

A PALYNOLOGICAL STUDY OF THE FAMILY GRAMINEAE FROM KARACHI-PAKISTAN

TAHMEENA SIDDIQUI AND M. QAISER

*Department of Botany,
University of Karachi, Karachi-75270, Pakistan.*

Abstract

A palynological study of 64 species belonging to 40 genera of the family Gramineae from Karachi was conducted. The family is stenopalynous and the pollen are spheroidal, usually monoporate, rarely diporate, smooth, mostly annulate and operculate. On the basis of sculpturing pattern 3 groups are recognized viz., (i) areolate, (ii) scabrate (iii) scabrate-areolate. Each group is further divided on the basis of proximity of the granules (scabrae). Various quantitative characters were also statistically tested.

Introduction

Gramineae one of the largest families of flowering plants, comprising of 620 genera and c. 10000 species is widely dispersed in all the regions of the world (Willis, 1973). In Pakistan, it is represented by 158 genera and 492 species (Stewart, 1972; Cope, 1982). Jafri (1966) reported 43 genera and 72 species from Karachi, while Cope (1982) in the taxonomic revision of the family for the Flora of Pakistan enumerated 48 genera and 86 species from the area.

Grass family is a natural and fairly homogenous taxon. It is remarkable in the constancy of its basic characters. Pollen also do not show much deviation from this contention and has long been recognized as remarkably uniform. Therefore palynology plays a little role in the taxonomy of this family (Wodehouse, 1935; Rowley, 1960). However according to Wodehouse (1935) two characters viz., grain size and sexine pattern are of some significance. Firbas (1937) used the grain size as a basic character to separate wild and cultivated grasses. Rowley (1960) while studying the fine structure of some of the grass pollen used the arrangement of the spinules on the ektexine as a key character for delimiting various taxa.

Faegri & Iversen (1964) in their study of grass pollen, found two different types of sexine i.e., scabrate and areolate. Anderson & Bertelsen (1972) and Grant (1972) also used these two types to distinguish various members of the tribe Triticeae, *Zea* and *Trip-sacum*, respectively. Page (1978) in his scanning electron microscopic survey of grass pollen further divided these two basic types on the basis of the proximity of granules, whether they are closely or widely spaced, while the fused type are differentiated on the basis of height of granules. Chaturvedi (1971) has reported 4 types of grains in Sac-

charum robustum viz., (i) normal monoporate grains (ii) double grains with two pores on either side of the dumbbell shaped pollen grains (iii) single diporate grains and (iv) double grains with single pore. There does not appear to be any report on the palynological aspects of the family Gramineac from Pakistan. A palynological study of the grasses of Karachi was therefore carried out to correlate the various quantitative characters and sculpturing pattern within the different tribes of Gramineae.

Materials and Method

Pollen grains of 64 species belonging to 40 genera were obtained from freshly collected specimens, where fresh material was not available herbarium specimens were used. All the specimens have been deposited in Karachi University Hebarium (KUH). A list of vouchers is given in Appendix I.

Slides of pollen grains were prepared by the classical acetolysis technique devised by Erdtman (1952). Since grass pollen tend to collapse after acetolysis due to the presence of a thin and delicate exine as also found with acetic acid only distilled water was used. Anthers were removed and crushed in a centrifuge tube with distilled water. The material was sieved and centrifuged for 2-3 minutes only, and the water was decanted. For light and scanning microscopy the material was divided into two parts. For SEM, the material was mounted on metallic stubs with a fine jet and coated with gold in a conventional vacuum coater, and examined with a Jeol scanning microscope (JSM-T200). For light microscopic studies, the remaining part of the material was centrifuged for 2-3 minutes in 50% glycerine. After decanting the liquid, the material was mounted in glycerine jelly, prepared according to the method of Kissner (1937).

For light microscopy, 15 pollen grains for each species were observed under compound microscope Erma No. 244825 and the following measurements were taken: (i) longest diameter, (i) Length and breadth of the pore (iii) Exine thickness (iv) Annulus thickness and diameter (v) Scabrate size (vi) Areolae length and breadth (Measurements of iv-vi were taken with SEM). Terminology followed here is that of Kremp (1965), Andersen & Bertelsen (1972), Page (1978) and Solomonon (1983).

Results

The pollen grains in Gramineae are fairly uniform, with an average diameter (26.8-) 18.89-35.57 (-37.18) μm , ± spheroidal, rarely oblate (only in *Cymbopogon martinii*); usually monoporate, but diporate grains may also be present as in *Cenchrus pennisetiformis*; operculate or non-operculate, annulate, annulus 0.83-3.12 μm thick, 3.42-10.0 μm in diameter. Sometimes annulus reduced or even absent. Pore (2.14-) 2.01-3.57 (-4.29) x (0.71-) 0.70-2.47 (-3.57) μm . Exine (0.71-) 0.72-1.43 (2.86) μm thick, usually sexine as

