematic study was conducted by using the molecular data of *A. taurica*. 54 individuals taken from 22 different populations belonging to this species were examined by analyzing the base slice of the regions. An attempt to find some information about the populations' closeness with and distance from each other. This research is important as it is the first molecular-based study of the species *A. taurica* growing naturally in Turkey.

Molecular diversity parameters for the individuals of the *A. taurica* species are shown in Table 4. As seen in the table, the number of informative regions is very low, which indicates that the sequences between populations do not differ much. However, when the positions of the variable bases were examined in terms of the variation we obtained in our studies, it was seen that the individuals of the *A. taurica* species in both close and same localities generally showed similar base sequences and were placed sideways in the phylogenetic tree in Figure 2.

It was reported that the apomictic reproduction pattern was very common in the autopolyploid species and populations of the genus *Artemisia* (Czapik, 1996; Noyes, 2007; Melanie *et al.*, 2014). According to Gustavsson (1947), the apomictic breeding pattern guarantees individual genotypes a long-lasting existence in large areas (Gustavsson, 1947). Apomictic reproduction may cause high similarities and fewer variations at the molecular level between samples of different populations studied. This is because apomictic seed development is a form of reproduction that generates offspring genetically identical to the parent plant. In this study, considering the high numerical values of the conserved region and homologous base pair and the low numerical values of regions with variation and informative regions; we can say that the studied populations of the *A. taurica* species have a high probability of apomictic reproduction.

It has been reported that the populations of the *A. taurica* species in the world and in Turkey are autotetraploid ($2n=4x=36$), and the only Van (Zernek Dam) population was autohexaploid ($2n=6x=54$) (Civelek *et al.*, 2010; Kursat, 2010). We believe that the differences in morphological structures in Van (Zernek Dam) population samples belonging to the *A. taurica* species (Civelek *et al.*, 2010; Kursat, 2010) were caused by adaptation to different ecological conditions and higher ploidy levels.

In addition, according to the phylogenetic tree in Figure 2, samples belonging to the Zernek dam (Van) population, together with samples of other Van populations in close localities, sitting side by side on neighboring branches that can be explained in two ways:

i. It can show that different ploidy levels do not inhibit gene flow between populations and do not result in genetic isolation. However, the fact that autopolyploid species generally have apomictic reproduction thus making this less possible. Apomixis produces populations of plants of the same genotype with high levels and good adaptation.

ii. 2. Populations of the *A. taurica* species reproduce apomictically because they are autopolyploid and autohexaploid; due to apomictic reproduction (asexual seed production), populations cannot find the opportunity to differentiate genetically. This is because meiosis and crossing-over do not occur in apomictic reproduction. For this reason, newly formed embryos are genetically identical to the parent (mother) plant, as variations do not occur.

For this reason, the Van (Zernek dam) population of the *A. taurica* species were evaluated within the framework of the evolutionary species concept and intra-specific variation (micro-species) was accepted and a new variety has published (Kursat *et al.*, 2018).



Fig. 2. Maximum Likelihood tree showing the phylogenetic relationship between individuals.

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