VEGETATIVE GROWTH PERFORMANCE OF FIVE MEDICINAL PLANTS UNDER NaCI SALT STRESS

ZAHIR MUHAMMAD AND FARRUKH HUSSAIN

Department of Botany, University of Peshawar, Peshawar, Pakistan

Abstract

Seeds of *Lepidium sativum* L., *Linum usitatissimum* L., *Nigella sativa* L., *Plantago ovata* Forssk, and *Trigonella foenum-graecum* L. were grown in pots containing loamy soil with 0.21 (Control) 5.0, 7.5, 10.0, 12.5, or 15.0 dS/m concentration of NaCl to see their salinity tolerance.

Various concentrations of salt had a highly significant effect upon the survival %age, plant height, number of branches, shoot fresh and dry weight, root fresh and dry weight and root moisture contents. Number of leaves also varied significantly. However, leaf length and shoot moisture contents exhibited non-significant differences. Differences among the test species for all the parameters under consideration were also highly significant.

The findings suggest that the test species are tolerant to moderate salinity i.e., 7.5 dS/m and might be tried on saline soils to obtain some biomass.

Introduction

Salinity adversely reduces the overall productivity of plants including crops by inducing numerous abnormal morphological, physiological and biochemical changes that cause delayed germination, high seedling mortality, poor crop stand, stunted growth and lower yields. Out of 20.2 million hectares of cultivated land in Pakistan, 6.8 million hectares are affected with some degree of salinity (Anon., 2002). Generally screening and cultivation of crops on the salt affected soil is the main approach. Germination and seedling growth under saline environment are the screening criteria which are widely used to select the salt tolerant genotype (Ashraf *et al.*, 1990; Khan *et al.*, 1993). Medicinal plants are important source of medicines and livelihood. They are either collected from natural habitat or imported. Salted soils that can not support crops might be used for medicinal plants as novel crops.

Some work on the effect of salinity on germination and growth of medicinal plants include Linum usitatissimum (Ashraf & Fatima, 1994; Ashraf & Tufail, 1995; Beke & Graham, 1995), Plantago spp. (Boss, 1992; Tanczos et al., 1992), Trigonella foenumgraecum (Ali et al., 1992), Nigella sativa (Hajar et al., 1996), Helianthus annuus (Ashraf & Tufail, 1995; Ashraf et al., 2003; Mutlu & Bozcuk, 2007), Glycine max (Umezawa et al., 2000; Essa 2002), Brassica spp. (Ashraf & Naqvi, 1997; Farhandi & Sharif Zadah, 2006; Gul & Ahmad, 2007; Ulfat et al., 2007; Jenagand et al., 2008), Ricinus communis (Raghavaiah, 2002, 2006). It appears that little information is available regarding the effect of salinity on the growth and productivity of medicinal plants. Lepidium sativum L., Linum usitatissimum L., Plantago ovata Forssk and Trigonella foenum-graecum L. have been evaluated and proved to be moderately salt tolerant at germination and seedling growth stage (Muhammad & Hussain, 2009). The present study was focused to assess the tolerance of these medicinal plants towards salinity at their vegetative growth. The findings might help in utilizing saline habitats. The successful cultivation of medicinal plants will provide raw material to pharmaceutical companies and for local medicinal uses.

Materials and Methods

Seed of *Plantago ovata* Forssk, *Lepidium sativum* L., *Trigonella foenum-graecum* L., *Linum usitatissimum* L. and *Nigella sativa* L. were obtained from the local market. They were grown in pots salinized with 0.21 (Control), 5.0, 7.5, 10.0, 12.5, or 15.0 dS/m NaCl concentration. There were four replicates for each treatment. Each pot had 5 Kg loamy soil lined with polyethylene bags to avoid salt leaching. The soil used had 8.75% CaCO₃, 0.65% organic matter, 0.032% nitrogen, 20.9 ppm phosphorus, 413.5 ppm potassium, 7.6 pH level, 0.21 dS/m electrical conductivity, 0.067% total soluble salts, 5.2% clay, 17.0% silt and 77.8% sand.

Generally, it took 3-4 days for the first emergence of seedlings and within 6-7 days the germination completed in all the species. However, *Nigella* took 14 days to germinate and more than 25 days for the maximum possible germination. Seedlings were thinned to 10 individuals/pot after one month of the maximum germination for each species. Uniform equi-distant and healthy seedlings were left in each pot. Pots were brought to the desired salinity levels by providing 2.5 dS/m salinity levels in 6 equal increments. Salinity was measured with EC meter.

The pots were kept in the net house of Botany Department, University of Peshawar under uniform open environmental conditions. They were maintained at field capacity and protected from rain or dew by polyethylene sheets. Growth parameters such as height of plants, number of leaves, number of branches or tillers and leaf length were recorded at the time of the harvest. Weekly data for the plant survival was also recorded after salt treatment. Crops were harvested when matured. Roots were carefully taken out of soil. Fresh weight of shoots and roots were determined. They were oven dried at 65°C for 72 hours for dry weight determination.

The moisture contents of shoots and roots were determined on oven dry basis (Hussain, 1989). The results were subjected to ANOVA (Steel & Torie, 1980) and significant differences were accepted at p = 0.05.

Results

1. Effect on survival of seedlings: Highly significant differences were observed in the survival % age under different salt treatments and among the species (Table 1). In control the survival was 100%, which decreased gradually with increasing concentration of salt. The differences were non-significant up to 10.0 dS/m salt concentration thereafter the differences were statistically sound at 12.5 dS/m (59.2%) and 15.0 dS/m (33.8%). *Lepidium* exhibited significantly higher survival % age (89.5%), closely followed by *Trigonella* (83.3%) and *Plantago* (83.0%) showing insignificant differences. Survival rate in *Linum* (68.7%) and *Nigella* (68.2%) was statistically at par with each other but significantly different from the aforementioned species (Table 2).