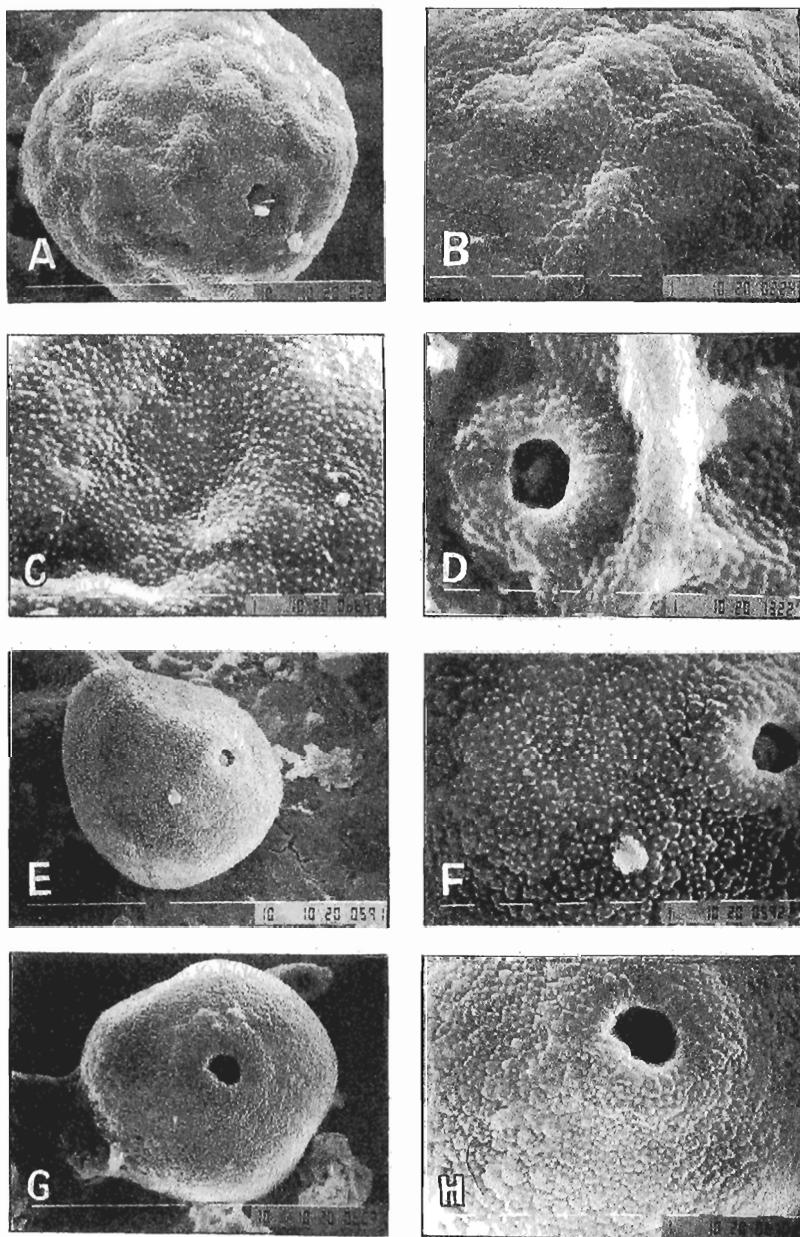


Fig. 1. Scanning electron micrographs of grass pollen. *Cenchrus biflorus*: A. entire grain, B. sculpturing pattern, *Panicum turgidum*: C. sculpturing pattern, *Digitaria violascens*: D. sculpturing pattern, *Panicum antidotale*: E. entire grain, F. sculpturing pattern, *Eragrostis tenella*: G. entire grain, H. sculpturing pattern. (scale: Fig. A, E, G bar = 10 μm and Fig. B, C, D, F, H bar = 1 μm).

