COMPARATIVE STUDY OF PHYSIOLOGICAL AND GROWTH PARAMETERS FOR SODIUM AND IRON TOXICITIES IN UPLAND AND LOWLAND CULTIVARS OF *ORYZA SATIVA* L.

NUSRAT JAHAN^{1, 2}, MUHAMMAD ARSHAD JAVED^{1*}, FAZILAH ABD MANAN¹, SAMIULLAH KHAN^{1,2}, AZMAN ABD SAMAD¹ AND MUDASSIR ISRAR ZAIDI³

¹Faculty of Biosceinces and Medical Engineering, Universiti Teknologi Malysia (UTM), Johor Bahru 81310, Malaysia ²Faculty of life sciences and informatics, BUITEMS, Quetta, Pakistan ³ University of Balochistan, Pakistan

**Corresponding author's email: majaved@fbb.utm.my*

Abstract

Abiotic stresses are common limitations to rice productivity worldwide. Sodium and iron toxicities affect the rice crop yield adversely. The aim of this study was to compare the effects of these toxicities on seedling growth and ionic distribution in (two upland: *SK1* and *Panderas* and three lowland: *Pokkali*, *MR211* and *Firat*) rice varieties. Sodium and iron toxicities (NaCl: 0, 80mM and 120mM and FeCl₂: 0, 3.5mM and 7.5mM) were imposed separately, to three weeks old seedling for two weeks, using a hydroponic system. Results showed a significant adverse effect on growth traits with an increase in iron and sodium toxicities. Ionic distribution analysis of roots, leaves and flag leaves revealed that an increase in Na⁺ and Fe²⁺ ions reduced K⁺ uptake. *Pokkali* showed low accumulations of Na⁺ and Fe²⁺ in root, leaves and flag leaf in all stress levels of these toxicities. *Firat* showed high accumulation of Na⁺ and low accumulation of Fe²⁺ in high stress levels of these toxicities. Therefore, it is concluded that the tolerance for sodium and iron toxicities may be independent mechanisms as reflected by physiochemical analysis. However, ability to uptake potassium would help the seedling to ameliorate the effects of these toxicities.

Key words: Oryza sativa L., Abiotic Stresses, toxicity, ion distribution.

Introduction

Rice (*Oryza sativa* L) is the major nutrition source for about 40% of the world's population, including many of the people living in the developing countries (Greenland, 1997). It is cultivated on approximately 128 million hectares of irrigated and rain fed lowlands (Maclean *et al.*, 2002) and about 55% of rice cultivated fields are affected due to toxic elements like sodium and iron toxicities (Chérif *et al.*, 2009). Excess of iron and sodium in soil are the major agricultural constrains that strongly influencing the rice crop productivity (Engel *et al.*, 2012; Borsani *et al.*, 2003). They are the most important toxicities that can directly effect on plant growth and development (Galvani, 2007; Lauchli & Grattan, 2007, Arshad *et al.*, 2012).

The reduction in height and dry matter has a related with rice production and ultimately affect the rice yield. The salt injury is the main symptom of sodium toxicity whereas, iron toxicity causes the typical leaf-bronzing symptoms and these toxicities effect the rice growth. The excess ions up take, nutrient disorder and water deficiency are reported as major constraints for plants grown in sodium toxicity.

Plants possess different mechanisms to maintain the physiological concentrations of essential ions when they are exposed to high levels of toxicities (Hall 2002). Rice varieties have different level of tolerance to toxicities which have been developed by plant breeders (Sahrawat *et al.*, 1996).

Many studies have been conducted to identify the genetic and biochemical mechanisms helping plants to overcome with toxicities of toxic substance like sodium and iron ions. The mechanisms of plants play very crucial role to exclude, detoxify or compartmentalize the toxic substance toxicity for the protection of plant in a contaminated environment (Lin *et al.*, 2012).

The main objective of this study was to observe the effects of sodium and iron toxicities on plant growth and development and to screen the rice varieties possessing tolerance mechanism when exposed to high levels of sodium and iron toxicities.