2. Effect on plant height and branching/tillering: ANOVA revealed highly significant differences in the plant height under various applied salt levels and between the test species (Table 1). The plant height was significantly greater in control (40.56 cm), which was closely followed by 5.0 dS/m (31.44 cm). The plant height decreased gradually with every increasing level in the salt concentration reaching to minimum of (7.26 cm) at the highest dose. *Lepidium* exhibited maximum height (32.32 cm), that was respectively followed by *Linum* (26.80 cm), *Trigonella* (23.07 cm), *Nigella* (20.62 cm) and *Plantago* (16.22 cm) (Table 3).

			Ta	ible 1. Mear	n squares of	the analysi	is of variance f	or various par	rameters.			
Source	d.f	Survival (%)	Plant height (cm)	Number of branches	Number of leaves	Leaf length (mm)	Shoot fresh weight (mg)	Shoot dry weight (mg)	Root fresh weight (mg)	Root dry weight (mg)	Shoot moisture contents (%)	Root moisture contents (%)
Concentration	5	3635.25**	723.50^{**}	22.94**	996.71*	45.00 ^{NS}	3406093.92*	355225.91**	6707036.38*	180620.53**	8121.19 ^{NS}	22505.36**
Species	4	552.12*	224.57**	59.18**	3720.38**	706.84**	3768146.26*	348869.28**	9273574.48*	192214.87**	153627.95*	73205.13**
Error	20	162.14	30.67	5.17	364.48	26.87	338633.05	19419.78	972143.98	22707.12	15225.07	5299.31
Total	29											
Table 2. Effec	ct of va	arious level	s of NaCl	(dS/m) sal	linization o	n the surv	vival rate (%). Each value	e is a mean of	4 replicates	, each with	10 plants.
Concentra	ation	г 2	epidium sativum	511	Linum sitatissimun		Nigella sativa	Plantago ovata	Tri foenui	igonella n-graecum	Tre	atment iean
Contro	<u> </u>		100.0		100.0		100.0	100.0		100.0	10	0.0 ^A
5.0			100.0		100.0		100.0	97.0		100.0	6	9.4 ^A
7.5			100.0		100.0		95.0	92.0		100.0	6	7.4 ^A
10.0			95.0		67.0		75.0	90.0		80.0	81	.4 ^{AB}
12.5			82.0		40.0		32.0	62.0		80.0	5	9.2 ^B
15.0			60.0		5.0		7.0	57.0		40.0	Ω.	3.8 ^c
Species n	ıean		89.5 ^A		68.7 ^B		68.2 ^B	83.0 ^{AB}	8	(3.3 ^{AB}		
LSD value at 0.0)1 alpha	a level for trea	atment mean	n = 22.91 an	id for species	mean = 15	.34					

GROWTH PERFORMANCE OF MEDICINAL PLANTS UNDER SALT STRESS

305

Means in the last column/row sharing the same letter do not differ significantly from each other at 1% or 5% level of probability.

	Treatment	mean	40.56^{A}	31.44^{AB}		28.04^{BC}		20.09^{CD}		14.62 ^{DE}		7.26^{E}				Ireatment	mean	7.58^{A}	7.00^{Λ}		6.24^{A}		$5.30A^{B}$		3.84^{AB}		1.86 ^B		
sight (cm).	Trigonella	foenum-graecum	38.8	29.7	76.6	25.1	64.6	19.5	50.3	17.2	44.4	8.1	20.9	23.07^{BC}	branches or tillers.	Irigonella	foenum-graecum	5.6	4.6	81.3	4.5	80.4	3.1	54.5	3.6	64.3	1.4	25.2	3.80 ^B
linization on the he ch with 10 plants.	Plantago	ovata	23.3	20.4	87.6	18.3	78.6	16.1	69.1	10.0	43.0	9.2	39.6	16.22 ^C	on the number of ch with 10 nlants	Plantago	ovata	5.3	4.8	90.9	4.4	83.9	4.0	76.3	3.1	58.3	3.0	56.8	4.10^{B}
of NaCl (dS/m) sal of 4 replicates, ea	Nigella	sativa	40.7	29.3	72.0	26.1	64.0	18.1	61.7	7.5	18.4	2.0	5.0	20.62^{BC}	dS/m) salinization of 4 renlicates, ear	Nigella	sativa	5.2	5.5	105.7	4.3	81.8	4.0	76.5	2.8	53.2	1.1	20.7_{-}	3.78 ^B
t of various levels o ch value is a mean	Linum	usitatissimum	52.9	41.6	78.6	34.4	65.0	20.6	39.0	11.1	21.0	0.2	0.3	26.80^{AB}	us levels of NaCl (ch value is a mean	TIMUM	usitatissimum	16.9	16.2	95.9	13.4	79.2	11.6	68.8	6.2	36.6	1.2	7.1	10.92^{A}
Table 3. Effect Eac	Lepidium	sativum	47.3	36.2	76.5	36.3	76.7	30.2	63.8	27.3	57.7	16.8	35.5	32.32^{A}	e 4. Effect of vario	 Lepidium	sativum	4.9	4.1	84.3	4.6	93.9	3.8	77.6	3.5	71.4	2.6	52.0	3.92 ^B
	Concentration	(dS/m)	Control	5.0	% of Control	7.5	% of Control	10.0	% of Control	12.5	% of Control	15.0	% of Control	Species mean	Tabl	Concentration	(dS/m)	Control	5.0	% of Control	7.5	% of Control	10.0	% of Control	12.5	% of Control	15.0	% of Control	Species mean

