RE-USE OF FUNGAL MYCELIUM FOR THE PRODUCTION OF CITRIC ACID BY ASPERGILLUS NIGER

IKRAM-UL-HAQ, SIKANDER ALI AND ASAD-UR-REHMAN

Biotechnology Research Laboratories, Department of Botany, Government College University, Lahore, Pakistan.

Abstract

The present study deals with the re-use of fungal mycelium for citric acid fermentation. Among the 16 – different isolates of *Aspergillus niger*, IS-6 was optimised for citric acid production. The re-use of mycelium was found to be economical due to decrease in fermentation period and high volumetric productivity ($Yp/x = 3.233 \& 2.914 \text{ gg}^{-1}$) but in the third batch, the production of citric acid was markedly decreased due to the mycelial age factor.

Introduction

Citric acid has a wide range of applications in food, pharmaceutical and beverage industries. The entire worldwide demand for citric acid is met by fermentative production, mainly by the moulds involving the filamentous fungus *Aspergillus niger* (Usami & Fukutomi, 1977; Steinboch *et al.*, 1991; Suzuki *et al.*, 1996). Authors have used sucrose salt medium (Begum *et al.*, 1990; Singh *et al.*, 1998) and raw materials such as cane and beet molasses (Mattey 1992; Maddox & Brooks, 1995; Haq *et al.*, 2000) as well as starch hydrolysate (Takatomi & Usami, 1960; Crueger & Crueger, 1984) for citric acid fermentation. The re-use of fungal mycelia may also increase citric acid production, regarding the consistency of fermentative product (Kirimura *et al.*, 1988; Suzuki *et al.*, 1996). The present report describes the production of citric acid by re-using the fungal mycelium, from synthetic medium like sucrose salt medium in shake flask.

Materials and Methods

Sixteen different cultures of *Aspergillus niger* were isolated from soil samples of Lahore, by serial dilution method (Clark *et al.*, 1958). These cultures were maintained on potato dextrose agar (PDA) slants, incubated at 30°C and stored at 4°C in a refrigerator. The conidial suspension was prepared in 10 ml of 0.005 % Monoxal O. T. (Dioctyle ester of sodium sulpho succinic acid) from a 3-6 days old slant culture. The inoculum needle was used for breaking clumps of conidia. The fermentation medium contained (%age, w/v); Sucrose 15.0, KH₂PO₄ 0.1, MgSO₄.7H₂O 0.025 and NH₄NO₃ 0.25. The optimal conditions for citric acid fermentation were investigated in 250 ml Erlenmeyer flask containing 25 ml of the medium having pH 3.5. One ml of the conidial suspension (1.2 x 10⁶ conidia) was added to each flask. The flasks were rotated (160 rpm) in the rotary incubator shaker (Model: GLSC. OSIR-195-0.004, Pak made) at 30°C for 8 days. All the experiments were run parallel in triplicates.

Sugar was determined colorimetrically by DNS method as reported by Tasun *et al.* (1970) while citric acid monohydrate was estimated by the method of Marrier & Boulet (1958). The colour intensity was determined by photoelectric colorimeter (Model: AE-11B, ERMA, Japan) using green wratten filter of 530 nm. Dry cell mass was determined according to Haq & Daud (1995).

Table 1. Screening of Aspergillus niger isolates for the production of citric acid

using sucrose salt media.

Isolates of	Sugar (g/l)		Dry cell	Citric acid	% age
Aspergillus	Used	Residual	mass	monohydrate	citric acid*
niger			(g/l)	(g/l)	
IS-1	66.5	83.5	8.5	4.56	6.86
IS - 2	81.0	69.0	11.0	9.21	11.37
IS - 3	68.0	82.0	9.0	6.55	9.63
IS - 4	57.0	93.0	6.9	7.62	13.37
IS-5	75.0	75.0	10.5	12.50	16.67
IS-6	87.5	62.5	13.5	36.45	41.66
IS-7	90.0	60.0	18.6	26.52	29.47
IS - 8	72.5	77.5	12.5	11.27	15.54
IS - 9	62.0	88.0	8.8	6.72	10.84
IS - 10	52.8	97.2	7.5	5.42	10.26
IS - 11	60.5	89.5	11.5	14.21	23.49
IS - 12	55.5	94.5	12.0	19.12	34.45
1S - 13	83.0	67.0	19.5	20.25	24.40
IS – 14	64.0	86.0	14.2	23.56	36.81
IS - 15	50.5	99.5	9.0	3.12	6.18
IS - 16	71.0	79.0	13.0	25.76	36.28