Materials and Methods

Rice varieties and growth conditions: Five rice varieties, two upland (Sk1 and Panderas) and two lowland rice (MR211, Firat and Pokkali), were used to investigate the effects of sodium and iron toxicities. Seeds were soaked for 24 h in demineralized water and then transferred to Petridishes containing moist filter paper for germination. Seven days later, the germinated seeds were transferred into half strength hydroponic system for 7 days consisting of 4 liters of Yoshida solution (Samaranayake et al., 2012). The pH of the Yoshida solution for the whole experiment was retained at pH 5.0-5.5 (Wu et al., 2006). Sodium and ferrous toxicities were imposed after one week (Sodium: omM, 80mM and 120mM Iron: 0mM, 3.5mM and 7.5mM, respectively). Solution pH and nutrient solution were adjusted as described by Wu et al. (2006). Toxicities were imposed for two weeks.

Growth traits: Then seedlings were harvested to record the data for growth traits. For dry weight, the plants were washed thoroughly with deionized water and allowed to dry in an oven at 70°C for 48 hours until a constant weight was obtained. Based on these measurements, the relative decrease in shoot and root dry weight was calculated using the following formula:

Mean weight in control - Mean weight under stresses Mean weight in control x100

Leaf bronzing and salt injury score were computed by the effects reflected by the leaves under sodium and iron toxicities, respectively. Toxicity symptoms were converted to a scale of 1-5 as described by Standard Evaluation System of International Rice Research Institute (Anon., 1988). A score of 1 indicates normal growth and 5 indicates that all the leaves are dead.

Physio-chemical parameters: From each sample, 0.3 grams of ash in the test tube were dissolved in aqua regia (HCl+ HNO₃), 6ml: 2ml and kept in the aluminium block for digestion at 180°C. The samples were digested to cool and dry and were diluted with 20ml of 10% HCl, till the transparent colour appears.

Determination of the sodium, iron and potassium ions in the leaves, flag leaf and root samples were carried out separately using the Atomic Absorption Spectrophotometer (AA-7000, Shimadzu).

Statistical analysis

Data of the experiment was subjected to statistical analysis with version 16.0 of IBM-SPSS statistics. Mean and standard deviation of data were calculated.

Results and Discussion

The results of hydroponic study indicated that both Na⁺ and Fe²⁺ toxicities severely influenced the growth and physiological parameters in all varieties. However, decrease in the growth trait and physiological parameters were more pronounced at 120mM NaCl and 7.5mM FeCl₃ concentrations in the hydroponic growth medium for all rice varieties.

Effect of Na and Fe toxicities on growth parameters

Root dry Biomass: Data (Fig. 1), regarding the reduction percentage of root dry biomass showed the comparison Na⁺/Fe²⁺ toxic effect on all rice varieties of different Na⁺ and Fe²⁺ toxicity levels. In control, maximum root dry weight was recorded in *Pokkali* (3.93mg \pm 0.01, and 3.92mg±0.00) followed by Sk1 (3.89mg±0.56) and Firat $(2.46 \text{ mg} \pm 0.08)$ in both experiments. The comparison of rice cultivars response at low levels of Na⁺ toxicity (80mM) Pokkali and MR 211 showed a minimum reduction 38.73% and 37.83%, respectively, followed by Panderas (34.25%) and variety Firat showed the lowest dry root biomass (14.61%). Whereas, at low levels of Fe^{24} toxicity (3.5mm) showed the minimum reduction percentage in root dry biomass in varieties Firat (41.74%) and Panderas (40.60%) followed by MR211 (39.27%) and Pokkali (33.73%) while the lowest root dry weight was observed in SK1 (24.19%). At the highest concentration of Na⁺ toxicity, varieties MR211 (28.50%) and Pokkali (26.02%) showed minimum reduction while varieties SK1 (11.54%) and the Firat (9.33%) were

showed a maximum reduction in root dry weight. However, at high concentration of Fe²⁺ toxicity (7.5mM) varieties *Panderas* (39.12%), *MR211* (32.62%) and *Pokkali* (30.62%) performed better than other varieties, whereas *SK1* yielded lowest root dry mass. The results are in confirmation with Aboa & Dogbe (2006), Costa *et al.* (1995).

Shoot dry biomass: The results of the shoot dry biomass at different levels of Na⁺ and Fe²⁺ gevarieties MR 211 (33.09%) and Panderas (34.08%) at high toxicity of sodium are shown in Fig. 2. Var. Pokkali yielded highest (46.15%) and var. Firat produced lowest (28.29%) shoot dry weight at high (120mM) sodium toxicity. While in Fe²⁺ toxicities showed that maximum shoot dry weights 86.08%, 77.26%, 76.43%, 75.69% were produced by MR211 and Pokkali in 3.5mM and 7.5mM of Fe^{2+} toxicities, respectively. Whereas, the variety SK1 yielded 64% and 53% the lowest shoot dry mass. Similar results were also found by Purnendu et al., (2004), Maiti et al., (2006), Audebert et al., (2006) under iron and salinity stresses. Munns & Tester, (2008) also reported that the stresses might directly or indirectly inhibit cell division and enlargement during plant growing period. As a result, shoot and leaves of the affected plants appeared stunted.