306

ZAHIR MUHAMMAD & FARRUKH HUSSAIN

 % of Control
 77.6 6.8.8 7.5 7.6.5 7.6.3

 12.5 3.5 6.2 2.8 3.1 5.3.2 58.3 3.1

 % of Control
 71.4 36.6 53.2 58.3 3.1

 % of Control
 2.6 1.2 1.1 3.0 58.3

 % of Control
 2.6 1.2 1.1 3.0 58.3

 % of Control
 52.0 7.1 20.7 56.8 4.10^8

 Species mean
 3.78^8 4.10^8 4.10^8 Means in the last column/row sharing the same letter do not diffic significantly from each other at 1% level of probability

ANOVA showed highly significant differences in the average number of branching both under different salt treatments and between the test species (Table 1). The number of branching/tillering gradually decreased with increasing salt concentration. They were maximum (7.58) in control and minimum (1.86) at 15.0 dS/m treatment. The differences were insignificant within the range of 5.0 to 12.5 dS/m treatments. The number of branches in all the treatments in *Linum* (10.92) was significantly greater than *Plantago* (4.10), *Lepidium* (3.92), *Trigonella* (3.80) and *Nigella* (3.78), which were statistically at par with each other (Table 4).

3. Effect on the number and length of leaves: The number of leaves varied significantly under various salt concentrations and among the species (Table 1). The number of leaves at 7.5 dS/m level (47.14) differed insignificantly from control (45.34) but varied significantly from 15.0 dS/m salt treatment. The number of leaves in *Linum* (78.43) differed significantly from *Trigonella* (33.98), *Nigella* (23.17), *Plantago* (22.13) and *Lepidium* (18.05), which differed non-significantly from each other (Table 5).

Statistically non-significant differences were observed for average leaf length under various salt concentrations but the differences between the test species were highly significant (Table 1). The leaf length showed a declining trend with increasing level of salt concentration as it was maximum at control (33.98 mm) and the minimum (25.80) at 15.0 dS/m. *Nigella* had the maximum leaf length (43.67 mm) in all the salt treatments, which was significantly higher than *Trigonella* (32.78 mm), *Lepidium* (28.22 mm), *Linum* (20.72 mm) and *Plantago* (15.75 mm) (Table 6).

4. Effect on fresh and dry weight of shoot: Differences in the fresh and dry weight of shoots were highly significant both under various levels of salinization and among the species (Table 1). The shoot fresh weight was maximum (2874.4 mg) in control. that decreased gradually to a minimum of 416.0 mg at 15.0 dS/m. Average shoot fresh weight in all the treatments for *Trigonella* (2335.0 mg), *Plantago* (2054.7 mg) and *Linum* (1794.2 mg) differed insignificantly among themselves, while, shoot fresh weight in *Nigella* (743.7 mg) and *Lepidium* (578.8 mg) were statistically at par with each other however, but varied significantly from the aforementioned species (Table 7).

Average dry weight of shoot for all the tested species was maximum (951.0 mg) in control that dwindled to 172.5mg at 15.0 dS/m linearly with increasing salt stress. *Plantago* had the highest weight (780.0 mg), differing significantly from *Lepidium* (298.3 mg) and *Nigella* (201.7 mg) (Table 8).

5. Effect on fresh and dry weight of roots: The differences in fresh and dry weight of roots were highly significant in various salt treatments and among the species (Table 1). The fresh weight of roots was maximum in control (3426.0 mg). It reduced considerably under salt stress, especially under higher concentration, yet the effect was non-significant within various salt concentrations. The fresh weight of *Linum* (3350.0 mg) was significantly higher in all the treatments followed by *Plantago* (2067.9 mg). The root fresh weight in *Nigella* (819.6 mg), *Trigonella* (503.3 mg) and *Lepidium* (502.9 mg) were statistically at par with each other but differed significantly from *Linum* (Table 9).

The dry weight of roots in all the species was significantly greater in control that reduced gradually with increasing salt level with a minimum at 15.0 dS/m. The root dry weight in all the treatments was significantly higher in *Linum* (516.3 mg) which was statistically at par with *Plantago* (347.1 mg), quickly followed by *Nigella* (155.0 mg), *Lepidium* (115.4 mg) and it was the minimum in *Trigonella* (106.1 mg) (Table 10).

	5	THAT I I I ANTH IA	a forestation and a second			
Concentration	Lepidium	Linum	Nigella	Plantago	Trigonella	Treatment
(dS/m)	sativum	usitatissimum	sativa	ovata	foenum-graecum	mean
Control	21.6	105.0	31.9	28.2	40.0	45.34^{MB}
5.0	22.6	115.1	32.3	25.1	38.2	46.66^{A}
% of Control	104.9	109.6	101.1	89.0	95.4	
7.5	19.6	127.6	28.5	23.0	37.0	47.14^{Λ}
% of Control	90.7	121.6	89.3	81.4	92.5	
10.0	16.4	80.9	27.5	22.4	32.8	36.00^{ABC}
% of Control	76.1	77.0	86.2	79.3	81.9	
12.5	14.8	32.4	13.9	15.2	29.0	21.06^{BC}
% of Control	68.4	30.9	43.8	53.8	72.5	
15.0	13.3	9.6	4.9	18.9	26.9	14.72^{C}
% of Control	61.4	9.2	15.4	67.1	67.3	
Species mean	18.05^{B}	78.43^{A}	23.17 ^B	22.13 ^B	33.98 ^B	
D value at 0.05 alpha level f	or treatment mean $= 2$	5.19 and for species mea	an = 31.36			
ans in the last column/row s	haring the same letter	do not differ significant	ly from each other a	tt 1% level of probabi	lity	
	Table 6. Effect of	various levels of Na(Cl (dS/m) saliniz	ation on the length	(mm) of leaves.	
Concentration	Lepidium	ican of 20 leaves sele Linum	cted alternately <i>Nigella</i>	Plantago	eacn treatment. Trigonella	Treatment
(dS/m)	sativum	usitatissimum	sativa	ovata	foenum-graecum	mean
		~ • • •	~ • • •			00.00