Sugar added = 150 g/l

Incubation temperature = 30°C

Initial pH = 3.5

Fermentation period = 8 days

Agitation rate = 160 rpm

Results and Discussion

Citric acid is one of the most important organic acids produced commercially by fermentation with specific moulds, mostly strains of *Aspergillus niger* (Mattey, 1992). The present investigation deals with the isolation of *A. niger* strains and then re-use of the fungal mycelium for hyper production of citric acid in shake flask, using sucrose salt media. Among the 16 different isolates of *A. niger* screened for citric acid production (Table 1), isolate IS-6 gave maximum production of citric acid i.e., 36.45 g/l (41.66 % on the basis of sugar used). The sugar consumption was 87.5 g/l while the mycelium were round pellets having dry cell mass 13.5 g/l. Rest of the *A. niger* strains produced relatively lower yields of citric acid monohydrate. This highlighted the idea that it is the type of the strain, which is responsible for high degree product formation. Similar observation has been reported by Maddox & Brooks (1995).

To study the effect of re-use of fungal mycelia on the production of citric acid by *A. niger* IS-6, the mould mycelium after fermentation was separated from fermented broth under aseptic conditions, using sterile centrifuge tubes. The mycelium thus obtained from previous batch was transferred to fresh sterile medium contained in shake flasks. Each fermentation was run for 4-8 days. The experiments were performed in triplicates. In the first batch, the maximum amount of citric acid produced was 48.50±0.12 g/l and glucose consumed was noted as 72.0 g/l (Table 2). The dry cell mass, citric acid production on the basis of sugar used and kinetic parameter, Yp/x values were 15.0 g/l, 67.36 % and

^{*}On the basis of sugar used.

3.233 gg⁻¹, respectively. Further use of the mould mycelium resulted in lowering the consumption of glucose and subsequently the production of citric acid as 88.5 g/l and 52.45 ± 0.34 g/l, respectively (59.26 % citric acid on the basis of sugar used having Yp/x =2.914 gg⁻¹). The re-use of mycelium for the next batch, greatly reduced both glucose consumption and citric acid excretion (second and third batch). But the incubation period decreased from 8 to 6 days (first to second batch) and then only 4 days in the third repeated batch. The decreased incubation period and a fairly high yield of citric acid monohydrate makes the process economical. While in the third fermentative batch, the formation of the product was totally unsatisfactory (12.16 - 18.25 g/l citric acid, 4-5 days after incubation). This might be due to the fact that with the increase in age of mycelium, the metabolic pathways loose their efficiency to secrete the enzyme citrate synthase and also the substrate (sugar) consumption capability, resulting in the lower product formation and finally reaching to about nill in the fourth batch (The data is not given). In a similar study Kirimura et al., (1988) obtained consistent yields of citric acid production, many folds than the first batch. Haq et al., (2000) got a high yield of citric acid in fedbatch culturing of A. niger fermentatation in SS-stirred fermentor.

The present study could suggest that the re-use of fungal mycelia in citric acid fermentation is economical due to consistent yield of the product and decrease in incubation period but one has to find out a potent strain for all this.

Table 2. Effect of Re-use of fungus mycelium on the production of citric acid by

Mycelia	Sugar (g/I)		Incubatio	Ulus niger 1 Dry cell	Citric acid	% age citric	Yp/x***
(g/I)	Use d	Residual	n period (Days)	mass (g/l)	monohydrate (g/l)	acid**	(gg^{-1})
FIRST BATCH		7107					
5.0	64.0	86.0	8	9.0	16.24±0.20*	25.38	1.804
7.5	69.8	80.2	8	10.3	25.55±0.16	36.60	2.480
10.0	73.5	76.5	8	16.5	34.62±0.08	47.10	2.098
12.5	72.0	78.0	8	15.5	48.50±0.12	67.36	3.233
15.0	81.5	68.5	8	17.0	40.26±0.02	49.40	2.368
17.5	88.0	62.0	7	22.5	32.15±0.26	36.53	1.429
20.0	89.5	60.5	7	25.0	29.96±0.29	33.47	1.198
SECOND							
BATCH							
10.0	91.4	58.6	7	16.5	34.56±0.16	37.81	2.094
12.5	88.5	61.5	6	18.0	52.45±0.34	59.26	2.914
15.0	88.0	62.0	6	22.0	39.12±0.31	44.45	1.778
THIRD							
BATCH							
10.0	48.0	102.0	5	13.5	12.16±0.07	25.33	0.901
12.5	67.4	82.6	4	14.0	18.25±0.10	27.08	1.304
15.0	59.5	90.5	4	17.5	18.04±0.23	30.32	1.031

Sugar added = 150 g/l

Incubation temperature = 30°C

Initial pH = 3.5

Agitation rate = 160 rpm

^{*}standard deviation (SD) between the replicates ranging from ± 0.02 to ± 0.34 .