Leaf bronzing and salt injury score: As shown in results (Fig. 3) leaf injury score in Na⁺ toxicity and leaf bronzing in Fe²⁺ toxicity and were significantly high in all rice varieties. At high toxicity level of Na⁺, *Pokkali* (36.29%) and *SK1*(44.37%) showed the lowest leaf injury score while *Firat* (57.28%) and *Panderas* (51.67%) showed the highest leaf injury score as represented in Fig. 5. However, at the highest level of Fe²⁺ toxicity (7.5mM) *Firat* (57.28%) and *Pokkali* (65.67%) showed minimal leaf bronzing while *Panderas* (84.75%), *MR211* (83.56%) and *SK1*(81.76%) showed maximum leaf bronzing as shown in Fig. 4.



Fig. 1. Root dry weight as affected by sodium and iron toxicities in upland and lowland rice varieties.





Fig. 2. Shoot dry biomass as affected by sodium and iron toxicities in upland and lowland rice varieties.

Fig. 3. Leaf bronzing and leaf injury score as affected by sodium and iron toxicities in upland and lowland rice varieties.



Fig. 4. Leaf brozing score in (Left to right) SK1, Panderas, MR211, Firat, Pokkali under iron toxicity



Fig. 5. Salt injury score in (left to right) SK1, Panderas, MR211, Firat, Pokkali under sodium toxicity

Physio-chemical response of varieties under sodium and iron toxicities: The Na⁺ and Fe²⁺ions quantities of the root, leaves and flag leaves of all varieties were significantly different than that of their relevant control. The Na⁺/ Fe²⁺ ions distribution in the root, leaves and flag leaf at various rice varieties at different toxic levels is given in Table 1a, b. The Na⁺ and Fe²⁺ quantities are significantly higher than their relevant controls. The highest accumulations of Na⁺ and Fe²⁺ were found at 120mM and 7.5mM concentrations.

The comparison of rice varieties at different levels showed that minimum Fe^{2+} accumulations 1.5ppm, 0.32 ppm and 0.23 ppm were found in root, leaf and flag leaf respectively, in *Pokkali*. Whereas maximum Fe^{2+} accumulations 1.86ppm and 1.7ppm were recorded in the roots of *Panderas* and *SK1* and 0.63ppm and 0.51ppm were obseved in the leaves of *SK1* and *MR211* while 0.50ppm and 0.36ppm were found in the flag leaves of *MR211* and *Sk1* respectively. Akhtar *et al.*, (2001) and Samaranayake *et al.* (2012) reported the similar findings in cotton and rice, respectively. *Pokkali* retained most of absorbed Fe^{2+} in the roots and stopped or slow the transport of Fe^{2+} to leaves and flag leaf.

 $\mathbf{K}^{+}/\mathbf{F}\mathbf{e}^{2+}$ and $\mathbf{K}^{+'}N\mathbf{a}^{+}$ Ratio: Table 1 (a) and (b) describes the K^{+} concentration in the rice varieties at different toxicity levels of $\mathbf{F}\mathbf{e}^{2+}$ and $\mathbf{N}\mathbf{a}^{+}$. On an overall mean indicated that increase in $\mathbf{N}\mathbf{a}^{+}$ / $\mathbf{F}\mathbf{e}^{2+}$ decreased K^{+}

concentration significantly at all levels of stresses, however, the decrease was more pronounced at 7.5mM FeCl₃ and 120mM NaCl concentration in growth medium. The comparison of rice varieties at 80mM and 120 mM level of Na⁺, 3.5mM and 7.5mM of Fe²⁺ showed the maximum K⁺ concentration of the minimum K⁺ concentration in different tissues of rice varieties. Decreased in K⁺ concentration with increasing Fe²⁺/Na⁺ was reported in rice (Aslam *et al.*, 1993; Li *et al.*, 2001) and in cotton (Akhtar *et al.*, 2001). At high K⁺ supplies, increased root oxidizing power may have increased K uptake by decreasing Fe²⁺/Na⁺ was reported concentration between K and Fe²⁺/Na⁺ was reported concentration dependent (Li,*et al.*, 2001).