Concentration	Levidium	Linum	Nigella	Plantago	Trigonella	Treatment
(dS/m)	sativum	usitatissimum	sativa	ovata	foenum-graecum	mean
Control	27.3	24.6	64.6	18.3	35.1	33.98
5.0	25.1	22.4	45.0	15.9	32.9	28.26
% of Control	91.9	91.1	69.7	87.1	93.9	
7.5	28.0	18.3	44.9	15.5	34.2	28.18
% of Control	102.6	74.4	69.5	85.0	97.4	
10.0	29.2	18.5	39.8	15.0	32.0	27.08
% of Control	107.2	75.0	61.6	82.2	91.2	
12.5	29.6	18.5	38.0	14.5	29.7	26.06
% of Control	108.6	75.3	58.8	79.2	84.7	
15.0	30.1	22.0	29.7	15.3	31.9	25.80
% of Control	110.3	89.4	46.0	83.6	90.9	
Species mean	28.22BC	20.72CD	A3 67A	15 75 ^D	22 78 ^B	

LSD value at 0.01 alpha level for species mean = 8.516 LSD value at 0.01 alpha level for species mean = 8.516 Means in the last row sharing the same letter do not differ significantly from each other at 1% level of probability

(dS/m) Control 5.0 % of Control	sativum		angena a			TICALINE
Control 5.0 % of Control		usitatissimum	sativa	ovata	Joenum-graecum	mean
5.0 % of Control	1087.5	3117.5	1572.2	3260.0	5335.0	2874.4^{Λ}
% of Control	655.0	2660.0	957.5	2365.0	2305.0	1788.5 ^B
	60.2	85.3	60.9	72.6	43.2	
7.5	632.5	1735.0	862.5	2020.0	2400.0	1530.0^{B}
% of Control	58.2	55.7	54.3	62.0	45.0	
10.0	397.5	2057.5	677.5	1945.0	1900.0	1395.5 ^{BC}
% of Control	36.6	66.0	42.6	82.2	35.6	
12.5	427.5	950.0	302.5	1525.5	1810.0	1003.1^{BC}
% of Control	39.3	30.5	19.0	46.7	33.9	
15.0	272.5	245.0	90.0	1212.5	260.0	416.0°
% of Control	25.1	7.9	5.7	37.2	4.9	
Species mean	578.8 ^B	1794.2^{A}	743.7^{B}	2054.7^{A}	2335.0^{A}	
Concentration	Lepidium	Limum	Nigella	Plantago	Trigonella	Treatment
(dS/m)	sativum	usitatissimum	sativa	ovata	foenum-graecum	mean
Control	480.0	1062.5	767.5	1095.0	1380.0	951.0^{A}
5.0	327.5	827.5	182.5	970.0	660.0	593.5 ^B
% of Control	68.2	9.77	23.8	88.6	47.8	
7.5	332.5	647.5	145.0	842.5	635.0	520.5^{BC}
% of Control	96.3	60.9	18.9	76.9	46.0	
10.0	285.0	552.5	92.5	735.0	489.0	430.8^{BC}
% of Control	59.4	52.0	12.1	67.1	34.8	l
12.5	232.5	330.0	42.5	545.0	475.0	325.0^{CD}
% of Control	48.4	28.2	5.5	49.8	34.4	4
15.0	132.5	57.5	10.0	492.5	170.0	172.5 ⁰
% of Control	27.6	5.4	1.3	45.0	12.3	
Species mean	298.3 ^B	579.6 ^A	201.7^{B}	780.0^{Λ}	634.8^{Λ}	

(dS/m) Control		Linum	Nigella	Plantago	Trigonella	Treatment
Control	sativum	usitatissimum	sativa	ovata	foenum-graecum	mean
	772.5	7235.0	3060.0	4742.5	1320.0	3426.0^{Λ}
5.0	710.0	4927.5	792.5	2617.5	485.0	1912.5^{AB}
% of Control	19.9	68.1	25.9	55.2	36.7	
7.5	610.0	4072.5	590.0	2167.5	495.0	1587.0^{B}
% of Control	79.0	56.3	19.3	45.7	37.5	
10.0	375.0	2510.0	320.0	1490.0	305.0	1000.0^{B}
% of Control	48.5	34.7	10.5	31.4	23.1	
12.5	387.5	1105.0	132.5	987.5	380.0	598.5 ^B
% of Control	50.2	15.3	4.3	20.8	28.8	
15.0	162.5	250.0	22.5	372.5	35.0	168.5^{B}
% of Control	21.0	3.5	0.7	7.9	2.7	
species mean	502.9^{B}	3350.0^{A}	819.6 ^B	2067.9^{AB}	503.3 ^B	
oncentration	Lepidium	Linum	Nigella	Plantago	Irigonella	Ireatment
(dS/m)	sativum	usitatissimum	sativa	ovata	Joenum-graecum	mean
Control	162.5	1145.0	457.5	875.0	245.0	577.0 ^A
5.0	165.0	695.0	182.5	422.5	110.0	315.0^{AB}
% of Control	101.5	60.7	39.9	48.3	44.9	
7.5	140.0	612.5	145.0	330.0	110.0	267.5 ^{BC}
% of Control	86.2	53.5	31.7	37.7	44.9	
10.0	90.0	405.0	92.5	235.0	70.0	178.5 ^{BC}
% of Control	55.4	35.4	20.2	26.9	28.6	
12.5	90.0	195.0	42.5	147.5	85.0	112.0^{BC}
% of Control	55.4	17.0	9.3	16.9	34.7	
15.0	45.0	45.0	10.0	72.5	16.5	37.8 ^C
% of Control	27.7	3.9	2.2	8.3	6.7	
species mean	115.4 ^B	516.3^{A}	155.0^{B}	347.1^{AB}	106.1 ^B	