^{**}on the basis of sugar used.

^{***}Product yield coefficient (Yp/x, gg-1) = Citric acid produced. g / Dry cell mass. G

Acknowledgment

This work is a part of the research project No. PSF/GC/Bio/Res(283). Financial assistance provided by the Pakistan Science Foundation, Islamabad is gratefully acknowledged.

References

- Begum, A.A., N. Chaudhary and M.S. Islam. 1990. Citric acid fermentation by gamma ray induced mutants of *Aspergillus niger* in different carbohydrate media. *J. Ferment. & Bioengin.*, 4: 286-288.
- Clark, H.E., P. Bordner, E.F. Geldrich, P.W. Kabler and C.B. Huff. 1958. *Applied Microbiology*. IBS Publishers, N. Y., USA., pp. 27-53.
- Crueger, W. and A. Crueger. 1984. Organic acids. In: *Biotechnol, a textbook of Industrial Microbiology*. (Ed.): T.D. Brock. Sinauer Associates, Sunderland, MA, USA.
- Haq, I., S. Khurshid, S. Ali, H. Ashraf, M.A. Qadeer and M.I. Rajoka. 2000. Mutation of Aspergillus niger for hyper-production of citric acid. World J. Microbiol & Biotechnol., 17(1): 35-37.
- Haq, I., S. Ali and M.A. Qadeer. 2000. Fed-batch culture study during citric acid fermentation by *Aspergillus niger* GCMC-7 in stirred fermentor. *Biologia*, 45(2): 11-17.
- Haq, P.B. and D.A. Daud. 1995. Process of mycelial dry weight calculation for citric acid. J. Biotechnol., 9: 31-35.
- Kirimura, K., I. Nakajima, S.P. Lee, S. Kawabe and S. Usami. 1988. Citric acid production by diploid strains of *Aspergillus niger* obtained by protoplast fusion. *Applied Microbiol*. & *Biotechnol*., 27: 504-506.
- Maddox, I.S. and J.D. Brooks. 1995. Application of a multipacked layer bed reactor to citric acid production in solid-state fermentation using *Aspergillus niger*. *Process Biochem.*, 33(2): 117-123.
- Marrier, J.R. and M. Boulet. 1958. Direct determination of citric acid in milk with an improved pyridine acetic anhydride method. *J. Dairy Sci.*, 41: 1683.
- Mattey, M. 1992. The production of organic acids. Crit. Review Biotechnol., 12: 87-132.
- Singh, S.P., H. Verma, U.N. Kishor and H.K. Samdane. 1998. Effect of concentration of substrate on citric acid production by surface culture fermentation. *Orient J. Chem.*, 14(1): 133-135.
- Steinboch, F.A., Î. Held, S. Choojun, H. Harmsen, M. Rohr, E.M. Kubicek Pranz and C.P. Kubicek. 1991. Regulatory aspects of carbohydrate metabolism to citric acid accumulation by *Aspergillus niger. Acta Biotechnol.*, 11: 571-581.
- Suzuki, A., S. Sarangbin, K. Kirimura and S. Usami. 1996. Direct production of citric acid from starch by a 2-deoxy resistant mutant strain of *Aspergillus niger*. *J. Ferment. & Bioengin.*, 81(4): 320-323.
- Taketomi, N. and S. Usami. 1960. Production of citric acid by submerged fermentation of acid hydrolysate of sweet potato starch. *Kogyo Kogaku Zasshi*, 63: 1602-1605.
- Tasun, K., P. Chose and A. Ghon. 1970. Sugar determination by DNS method. *Biotecnol. & Bioengin.*, 12: 921.
- Usami, S. and N. Fukutomi. 1977. Citric acid production by solid-state fermentation method using sugar-cane bagasse and concentrated liquor of pineapple waste. *Hakko Kogaku*, 55: 44-50.

(Received for publication 12 April 2003)