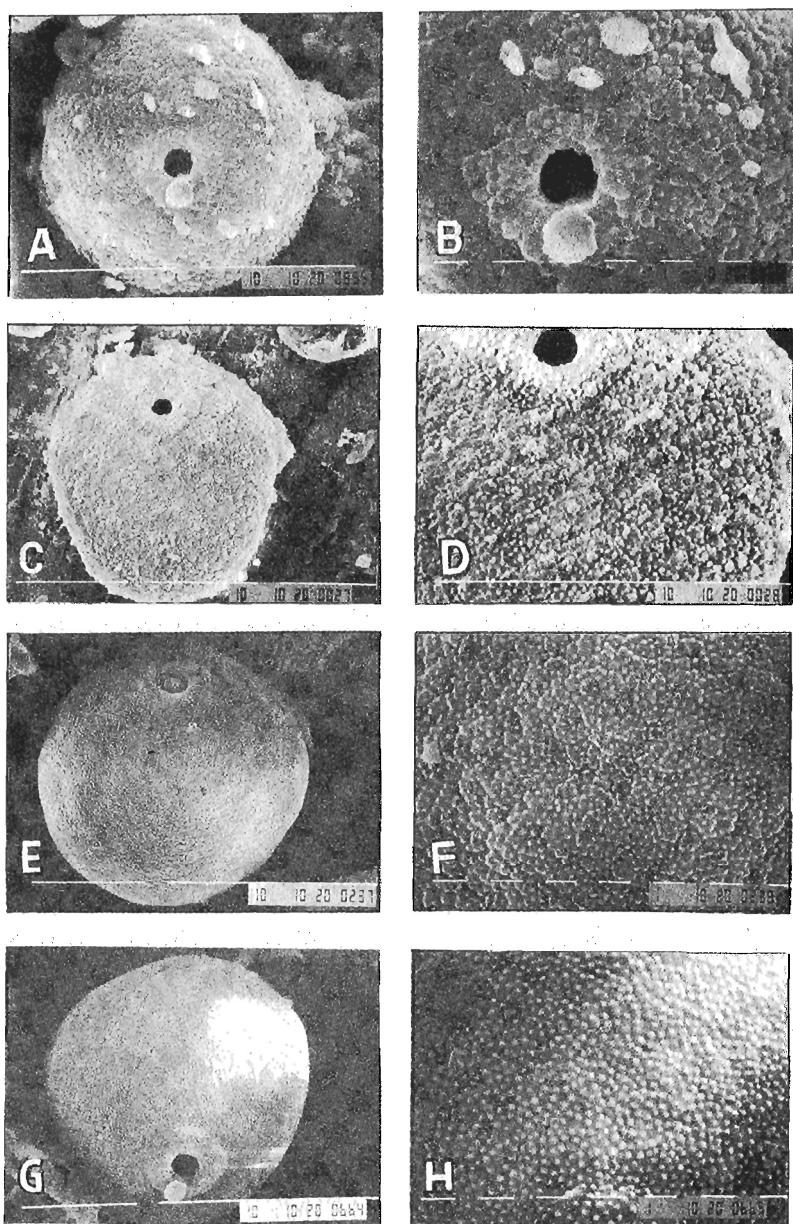


Fig. 2. Scanning electron micrographs of grass pollen. *Eragrostis pilosa*: A. entire grain, B. sculpturing pattern, *Ottochloa compressa*: C. entire grain, D. sculpturing pattern, *Cymbopogon jawarancusa*: E. entire grain, F. sculpturing pattern, *Lasiusurus scindicus*: G. entire grain, H. sculpturing pattern (scale: Fig. A, C, D, E, G bar = 10 μ m and Fig. B, F, H, bar = 1 μ m).

thick as nexine or sexine thicker than nexine or vice versa. Sculpturing pattern areolate, scabrate or scabrate-areolate. Scabrae 0.05-0.20 μm , grouped on small (0.40 x 0.10 - 0.60 x 0.30 μm), medium (0.90 x 0.40 - 1.3 x 0.60 μm) or large (1.5 x 0.80 - 1.7 x 0.10 μm), regularly or irregularly shaped areolae. Incisions present or absent, if present wide or narrow. Granules closely, medium or widely spaced. A detailed description of all the taxa studied is given in Table 1.

Discussion

Gramineae is a stenopalynous family and the pollen show very little character differences, not only at the specific but also at the generic or even at the tribal level. Two main types of the pollen are recognized on the basis of the sculpturing pattern (i) scabrate, (ii) areolate, the third intermediate type i.e. scabrate-areolate have also been observed in few cases. Of the 64 species, 62 wild species showed an areolate pattern, only three species have scabrate, and three have intermediate pattern (Table 1). Present findings are in conformity with Faegri & Iversen (1964), who found that the wild grasses and cereals had predominantly areolate sculpturing. These two basic types have been further subdivided into separate and fused subtypes on the basis of the proximity of the granules (scabrae) both on the smooth surface, as well as on the islands (areolae) as done previously by Andersen & Berltsen (1972) in cereals and wild grasses of tribe Triticeae. The separate subtype can be further classified into closely medium or widely spaced type while in case of fused subtypes the differences seems to be indicated by the height of the granules, which may be in high or low relief. Nevertheless the taxonomic significance of grass pollen is very limited as demonstrated by Page (1978). This is also evident from Table 1, that at the tribe level; on the basis of the granule pattern and sculpturing type; even if arranged in sequence; the tribes form no definite order. All type of granule patterns occur within the same tribe. In tribe Paniceae (12 genera, 24 species), areolate sculpturing was present (Fig. 1, D-F) except in *Panicum turgidum* (Fig. 1, C) and *Paspalidium geminatum* which have scabrate sculpturing and *Cenchrus biflorus* having scabrate-areolate sculpturing (Fig. 1 A&B). All the three types of separate granule pattern i.e. closely, medium and widely spaced type are present within the tribe. Tribe Eragrostideae (7 genera, 12 species) seems to be fairly uniform and have only areolate type (Fig. 1, G-H, 2 A-D), with separate and fused granule pattern. The former is further segregated by having closely and medium spaced type, and the later is with groups in high relief. However, the individual species of this tribe cannot be further delimited on the basis of sculpturing pattern. Tribe Andropogoneae (8 genera 12 species) has predominantly areolate sculpturing (Fig. 2, G&H, Fig 3 A-D), while *Dichanthium annulatum* and *Saccharum griffithii* have scabrate-areolate type. All the further subdivisions of separate type were also met, while the fused type with groups in low relief was also present. In Sporoboleae (2 genera, 4 species; Fig. 3 A&B), Chlorideae (3 genera, 4 species, Fig. 3 C&D), Aveneae (2 genera, 2 species; Fig. 3 G&H), Zoysieae (2 genera, 2 species, Fig. 3 E&F), and Arundineae (2

Table 1. Pollen characteristics of the family Gramineae.