ZAHIR MUHAMMAD & FARRUKH HUSSAIN

Moisture contents of roots showed highly significant differences under various salt treatments and between species (Table 1). Plants in control (471.4%) had the highest moisture contents that declined steadily under salt stress especially at 15.0 dS/m level (273.5%). Moisture contents in *Linum* (524.7%) were significantly higher and were statistically at par with *Plantago* (505.9%). The root moisture contents in *Lepidium* (325.0%), *Trigonella* (320.8%) and *Nigella* (298.8%) showed non-significant differences among themselves but varied significantly from *Linum* and *Plantago* (Table 12).

Discussion

Salt stress reduces plant growth and productivity by affecting morphological, anatomical, biochemical and physiological characteristics, processes and functions. Disturbed water and nutritional balance of plants may cause reduced crop yield in saline soil. Reduced plant height and other morphological characters are the most distinct and obvious effects of salt stress. Depressed growth due to high salinity is attributed to several factors such as, water stress, specific ion toxicity and ion imbalance stress or induced nutritional deficiency. In the present study, plant height of all the test species decreased at higher salinity levels. Plants growing at 15.0 dS/m were stunted as compared to the control. The reduced plant height might be attributed to the direct effect of excess salt on plant tissues and poor intake of minerals. Reduced plant height under saline conditions has been observed in Linum usitatissimum (Singh & Singh, 1991; Kheir et al., 1991). Mamo et al., (1996) reported reduced plant height in chickpea variety in response to salinity. Essa (2002) reported significantly reduced plant height in soybeen by soil salinity. Mensah et al., (2006) reported that plant height tended to decrease with increasing salinity in Arachis hypogaea. Sadat-Noori (2006) also reported reduced plant height in wheat under different saline conditions. All these workers support the present findings.

Salt tolerance at vegetative stage is crucial for yielding vigorous plants for tolerating salt stress at later stages of growth. The observed reduced degree of branching at higher salt concentration directly affects the productivity, biomass and seed yield. Decreased branching due to salt stress in different oil seed crops (Singh *et al.*, 1988; Narash *et al.*, 1993; Mensah *et al.*, 2006; Sadat-Noori, 2006) has been reported and our results agree with them.

The number of leaves and leaf length got suppressed in all the test species under higher salinity regimes. Generally, leaf thickness increases under salt stress, which decreases leaf area. Letchamo *et al.*, (1993) reported significant decrease in the number of leaves of *Passiflora edulis* under high salinity and our findings agree with them. Likewise, Singh & Singh (1991) observed a considerable reduction in the leaf length of *Linum* under high NaCl stress. Leaf area is a good indicator of water and salinity stress, since leaf expansion generally requires a high turgor pressure for cell enlargement (Krieg, 1983). It is well accepted that osmotic adjustment plays a crucial role in plant adaptation to drought (Quisenberry, 1982). Salinity induced osmotic stress is considered responsible for the reduced leaf area in Canola and wild mustard (Huang & Redmann, 1995). Hajar *et al.*, (1996) and Murillo-Amador & Trovo-Dieguez (2002) also reported decreased leaf area under increasing salinity levels in *Nigella sativa* and cowpea, respectively. Our findings agree with the previous workers in this respect.

Concentration	Lepidium	Linum	Nigella	Plantago	Trigonella	Treatmen
(dS/m)	sativum	usitatissimum	sativa	ovata	foenum-graecum	mean
Control	126.6	193.4	107.2	197.7	286.6	182.3
5.0	100.0	221.5	430.1	143.8	249.2	228.9
% of Control	79.0	114.5	401.4	72.7	87.0	
7.5	90.2	168.0	494.8	139.8	278.0	234.2
% of Control	71.3	86.8	461.7	70.7	97.0	
10.0	39.5	272.4	632.4	164.6	295.8	280.9
% of Control	31.2	140.8	590.1	83.3	103.2	
12.5	83.9	216.7	611.8	179.4	281.1	274.6
% of Control	66.3	112.0	570.8	90.7	98.1	
15.0	105.7	326.1	800.0	146.2	52.9	286.2
% of Control	83.5	168.6	746.5	73.9	18.5	
Species mean	91.0^{B}	233.0^{B}	512.7^{A}	161.9 ^B	240.6^{B}	

of shoots per plant.	
n on the moisture contents (%	tes, each with 10 nlants.
vels of NaCl (dS/m) salinization	ich value is a mean of 4 renlica
Table 11. Effect of various le	E.

plant.	
per	
roots	
90 (e	
્રે	
contents	monte
ure	110
oist	T.
on m	dooo
tion	400
niza	diag
saliı	1
Î	5
(dS/	100
õ	
Ž	
Is of	allu -
leve	ي واد
ous	ŗ
vario	
đ	
Effect	
4	
Table	
L	