Conclusion

The varieties showed a variable response to sodium and iron toxicities. *Pokkali* may possess both mechanisms for tolerance to iron and salinity toxicities; whereas, *Sk1* and *Firat* exhibited opposite responses to these toxicities as shown by growth traits and ion distribution patterns. These results reflected that tolerance mechanisms to these toxicities would be independent of each other. Uptake of K^+ is reduced with an increase in toxicity of sodium and iron. Therefore, availability of potassium and ability of the plant to uptake K^+ would minimize the effects of these toxicities.

Table 1. (a) Physio- chemical parameters of all rice varieties as affected by Na⁺ toxicities

Variety	Stress	Physio-chemical parameters (ppm)						
		Na-R	Na-L	Na-FL	K-R	K-L	K-FL	
SK1	Control	0.20±0.0	0.10±0.0	0.6±0.7	2.06 ± 0.0	3.20±0.0	3.42±0.0	
	80mM	$1.40{\pm}0.01$	$0.50{\pm}0.0$	1.45 ± 0.3	1.21 ± 0.0	1.79±0.0	2.17±0.0	
	120mM	$1.80{\pm}0.0$	1.47 ± 0.0	1.45 ± 0.0	1.09 ± 0.0	1.30 ± 0.0	1.84 ± 0.0	
Pendras	Control	$0.14{\pm}0.1$	0.25±0.0	0.92 ± 0.0	1.83±0.0	2.50±0.0	3.14±0.2	
	80mM	0.97 ± 0.0	1.7±0.2	1.46±0.1	1.02±0.0	1.39±0.0	1.85±0.0	
	120mM	1.01 ± 0.0	2.38±0.0	1.63±0.1	0.69±0.0	0.98±0.0	1.26±0.1	
MR211	Control	$0.18{\pm}0.0$	0.09 ± 0.0	0.06±0.0	1.79±0.0	2.68±0.0	2.56±0.2	
	80mM	$1.28{\pm}0.0$	1.17±0.1	0.50±0.0	0.97±0.0	1.78±0.0	1.58±0.0	
	120mM	$1.59{\pm}0.0$	1.90±0.1	0.85±0.0	0.48 ± 0.0	1.43±0.1	1.02±0.3	
Firat	Control	0.30±0.0	0.15±0.0	0.05±0.0	1.70±0.0	2.63±0.1	3.40±0.2	
	80mM	$0.39{\pm}0.0$	0.68 ± 0.0	0.58±0.0	1.39±0.0	1.75±0.0	2.20±0.1	
	120mM	$0.44{\pm}0.0$	1.84±0.0	1.34±0.0	0.99 ± 0.0	1.15±0.0	1.59±0.1	
Pokkali	Control	0.65 ± 0.0	0.06 ± 0.0	0.03±0.0	1.34 ± 0.0	2.56±0.0	3.64±0.1	
	80mM	$0.81{\pm}0.0$	0.48 ± 0.0	0.16±0.0	1.11±0.0	1.96±0.0	2.50±0.0	
	120mM	0.95±0.0	1.36±0.0	0.48±0.1	1.05±0.0	0.85±0.0	1.80±0.2	

Mean \pm Standard deviation. Na⁺--R = Na⁺-quantity in root, Na⁺--L = Na⁺-quantity in leaf, Na⁺--FL = Na⁺-quantity in flag leaf, K⁺-R = K⁺ quantity in root, K⁺-L = K⁺ quantity in leaf, K⁺-FL = K⁺ quantity in flag leaf.

At Na⁺ toxicity level 120mM maximum Na⁺ accumulations were 1.84ppm and 1.59 ppm in roots of *Sk1* and *MR211*, 1.63ppm and 1.45ppm in flag leaf of *Panderas* and *Sk1*, 2.38ppm and 1.9ppm in leaf *Panderas* and *MR211* respectively. While the lowest accumulations of Na⁺ were 0.95ppm in root, 0.48ppm in flag leaf and 1.36ppm in leaf found in variety *Pokkali*. Increase in a Na⁺ concentration resulting increase in toxicity was confirmed by findings of Nawaz *et al.* (1998) in wheat and Claudia *et al.* (1995) in rape seed.