Taxa	Tribe	Size in μm	Op.	Annulus μm		Pore Size μm		Exine thickness μm		BSP	GP
				D	T	Length	Breadth	Mean SE	Mean SE		
<i>Cenchrus pennisetiformis</i>	Paniceae	34.49 \pm 0.41	-	5.5	1.0	3.19 \pm 0.18	2.28 \pm 0.13	1.43	"	Ar.	SCST.
<i>Cenchrus segetius</i>	"	31.55 \pm 0.58	+	5.7	2.34	2.64 \pm 0.06	1.30 \pm 0.15	1.38 \pm 0.03	"	"	"
<i>Cenchrus ciliaris</i>	"	35.87 \pm 0.35	-	4.16	1.38	2.77 \pm 0.07	1.66 \pm 0.11	1.52 \pm 0.06	"	"	"
<i>Setaria pumila</i>	"	24.02 \pm 0.44	-	reduced		2.88 \pm 0.04	0.99 \pm 0.11	1.41 \pm 0.05	"	"	"
<i>Urochloa panicoides</i>	"	32.50 \pm 0.50	+	5.0	2.0	2.86	2.19 \pm 0.16	1.43	"	"	"
<i>Paspalidium punctatum</i>	"	24.59 \pm 0.31	-	reduced		2.31 \pm 0.07	2.33 \pm 0.08	1.43	"	"	"
<i>Digitaria violascens</i>	"	32.21 \pm 0.18	+	6.0	2.0	2.74 \pm 0.05	1.43	1.33 \pm 0.04	"	"	"
<i>Pennisetum glaucum</i>	"	32.98 \pm 0.44	-	5.88	1.47	2.14 \pm 0.03	1.18 \pm 0.09	1.43	"	"	"
<i>Echinochloa colona</i>	"	31.83 \pm 0.22	+	4.5	2.0	2.53 \pm 0.10	1.68 \pm 0.18	1.36 \pm 0.09	"	"	"
<i>Lephiorhynchus senecalese</i>	Zoysieae	22.30 \pm 0.45	-	reduced		2.64 \pm 0.08	0.89 \pm 0.08	1.16 \pm 0.04	"	"	"
<i>Desmostachya bipinnata</i>	Eragrostidaceae	32.31 \pm 0.33	+	3.88	1.10	2.09 \pm 0.08	0.71	1.43	"	"	"
<i>Haldyptium mucronatum</i>	"	30.03 \pm 0.30	-	4.05	1.35	2.86	1.83 \pm 0.18	1.43	"	"	"
<i>Eragrostis tenella</i>	"	19.73 \pm 0.25	-	6.08	1.90	2.95 \pm 0.11	1.20 \pm 0.17	1.33 \pm 0.06	"	"	"
<i>Eragrostis cilianensis</i>	"	24.21 \pm 0.25	+	5.40	1.89	2.71 \pm 0.04	0.19 \pm 0.06	1.43	"	"	"
<i>Eragrostis ciliaris</i>	"	18.78 \pm 0.19	+	reduced		2.81 \pm 0.03	0.84 \pm 0.07	1.43	"	"	"
<i>Dactyloctenium aegyptium</i>	"	31.07 \pm 0.49	+	10	2.0	2.83 \pm 0.23	1.97 \pm 0.16	1.26 \pm 0.07	"	"	"
<i>Dactyloctenium sinadicum</i>	"	26.21 \pm 0.36	-	absent		2.83 \pm 0.23	1.76 \pm 0.11	1.43	"	"	"
<i>Eleusine indica</i>	"	24.97 \pm 0.33	-	reduced		2.29 \pm 0.07	1.52 \pm 0.11	1.43	"	"	"
<i>Cymbopogon martinii</i>	Andropogoneae	35.26 \pm 0.16	+	4.72	1.94	2.69 \pm 0.05	1.60 \pm 0.08	1.43	"	"	"
<i>Saccharum griffithii</i>	"	35.17 \pm 0.19	-	8.22	2.22	3.16 \pm 0.10	2.47 \pm 0.21	1.43	"	"	"
<i>Sorghum halepense</i>	"	30.69 \pm 0.41	-	7.50	2.50	2.50 \pm 0.05	0.84 \pm 0.07	1.43	"	"	"
<i>Dichanthium annulatum</i>	"	29.93 \pm 0.38	+	4.76	1.60	2.76 \pm 0.13	1.95 \pm 0.17	1.43	"	"	"
<i>Dichanthium foveolatum</i>	"	32.80 \pm 0.18	reduced			3.57 \pm 0.25	1.68 \pm 0.27	1.43	"	"	"
<i>Hakea lochiae granularis</i>	"	22.59 \pm 0.15	-	6.75	1.35	2.59 \pm 0.07	0.82 \pm 0.04	1.19 \pm 0.04	"	"	"
<i>Lasiurus scindicus</i>	"	30.03	-	7.02	2.70	2.95 \pm 0.41	1.01 \pm 0.10	1.43	"	"	"
<i>Musotima laevis</i>	"	25.33 \pm 0.31	+	4.83	1.61	2.59 \pm 0.07	0.82 \pm 0.04	1.19 \pm 0.04	"	"	"
<i>Cynodon dactylon</i>	Chlorideae	34.15 \pm 0.27	+	4.91	1.50	2.71 \pm 0.05	1.43	1.43	"	"	"
<i>Phalaris minor</i>	Aveneae	35.08 \pm 0.19	+	5.94	2.70	2.83 \pm 0.02	1.08 \pm 0.19	1.14 \pm 0.03	"	"	"
<i>Arundo donax</i>	Arundineae	24.78 \pm 0.36	-	Absent		2.83 \pm 0.13	1.37 \pm 0.18	1.43	"	"	"
<i>Sporobolus arabicus</i>	Sporoboletae	24.02 \pm 0.39	-	5.40	1.35	2.74 \pm 0.06	1.40 \pm 0.17	1.43	"	"	"
<i>Sporobolus coronandeanthus</i>	"	21.16 \pm 0.39	-	4.0	1.0	2.37 \pm 0.07	0.99 \pm 0.41	1.43	"	"	"