0.01	1.001	320.1	800.0	140.2	6.75	280.2
% of Control	83.5	168.6	746.5	73.9	18.5	
Species mean	91.0^{B}	233.0^{B}	512.7 ^A	161.9 ^B	240.6^{B}	
LSD value at 0.01 alpha level 1	for specie mean = 202.	6				
Means in the last row sharing t	he same letter do not d	iffer significantly from e	each other at 1% lev	el of probability		
Table 1	2. Effect of various	s levels of NaCl (dS/1	m) salinization or	1 moisture content	s (%) of roots per plant.	
	E	ach value is a mèan e	of 4 replicates, ea	ch with 10 plants.		
Concentration	Lepidium	Linum	Nigella	Plantago	Trigonella	Treatment
(dS/m)	sativum	usitatissimum	sativa	ovata	foenum-graecum	mean
Control	375.4	531.9	568.9	442.0	438.8	471.4 ^A
5.0	330.3	609.0	334.3	519.5	340.9	426.8^{A}
% of Control	88.0	114.5	55.8	117.5	77.7	
7.5	335.7	565.4	306.9	556.8	350.0	423.0^{A}
% of Control	89.4	106.3	54.0	126.0	79.8	
10.0	316.7	519.8	246.0	534.0	335.7	390.4^{AB}
% of Control	84.4	97.7	43.2	120.8	76.5	
12.5	330.6	466.7	211.8	569.5	347.1	385.1 ^{AB}
% of Control	88.1	87.7	37.2	128.8	79.1	
15.0	261.1	455.6	125.0	413.8	112.1	273.5 ^B
% of Control	69.6	85.7	22.0	97.7	25.6	
Species mean	325.0^{B}	524.7^{A}	298.8^{B}	505.9^{A}	320.8^{B}	
LSD value at 0.01 alpha level 1	for treatment mean = 1	31.0 and for species me	an = 119.6			
Means in the last column/row s	sharing the same letter	do not differ significant	ly from each other a	t 1% level of probabili	ity	

Reduced fresh weight of shoots under test conditions was noticed in all the test species, except *Lepidium* at 12.5, *Linum* at 10.0, and *Trigonella* at 7.5 dS/m. Fresh weight of roots also dwindled significantly especially at higher salt treatments in all the test species except *Trigonella* at 7.5 and 12.5 dS/m. The reduced fresh weight of shoots and roots could be attributed to low absorption of water from the growth medium as a result of physiological drought (Hussain & Ilahi, 1992). The slight increase in fresh weight of shoots and roots at certain salt concentration might be due to the development of succulence (Ilahi & Hussain, 1990). Bharti & Singh (1994) reported declined fresh weight of roots of *Sesamum indicum* under saline conditions. The present findings are also in line with Hajar *et al.*, (1996) who reported decreased fresh weight of shoots and roots in *Nigella sativa* at higher salinity levels. Furthermore, Yagmur *et al.*, (2007) has also reported decreased fresh weight in wheat under saline conditions.

Salinity significantly inhibited the growth and resulted in a decrease of dry weight of both shoots and roots of all the tested plants under high salt concentration which could be attributed to the adverse lowering of osmotic potential. The present findings agree with those of Hajar *et al.*, (1996) and Mamo *et al.*, (1996) who reported decreased dry weight of shoots and roots under increasing salinity levels in *Nigella sativa* and chickpea, respectively. Significantly reduced shoot dry weight have also been reported in cowpea (Murillo-Amador & Trovo-Dieguez, 2000), *Glycine max* (Essa, 2002) and *Lens culinaris* (Turan *et al.*, 2007). Moreover, Iqbal *et al.*, (2005), Sadat-Noori (2006) and Yagmur *et al.*, (2007) recorded reduced dry matter in wheat under increasing salt stress, which also strengthen our findings.

Moisture content plays an important role in the growth and development of plants. The present study shows that shoot moisture contents of all the tested species enhanced with increasing salinity especially at higher levels. Root moisture contents in *Linum* and *Plantago* were significantly greater than *Lepidium*, *Nigella* and *Trigonella*. In the present study there was a non-significant increase in shoot moisture contents while, root moisture contents decreased significantly with increasing level of salinity which is in contrast to Ibrar & Hussain (2003) who reported enhanced root moisture contents with increasing level of salinity in *Medicago polymorpha*.

The tested species exhibited reasonably good survival percentage up to 10.0 dS/m application of salt. The survival of seedlings declined variously at high salt application. The observed resistance order regarding survival could be Lepidium (95%) > Plantago (90%) > Trigonella (80%) > Nigella (75%) > Linum (67%). On the basis of survival rate Linum and Nigella could be regarded as moderately tolerant while Lepidium, Plantago and Trigonella could be ranked as highly tolerant to salinity. The findings of the present study agree with those of El-Nakhlaway & El-Fawal (1989) who reported that Linum usitatissimum was least tolerant to different salinity levels. Ashraf & Fatima (1994) and Ashraf & Tufail (1995) screened linseed and sunflower accessions respectively for salt tolerance and established a great deal of variation for salt tolerance in these species. High or low salt tolerance at germination or vegetative stage in various other medicinal plants like Cajnus cajan (Ashraf, 1994), Nigella sativa (Hajar et al., 1996), Cicer arietinum (Mamo et al., 1996), Brasica sp.(Ashraf & Naqvi, 1997), Glycine max (Essa, 2002), Helianthus annuus (Ashraf et al., 2003), Medicago polymorpha (Ibrar & Hussain, 2003), Ricinus communis (Raghavaiah et al., 2002; 2006), Arachis hypogaea (Mensah et al., 2006), Lens culinaris (Turan et al., 2007) and canola cultivers (Jamagard et al., 2008) have also been reported at different degree of salinization.

The results indicate the existence of genetic potential for salt tolerance in these medicinal species under field conditions and they are tolerant up to 10 dS/m salinity level during the vegetative stage of growth. The findings suggest that the response of these species to salt stress depends on the species and concentration of the applied salt. Focusing at the survival percentage, growth and biomass production of the test species, it is suggested that the tested plants could be tried on moderately saline habitat. It will not only help in the utilization of the unproductive saline habitats but also provide raw matter for medicinal concerns. Salt tolerant legumes like *Trigonella* can also improve soil fertility through biological nitrogen fixation.