Physio-chemical parameters (ppm)										
Variety	Stress	Fe ²⁺ R	Fe ²⁺ L	Fe ²⁺ FL	\mathbf{K}^{+} - \mathbf{R}	\mathbf{K}^{+} -L	K^+ -FL			
SK1	Control	1.23±0.2	0.02±0.0	0.01±0.0	1.43±0.2	2.76±0.0	2.56±0.4			
	3.5mM	1.26±0.0	$0.40{\pm}0.0$	0.27±0.1	0.91±0.0	2.40±0.1	2.43±0.2			
	7.5mM	1.70±0.1	0.63±0.1	0.36±0.1	0.87 ± 0.0	2.20±0.1	2.03±0.1			
Panderas	Control	1.13±0.1	0.02 ± 0.0	0.01±0.0	1.40±0.2	2.23±0.1	2.70±0.0			
	3.5mM	1.23±0.1	0.29±0.0	0.18±0.0	$0.78{\pm}0.0$	2.13±0.2	2.30±0.1			
	7.5mM	1.86±0.3	0.39±0.0	0.38±0.0	$0.70{\pm}0.0$	1.63±0.0	1.93±0.0			
MR211	Control	1.13±0.4	0.01±0.0	0.01±0.0	1.83 ± 0.0	2.00±0.3	2.86±0.2			
	3.5mM	1.31 ± 0.1	0.28±0.0	0.22±0.0	0.93±0.1	2.60±0.2	2.40±0.1			
	7.5mM	1.41±0.1	0.51±0.0	0.50±0.0	$0.94{\pm}0.0$	2.10±0.1	2.10±0.2			
Firat 1	Control	0.90±0.1	0.01±0.0	0.01±0.0	1.90±0.1	2.70±0.3	2.76±0.2			
	3.5mM	1.16±0.1	0.20±0.0	0.17±0.0	1.30 ± 0.0	2.30±0.0	2.60±0.3			
	7.5mM	1.23±0.3	0.40±0.3	0.29±0.0	0.66 ± 0.0	1.96±0.1	2.36±0.1			
Pokkali	Control	0.94±0.0	0.01 ± 0.0	0.01±0.0	1.18±0.1	2.93±0.3	2.93±0.0			
	3.5mM	1.20±0.0	0.20±0.0	0.17±0.0	0.84±0.0	2.46±0.2	2.53±0.0			
	7.5mM	1.50±0.2	0.32±0.0	0.23±0.0	0.62±0.0	2.22±0.1	2.50±0.1			

Table 1. (b) Physio-chemical parameters of all rice varieties affected by Fe²⁺

Mean ± Standard deviation. $Fe^{2+}-R=Fe^{2+}$ -quantity in root, $Fe^{2+}-L=Fe^{2+}$ -quantity in leaf, $Fe^{2+}-FL=Fe^{2+}$ -quantity in flag leaf, $K^+-R=K^+$ quantity in root, $K^+-L=K^+$ quantity in flag leaf

Acknowledgements

Research Management Centre (RMC) of Universiti Teknologi Malaysia (UTM) is acknowledged for the financial assistance Cost Center No. Q.J130000.2545.05H93

References

- Aboa, K. and S. Dogbe. 2006. Effect of iron toxicity on rice yield in the Amouoblo lowland in Togo. *Iron Toxicity in Rice-based Systems in West Africa*. 1-5.
- Akhtar, J. and F.M. Azhar. 2001. Response of cotton (*Gossypium hirsutum* L.) hybrids to NaCl salinity at seedling stage. *Int. J. Agric. Bio.*, 3: 233-235.
- Anonymous. 1988. International Rice Research Institute, Standard Evaluation System for Rice, p: 37. Los Banos, Laguna, Philippine
- Audebert, A., L.T. Narteh, P. Kiepe, D. Miller and B. Beks. 2006. Iron Toxicity in Rice-based Systems in West Africa. Africa Rice Centre (WARDA).
- Arshad, M., M. Saqib, J. Akhtar and M. Asghar. 2012. Effect of calcium on the salt tolerance of different wheat (*Triticum* aestivum L.) genotypes. *Pak. J. Agri. Sci.*, 49: 497-504.
- Aslam, M., R.H. Qureshi and N. Ahmad, 1993. A rapid screening technique for salt tolerance in rice. *Plant & Soil*, 150: 99-107
- Borsani, O., V. Valpuesta and M.A. Botella. 2003. Developing salt tolerant plants in a new century: a molecular biology approach. *Plant Cell Tissue Organ Cult.*, 73: 101-115.
- Claudia, A.P., F.H. Boem and R.S. Lavado. 1995. The K/Na and Ca/Na ratios and rape seed yield under salinity or sodicity. *Plant & Soil.* 175: 225-251.
- Chérif, M., A. Audebert, M. Fofana and M. Zouzou. 2009. Evaluation of iron toxicity on lowland irrigated rice in West Africa. *Tropicultura*, 27: 88-92.