<i>Sporobolus helvolus</i>	"	30.31 ± 0.20	-	5.31	0.85	2.78 ± 0.03	2.78 ± 0.13	1.18 ± 0.13	"	SMST
<i>Urochondra setulosa</i>	"	24.21 ± 0.21	-	reduced	2.86	2.74 ± 0.17	1.43	"	"	"
<i>Setaria intermedia</i>	Panicaceae	25.93 ± 0.16	-	3.52	1.17	2.86	1.97 ± 0.08	1.43	"	"
<i>Setaria verticillata</i>	"	25.54 ± 0.16	-	4.16	0.83	2.34 ± 0.09	1.08 ± 0.50	1.43	"	"
<i>Panicum turgidum</i>	"	34.89 ± 0.27	+	4.05	1.62	2.86	1.97 ± 0.20	1.43	Scab.	"
<i>Panicum antidoteae</i>	"	30.60 ± 0.38	+	4.28	1.42	2.93 ± 0.17	1.42 ± 0.16	1.43	Ar.	"
<i>Cenchrus biflorus</i>	"	24.69 ± 0.18	-	6.0	2.0	2.58 ± 0.51	1.54 ± 0.14	0.34 ± 0.05	Ar.-Scab.	"
<i>Eriochloa falmeensis</i>	"	33.55 ± 0.19	-	6.38	1.94	2.01 ± 0.04	1.50 ± 0.07	1.43	Ar.	"
<i>Echinochloa crus-galli</i>	"	31.26 ± 0.19	+	6.60	1.0	2.86	1.08 ± 0.09	1.28 ± 0.04	"	"
<i>Bracharia eruciformis</i>	"	33.64 ± 0.50	+	5.71	1.42	2.90 ± 0.24	1.11 ± 0.19	1.16 ± 0.05	"	"
<i>Paspalum paspalodes</i>	"	30.98 ± 0.22	-	6.25	3.12	2.95 ± 0.20	2.11 ± 0.18	1.43	"	"
<i>Digitaria nodosa</i>	"	35.87 ± 0.35	+	7.14	1.42	2.90 ± 0.14	1.97 ± 0.20	1.21 ± 0.04	"	"
<i>Digitaria ciliaris</i>	"	26.02 ± 0.28	-	reduced	-	2.95 ± 0.15	1.18 ± 0.19	1.43	"	"
<i>Eragrostis minor</i>	Eragrostideae	26.46 ± 0.40	-	4.16	1.38	1.54 ± 0.08	2.62 ± 0.11	1.43	"	"
<i>Ottochloea compressa</i>	"	25.25 ± 0.16	-	7.29	2.70	2.25 ± 0.09	1.08 ± 0.10	1.43	"	"
<i>Saccharum ravennae</i>	Andropogoneae	28.31 ± 0.28	+	3.42	0.85	2.69 ± 0.03	2.86	1.43	"	"
<i>Chrysopogon aucheri</i>	"	31.93 ± 0.40	+	6.57	2.0	2.54 ± 0.04	1.54 ± 0.16	1.43	"	"
<i>Cymbopogon jwarancusa</i>	"	35.27 ± 0.47	-	reduced	-	2.90 ± 0.04	1.95 ± 0.13	1.43	"	"
<i>Cymbopogon communatus</i>	"	25.45 ± 0.31	-	7.05	2.94	2.74 ± 0.14	1.57 ± 0.11	1.43	Ar.	"
<i>Chloris barbata</i>	Chlorideae	25.54 ± 0.32	-	4.33	1.0	2.59 ± 0.12	1.33 ± 0.13	1.28 ± 0.07	"	"
<i>Phragmites karka</i>	Arundineae	29.36 ± 0.49	-	3.71	2.85	2.78 ± 0.05	2.03 ± 0.18	1.43	Scab.	"
<i>Aeluropoides lagopyoides</i>	Aeluropodeae	33.64 ± 0.30	+	5.0	1.13	2.86	1.50 ± 0.15	1.43	Scab.	SWST
<i>Paspalum geminatum</i>	Panicaceae	27.07 ± 0.24	-	4.25	1.06	2.83 ± 0.02	1.57 ± 0.09	1.43	Ar.	"
<i>Tricholaena teneriffae</i>	"	29.55 ± 0.41	-	4.34	1.09	2.50 ± 0.16	2.23 ± 0.10	1.21 ± 0.04	"	"
<i>Bracharia ramosa</i>	"	24.21 ± 0.88	+	4.86	1.08	2.76 ± 0.06	1.11 ± 0.15	1.43	"	"
<i>Tragus roxburghii</i>	Zoysieae	28.21 ± 0.03	+	5.42	1.07	2.81 ± 0.005	1.54 ± 0.03	1.21 ± 0.008	Ar.-Scab.	"
<i>Aristida mutabilis</i>	Aristideae	25.26 ± 0.22	-	6.12	3.06	2.05 ± 0.16	1.25 ± 0.10	1.20 ± 0.02	Ar.	FGHR
<i>Chloris quinqueveluta</i>	Chlorideae	30.88 ± 0.27	-	4.54	1.18	2.69 ± 0.14	2.26 ± 0.20	1.43	"	"
<i>Tetrapogon tenellus</i>	"	23.64 ± 0.73	-	Reduced	-	2.66 ± 0.09	0.19 ± 0.14	1.43	"	"
<i>Eragrostis pilosa</i>	Eragrostideae	27.55 ± 0.07	+	"	-	2.38 ± 0.09	0.76 ± 0.03	1.47 ± 0.04	FRGR	"
<i>Denebra retroflexa</i>	"	28.31 ± 0.20	+	In the form of ring	-	2.47 ± 0.05	1.28 ± 0.13	1.43	"	"
<i>Polygonum monspeliacum</i>	Aveneae	21.83 ± 0.18	-	7.0	2.2	2.66 ± 0.09	0.19 ± 0.14	1.43	"	"
<i>Eriochloa procera</i>	Panicaceae	23.64 ± 0.73	-	Reduced	-	2.38 ± 0.09	0.76 ± 0.03	1.47 ± 0.04	FRGR	"

Key to abbreviations: Ar. = Areolate, Ar.-Scab. = Areolate-Scabrate, BSP = Basis sculpturing pattern, D = Diameter, FGHR = Fused groups in high relief, FGLR = Fused groups in low relief, GP. = Granule patterning, OP. = Operculum, Scab. = Scabrate, SCST. = Separate closely spaced type, SMST. = Separate medium spaced type, SWST. = Separate widely spaced type, T = Thickness.

genera, 2 species), only areolate sculpturing was observed, while in Sporoboleae, Zoysieae and Chlorideae granule patterning is separate with closely and medium spaced type. In Aveneae separate type with closely and widely spaced subtype is present, while in Arundineae only separate medium spaced type was found (Fig. 4, C&D). In Aeluropodeae and Aristideae (1 genus, 1 species each) sculpturing was scabrate with separate widely spaced granule pattern (Fig. 4 A&B). However, *Eriochloa procera* is unique in having an unsculpturing pattern.