References

- Ali, R.M., Z.A. Ahmed and R. Abdel-Basset. 1992. Effect of NaCl salinity and Na-Ca combination on seed germination and endogenous levels of some metabolites and respiration on *Trigonella foenum-graecum* L. radicles. *Bull. Faculty Agric. Univ. Cairo*, 43(2): 817-828.
- Anonymous. 2002. *Agricultural Statistics of Pakistan*. Government of Pakistan, Ministry of Food, Agriculture and Live Stock, Economic Wing, Islamabad, 83-84.
- Ashraf, M. 1993. Responses of some local/exotic accessions of Lentil (*Lens culinaris* Medic.) to salt stress. *Journal of Agronomy and Crop Science*, 170(2): 135-147.
- Ashraf, M. 1994. Salt tolerance of pigeon pea (*Cajanus cajan* (L.) Millsp.) at three growth stages. *Annals of Applied Biology*, 124(1): 153-164.
- Ashraf, M. and H. Fatima. 1994. Intra-specific variation for salt tolerance in linseed (*Linum usitatissimum L.*). Journal of Agronomy & Crop Science, 173(3-4): 193-203.
- Ashraf, M. and M. Tufail. 1995. Variation in salinity tolerance in sunflower (*Helianthus annuus* L.). *Journal of Agronomy & Crop Science*, 174(5): 351-362.
- Ashraf, M. and M.I. Naqvi. 1997. Growth and ion uptake of four *Brassica* species as affected by Na/Ca ratio in saline sand culture. *Zeitschrift für Pflanzenernährung und Bodenkunde*, 155(2): 101-108.
- Ashraf, M., R. Zafar and M.Y. Ashraf. 2003. Time-course changes in the inorganic and organic components of germinating sunflower achenes under salt (NaCl) stress. *Flora - Morphology, Distribution, Functional Ecology of Plants*, 198(1): 26-36.
- Beke, G.J. and D.P. Graham. 1995. Growth and chemical composition of flax cultivars on artificially salinized soil. *Can. J. Pl. Sci.*, 75(1): 159-162.
- Bharti, N. and R.P. Singh. 1994. Antagonistic effect of Sodium chloride to differential heavy metal toxicity regarding biomass accumulation and nitrate assimilation in *Sesamum indicum* seedlings. *Phytochemistry*, 35(5): 1157-1161.
- Bos, M. 1992. Salinity and *Plantago*. In: *Plantago*: a multidisciplinary Study. (Ed.): P.J.C. Kuiper and P.J.C. Kuiper. *Ecological Studies*, 89: 148-157.
- El-Davier, S.M. and R.S. Yousaf. 2000. Effect of soil type, salinity, and allelochemicals on germination and seedling growth of a medicinal plant *Lepidium sativum* L. *Annals of Applied Biology*, 136(3): 273-279.
- El-Nakhlaway, F.S. and M.A. El-Fawal. 1989. Tolerance of five oil crops to salinity and temperature stresses during germination. *Acta Agronomica Hungarica*, 38(1-2): 59-65.
- Essa, T. A. 2002. Effect of salinity stress on growth and nutrient composition of three soybean (*Glycine max* (L.) Merrill) cultivars. *Journal of Agronomy & Crop Science*, 188(2): 86-93.
- Farhoudi, R. and F. Sharifzadeh. 2006. The effect of NaCl priming on salt tolerance in canola (*Brassica napus* L.) seedlings grown under saline conditions. *Indian J. Crop Science*, 1(1-2): 74-78.
- Gul, H. and R. Ahmad. 2007. Effect of different sowing dates on the vegetative and reproductive growth of canola (*Brassica napus* L.) cultivers under different salinity levels. *Pak. J. Bot.*, 39(4): 1161-1172.