- De Costa, W.A.J.M. and G.J.K. De Zoysa. 1995. Effects of water stress on root and shoot growth of soyabean (*Glycine max* (L.) Merill) and rice (*Oryza sativa* L.). Sri Lanka J. Agri. Sci., 32: 134-142.
- Engel, K., F. Asch and M. Becker. 2012. Classification of rice genotypes based on their mechanisms of adaptation to iron toxicity. J. Plant Nutr. & Soil Sci., 175(6): 871-881.
- Galvani, A. 2007. The challenge of the food sufficiency through salt tolerant crops. *Rev. Env. Sci. Biotech.* 6(1-3): 3-16.
- Greenland, D.J. 1997. *The sustainability of rice farming*. Wallingford, UK: CAB International.
- Hall, J.L. 2002. Cellular mechanisms for heavy metal detoxification and tolerance. J. Exp. Bot., 53 (366): 1-11.
- Koyro, H.W. and B. Huchzermeyer. 1999. Salt and drought stress effects on metabolic regulation in maize. In: *Handbook of plant and crop stress.* (Ed.): M. Pessarakli. Second ed. rev. and exp. Marcel Dekker, New York.
- Li, H., X. Yang and A. Luo. 2001. Ameliorating effect of potassium on iron toxicity in hybrid rice. J. Plant Nutr., 24(12): 1849-1860.
- Lin, Y.F., G. Mark and M. Aarts. 2012. The molecular mechanism of zinc and cadmium stress response in plants. *Cell. & Mol. Life Sci.*, 69(19): 3187-3206.
- Lauchli, A and S.R. Grattan. 2007. Plant growth and development under salinity stress. In: Advances in Molecular Breeding Toward Drought and Salt Tolerant Crops, (Eds.): M.A. Jenks, P.M.Hasegawa and S.M. Jain. Springer, Dordrecht, Netherlands
- Maiti, R.K., P. Vidyasagar and P.P. Banerjee. 2006. Salinity tolerance in rice (*Oryza sativa* L.) hybrids and their parents at emergence and seedling stage. *Crop Res. Hisar.*, 31(3): 427-433.
- Maclean, J.L., D.C. Dawe, B. Hardy and G.P. Hettel. 2002. Rice almanac: Source book for the most important economic activity on earth (3rd Eds). *International Rice Research Institute, Manilla, Philippines and CABI, Wallingford, UK*, 253.

- Munns, R. and M. Tester. 2008. Mechanisms of salinity tolerance. *Annu. Rev. Plant Biol.*, 59: 651-681.
- Nawaz, S., R.H. Qureshi, M. Aslam, J. Akhtar and S. Parveen. 1998. Comparative performance of different wheat varieties under salinity and water logging. II. Ionic relations. *Pak. J. Bio. Sci.*, 1(12): 357-359.
- Purnendu, G., M.A. Mannan, P.S. Pal, M.M. Hossain and S. Parvin. 2004. Effect of salinity on some yield attributes of rice. *Pak. J. Biol. Sci.*, 7(5): 760-762.
- Sahrawat, K.L., C.K. Mulbah, S. Diatta, R.D. Delaune, W.H. Patrick, B.N. Singh and M.P. Jones. 1996. The role of tolerant genotypes and plant nutrients in the management of iron toxicity in lowland rice. J. Agri. Sci., 126: 143-149.
- Samaranayake, P., B.D. Peiris and S. Dssanyake. 2012. Effect of excessive ferrous (Fe²⁺) on growth and iron content in rice (*Oryza sativa*). *Int. J. Agric. Biol.*, 14(2): 296-298.
- Wu, F., G. Zhang, Jia and S. Zheng. 2006. Genotypic difference in the responses of seedling growth and Cd toxicity in rice (Oryza sativa L.). Agri. Sci. China., 5(1): 68-76.

(Received for publication 15 January 2016)