In general, pollen are spheroidal, except in *Cymbopogon martinii* which has oblate grains. Firbas (1937) demonstrated that the pollen of wild grasses measured 20-30 μm and that of cereals were over 35 μm . However, Faegri & Iversen (1975) considered 40 μm as a dividing line between the two. Our findings are similar with Firbas (1937) as the pollen size ranges from (26.18-) 18.89-35.57 (-37.18) μm . In all the tribes studied so far, there seems to be all sorts of overlapping as far as the size of the grains was concerned. However Andropogoneae have the largest grains (29.05-32.49 μm) followed by Paniceae (28.18-32.08 μm), Aveneae (25.75-35.75 μm), Sporoboleae (22.8-34.96 μm), Arundineae (22.88-28.60 μm), Zoysieae (18.50-27.17 μm), Chlorideae (18.59-31.46 μm) and Eragrostideae (14.30-37.18 μm) respectively. Eragrostideae is unique by having the smallest as well as the largest grains. Grains are usually monoporate, however, in few species diporate and double grains with single pore were also observed (Table 2). Nevertheless such grains were larger than the normal monoporate ones, however, the pore size did not differ much, but the position and the interpore distance varied from grain to grain. Erdtman (1952) and Chaturvedi (1971) have already reported 0-7 pores in Triticale and diporate single and double grains in *Saccharum robustum*, respectively. Chaturvedi (1971) attributed the double grains as the non separation of individual grains during microsporogenesis, resulting with pores on the same face, or diporate single grains on the same pole, or double monoporate grains due to polyploidy. Similarly Gould (1957) while working on 9 different species of Andropogoneae, indicated a direct correlation between the chromosome number and pollen size.

The diporate characteristic is not confined to a particular tribe or a genus, they are scattered in several tribes e.g., Paniceae (5 species) Andropogoneae (2 species), Aveneae and Zoysieae (1 species each) have diporate grains (Fig. 4, E-H) which is absent from all the other tribes.

Operculate and non operculate grains were found in all the tribes. However, at the generic level, few genera like *Panicum* (2 species) and *Dichanthium* (2 species) showed exclusively operculate grains, with *Sporobolus* (3 species) and *Saccharum* (2 species) having non operculate grains only. In other genera both operculate and non operculate grains were present.

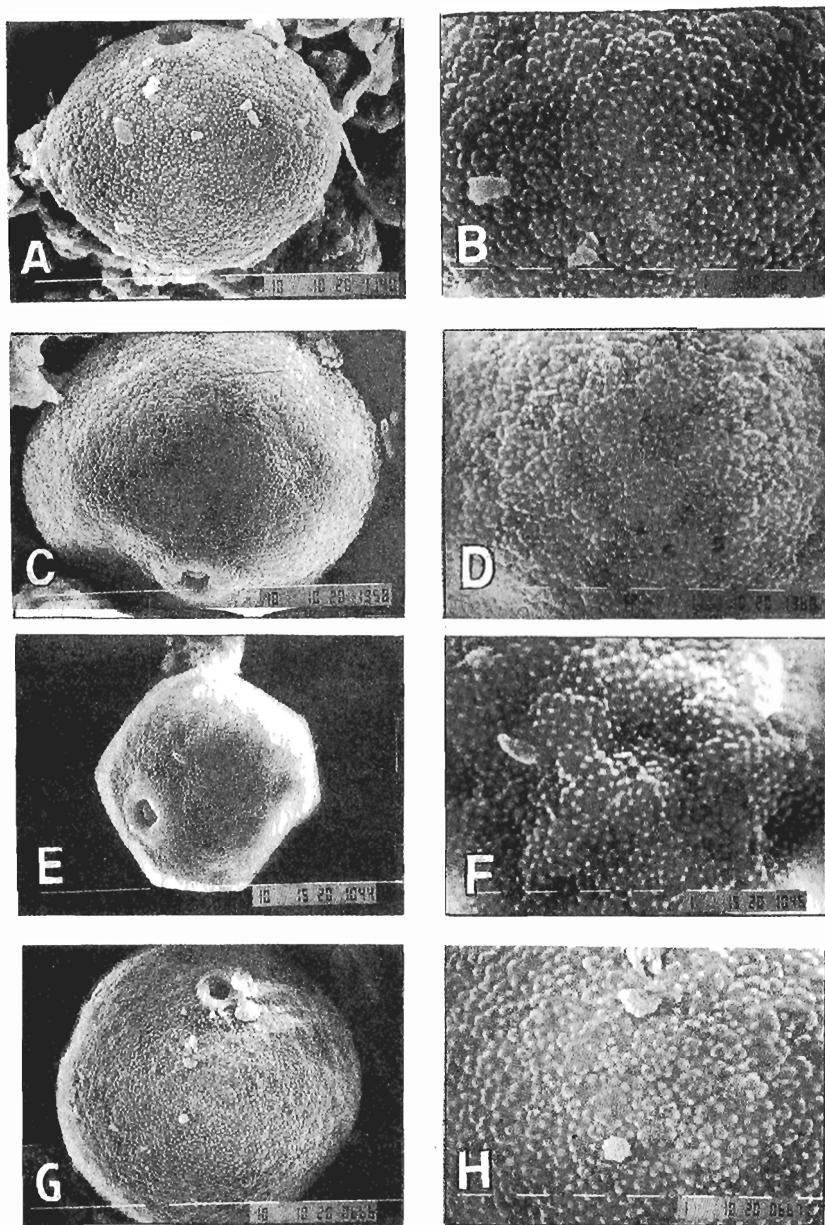


Fig. 3. Scanning electron micrographs of grass pollen. *Sporobolus helvolus*: A. entire grain, B. sculpturing pattern, *Cynodon dactylon*: C. entire grain, D. sculpturing pattern. *Tragus roxburgii*: E. entire grain, F. sculpturing pattern, *Polypogon monspeliensis*: G. entire grain, H. sculpturing pattern. (Scale: Fig. A, C, E, G bar = 10 μ m and Fig. B, D, F, H bar = 1 μ m).