314

- Hajar, A.S., M.A Zidan and H.S. Al-Zahrani. 1996. Effect of salinity stress on the germination, growth and some physiological activities of black cumin (*Nigella sativa L.*). Arab Gulf Journal of Scientific Research, 14(2): 445-454.
- Huang, J. and R.E. Redmann. 1995. Physiological responses of canola wild mustard to salinity and contrasting calcium supply. *J. of Plant Nutri.*, 18: 1931-1949.
- Hussain, F. 1989. *Field and Laboratory Manual of Plant Ecology*. University Grants Commission, Islamabad.
- Hussain, F. and I. Ilahi. 1992. Effect of magnessium sulphate, sodium sulphate and mixture of both salts on germination and seedling growth of three cultivars of *Brassica campestris* L. Sarhad J. Agric., 3(2): 175-183.
- Ibrar, M. and F. Hussain. 2003. The effect of salinity on the growth of *Medicago polymorpha* Linn. *J. Sci. & Tech. Univ. Peshawar*, 27: 35-38.
- Ilahi, I. and F. Hussain. 1990. Effect of Sodium chloride and Magnessium chloride on germination, growth performance and oil contents of three cultivars of *Brassica campestris*. Proceedings International Conference on Current Developments in Salinity and Drought Tolerance of Plants. January, 7-11, Tandojam, Sindh.
- Iqbal, R.M. 2005. Effect of different salinity levels on partitioning of leaf area and dry matter in wheat. *Asian journal of Plant Sciences*, 4(3): 244-248.
- Janagard, M.S., A. Tobeh and B. Esmailpour. 2008. Evaluation of salinity tolerance of three canola cultivars at germination and early seedling growth stage. *Journal of Food, Agriculture & Environment*, 6(2): 272-275.
- Kheir, N.F., E.Z. Harb, H.A. Mousi and S.H. El-Gayar. 1991. Effects of salinity and fertilization on flax plants (*Linum usitatissimum L.*). I. Growth, yield and technical properties of fibers. *Bull. Faculty Agric.*, Univ. Cairo, 42(1): 39-55.
- Krieg, D.R. 1983. Photosynthetic activity during stress. Agricultural water Management, 7: 249-263.
- Letchamo, W., H.C. Xu, B. Desroches and A. Gosselin. 1993. Effect of nutrient solution concentration on photosynthesis, growth, and content of the active substances of Passion fruit. *J. Pl. Nutrition*, 16(12): 2521-2537.
- Mamo, T., C. Richter and B. Heiligtag. 1996. Salinity effects on the growth and ion contents of some chickpea (*Cicer arietinum* L.) and lentil (*Lens culinaris* Medic.) varieties. *Journal of Agronomy & Crop Science*, 176(4): 235-247.
- Mensah, J.K., P.A. Akomeah, B. Ikhajiagbe and E.O. Ekpekurede. 2006. Effects of salinity on germination, growth and yield of five groundnut genotypes. *African Journal of Biotechnology*, 5(20): 1973-1979.
- Mer, R.K., P.K. Prajith, D.H. Pandaya and A.N. Pandey. 2008. Effect of salts on germination of seeds and growth of young plants of *Hordeum vulgare, Triticum aestivum, Cicer arietinum* and *Brassica juncea. Journal of Agronomy & Crop Science*, 185(4): 209-217.
- Muhammad, Z. and F. Hussain. 2009. Effect of NaCl salinity on the germination and seedling growth of some medicinal plants. *Pak. J. Bot.*, 41(6): (In press).
- Murillo-Amador, B. and E. Trovo-Dieguez. 2000. Effects of salinity on the germination and seedling characteristics of cowpea [Vigna unguiculata (L.) Walp.]. Australian J. Exp. Agric., 40(3): 433-438.
- Mutlu, F. and S. Buzcuk. 2007. Salinity induced changes of free and bound polyamine levels in sunflower (*Helianthus annuus* L.) roots differing in salt tolerance. *Pak. J. Bot.*, 39(4): 1097-1102.
- Narash, R.K., S.K. Minhas, A.K. Goyal, C.P.S. Chauhan and R.K. Guptha. 1993. Production potential of cyclic irrigation and mixing of saline and canal water in Indian mustard (*Brassica juncea*) and pearl millet (*Pennisetum typhoides*) rotation. Arid Soil Res. & Rehabilitation, 7(2): 103-111.
- Quisenberry, J.E. 1982. Breeding for drought resistance and plant water use efficiency. In: *Breeding Crops for Less Favourable Environments*. (Eds.): M.N. Christiansen and C.F. Lewis. PP. 193-212. Wiley, New York.

- Raghavaiah, C.V., Y. Muralidharudu, T.J.J. Royal, P. Ammaji, C. Lavanya and P. Lakshmamma. 2002. Influence of salinity stress on germination and early growth of castor (*Ricinus communis*) genotypes. *Indian Journal of Agricultural Sciences*, 72(10): 601-603.
- Raghavaiah, C.V., C. Lavanya, S. Kumaran and T.J. Royal. 2006. Screening castor (*Ricinus communis*) genotypes for salinity tolerance in terms of germination, growth and plant ion composition. *Indian Journal of Agricultural Sciences*, 76(3): 196-199.
- Sadat-Noori, S.A., S. Mottaghi and O. Lotfifar. 2008. Salinity tolerance of maize in embryo and adult stage. American-Eurasian Journal of Agricultural & Environmental Sciences, 3(5): 717-725.
- Saqib, M. and R.H. Qureshi. 1998. Combined effect of salinity and hypoxia on growth, ionic composition and yield of wheat line 234-1. *Pak. J. Biol. Sci.*, 1(3): 167-169.
- Singh, A.K. and B.B. Singh. 1991. Genotypic variability in biomass protein and nucleic acids in four *Linum* species to sodicity stress. *J. Agronomy & Crop Sci.*, 167(1): 1-7.
- Singh, K.N., D.K. Sharma and R.K. Chiller. 1988. Growth, yield and chemical composition of different oil seed crops as influenced by sodicity. *J. Agric. Sci.*, 111(3): 459-463.
- Steel, R.G.D. and J.H. Torrie. 1980. *Principles and Procedures of Statistics: A Biometric Approach.* 2nd Ed., McGraw-Hill Book, Co. Inc. New York.
- Tanczos, O.G., P.R. Van, Hasselt, P.R. Van-Hasselt, P.J.C. Kuiper and M. Bos. 1992. Low temperature tolerance of *Plantago coronopus* and *Plantago maritima* as affected by salt (NaCl). In: *Plantago:* a multidisciplinary Study. (Ed.) P.J.C. Kuiper. Pub. Springer Verlag Berlin, Germany, pp. 157-161.
- Turan, M.A., N. Turkmen and N. Taban. 2007. Effect of NaCl on stomatal resistance and proline, chlorophyll, Na, Cl and K concentrations of Lentil plants. *Journal of Agronomy*, 6(2): 378-381.
- Ulfat, M., H. Athar, M. Ashraf, N.A. Akram and A. Jamil. 2007. Appraisal of physiological and biochemical selection criteria for evaluation of salt tolerance in canola (*Brassica napus* L.) *Pak. J. Bot.*, 39(5): 1593-1608.
- Umezawa, T., K. Shimizu, M. Kato and T. Ueda. 2000. Enhancement of salt tolerance in soybean with NaCl pretreatment. *Physiol. Plant.*, 110: 59-63.
- Yagmur, M., D. Kaydan and N. Orkut. 2007. Alleviation of salinity stress during seed germination in wheat (*Triticum aestivum*) by potassium applications. *Indian Journal of Agricultural Sciences*, 77(6): 379-382.

(Received for publication 7 November 2008)