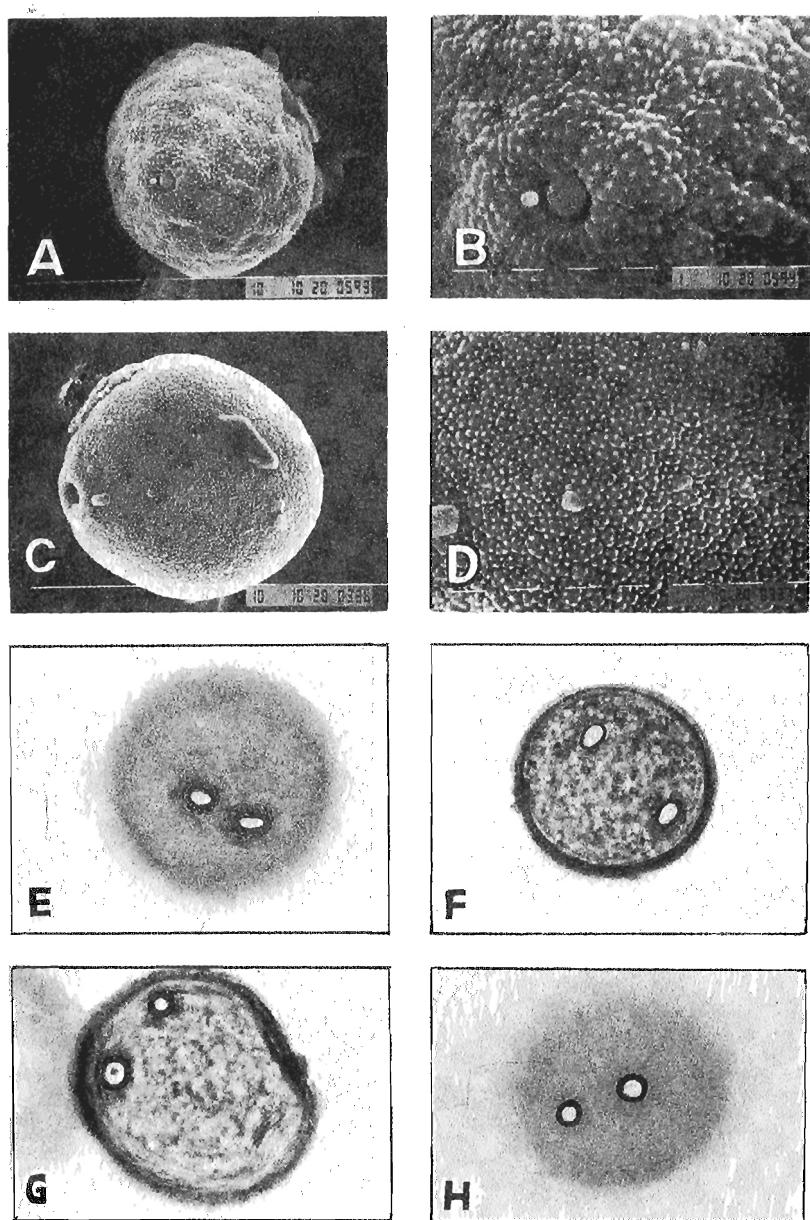


Fig. 4. Scanning electron micrographs of grass pollen. *Aristida mutabilis*: A. entire grain, B. sculpturing pattern, *Phragmites karka*: C. entire grain, D. sculpturing pattern. Light micrographs of diporate grains of grasses. *Digitaria violascens*: E. same face, *Tragus roxburghii*: F. same face, *Polypogon monspeliensis*: G. same pole, H. same face. (Scale: Fig. A, C, bar = 10 μm and Fig. B, D bar = 1 μm Fig. E, F, G, H = 800 x).

Table 2. Some characteristic features of diporate grains.

TAXA	Single grains	Diporate grains	Grain size μm	Interpore distance μm	Position of pores
	Double grains	Single grain	Double grain	Single grain	Double grain
<i>Cenchrus penicilliformis</i>	+	—	31.18 x 37.75	—	2.86 At the same pole
<i>Cenchrus setigerus</i>	—	+	—	25.74 x 24.31	— At the same face
<i>Setaria intermedia</i>	—	+	—	27.17 x 25.70	— At the same face
<i>Digitaria violascens</i>	—	+	—	34.32	" At the same face
<i>Echinochloa colona</i>	+	—	c. 32.89	—	3.57 "
<i>Dactyloctenium sibiricum</i>	—	+	—	30.03	— At the same pole
<i>Sorghum halense</i>	—	+	—	32.69 x 35.75	— At the same pole
<i>Tragus roxburghii</i>	+	—	c. 27.17	—	5.57 "
<i>Polygonum monspelliensis</i>	+	+	c. 28.60	30.03	2.86 At the same pole as well as at the same face.

+ present; - absent

It is interesting to note that not a single tribe can be characterized by the annulus. Three annulus types i.e., (i) well developed, (ii) reduced and (iii) completely absent can be recognized. Few genera like *Cenchrus*, *Digitaria*, *Echinochloa*, *Brachiaria*, *Panicum*, *Saccharum*, *Sporobolus* and *Chloris* have exclusively well developed annulus. In the remaining genera studied so far a mixture of all the three types was present. There is a continuous variation in the thickness (0.83-3.12 μm) and the diameter (3.42-10.0 μm) of the annulus which offers no assistance in the delimitation of the various species. Similarly no correlation exists between the presence of the operculum and the presence or absence of a well developed or reduced annulus, or vice versa. Operculate and non operculate condition may be present in any combinations with the annulus.

Exine thickness ranged from (0.71-) .72-1.43 (-2.86) μm . In most of the species sexine was as thick as nexine, but in a few cases viz., *Pennisetum glaucum*, *Hackelochloa granularis*, *Lasiurus scindicus*, *Phalaris minor*, *Arundo donax*, *Eriochloa fatmensis*, *Sporobolus helvolus*, *Phragmites karka*, *Digitaria nodosa*, *Eragrostis pilosa*, *Dichanthium fo-veolatum*, *Urochloa panicoides* and *Eragrostis ciliaris* sexine was thicker than nexine, while in *Eragrostis ciliaris*, *Digitaria violascens*, *Digitaria ciliaris* and *Panicum antidotale* nexine was thicker than sexine.

References

- Andersen, S.Th. and F. Bertelsen. 1972. Scanning electron microscopic studies of cereals and other grasses. *Grana*, 12: 79-86.
- Chaturvedi, M. 1971. Pollen gains of *Saccharum robustum*. *Grana*, 11: 87-90.
- Cope, T.A. 1982. Poaceae. In: Nasir, E. & Ali, S.I. (Eds.) *Flora of Pakistan*, 143: 1-678, Islamabad.
- Erdtman, G. 1952. Pollen morphology and plant taxonomy – Angiosperms (*An introduction to palynology-I*). Almqvist & Wiksell, Stockholm. Chronica Botanica Co., Waltham, Mass., U.S.A. pp. 538.
- Firbas, F. 1937. Der Pollenanalytische nachweis des Getreidebau. *Zeitchr. Bot.*, 31: 447-478.
- Faegri, K. and J. Iversen. 1964. *Text book of pollen analysis*. Blackwell Scientific Publications, Copenhagen. pp. 295.
- Gould, F.W. 1957. Pollen size as related to polyploidy and speciation in *Andropogon saccharoides* - *A. barbiflora* complex. *Brittonia*, 9: 71-75.
- Grant, C.A. 1972. A scanning electron microscope survey of some Maydeae pollen. *Grana*, 12: 177-184.
- Jafri, S.M.H. 1966. *The Flora of Karachi (Coastal West Pakistan)*. The Book Corporation, Karachi, pp. 375.
- Kremp, G.O. W. 1965. *Encyclopedia of pollen morphology*. Univ. Arizona Press, Tuscon, U.S.A. pp. 263.

- Kisser, J. 1937. Bemerkungen Zum einschluss in Glycerin-Gelatine. *Z. Wiss. Mikr.*, 51.
- Page, J.S. 1978. The scanning electron microscope survey of grass pollen. *Kew Bulletin* 32: 313-319.
- Rowley, J.R. 1960. The exine structure of cereal and wild type grass. *Grana*, 2: 8-15.
- Stewart, R.R. 1972. In: Nasir, E. & Ali, S.I. (Eds.) *Flora of W. Pakistan*. An Annotated Catalogue of Vascular Plants of West Pakistan. Fakhri Press, Karachi. pp. 1028.
- Solomon, A.M. 1983. Pollen morphology and plant taxonomy of white oak in North America. *Am J. Bot.*, 70: 481-494.
- Willis, J.C. 1973. *A dictionary of the flowering plants and ferns*. Cambridge Univ. Press. pp. 1245.
- Wodehouse, R.P. 1935. *Pollen grains, their structure, identification and significance in medicine*. Hafner Publishing Company, New York, pp. 574.

(Received for publication 25 May 1988)

APPENDIX I – LIST OF VOUCHER SPECIMENS

TAXA	LOCALITY	COLLECTOR
<i>Cenchrus penisetiformis</i>	Karachi University Campus, Botany Department.	<i>Tahmeena Siddiqui</i>
<i>Cenchrus setigerus</i>	Safari Park, Karachi.	<i>Tahmeena Siddiqui 45 (KUH)</i>
<i>Cenchrus ciliaris</i>	Cape Monze.	<i>Tahmeena Siddiqui 117 (KUH)</i>
<i>Setaria punnila</i>	Gijju.	<i>Tahmeena Siddiqui 81 (KUH)</i>
<i>Urochloa panicoides</i>	Karachi University Campus, near Girls Hostel.	<i>Tahmeena Siddiqui 137 (KUH)</i>
<i>Paspalidium punctatum</i>	Karachi University Campus, I.B.A. Department.	<i>Abrar Hussain</i> s.n. (KUH)
<i>Digitaria violascens</i>	North Nazimabad, Karachi.	<i>Tahmeena Siddiqui</i>
<i>Pennisetum glaucum</i>	Buffer Zone, Karachi.	<i>Saood Omer s.n. (KUH)</i>
<i>Echinochloa colona</i>	Karachi University Campus, Biochemistry Department.	<i>M. Qaiser s.n. (KUH)</i>
<i>Leptothrix senegalense</i>	Karachi University Camus, near Boys Hostel.	<i>Tahmeena Siddiqui</i>
<i>Desmostachya bipinnata</i>	Thatta Cement Factory.	<i>115 (KUH)</i>
<i>Halopyrum mucronatum</i>	Cape Monze.	<i>Abrar Hussain s.n. (KUH)</i>
<i>Eragrostis tenella</i>	Darsano Chano.	
<i>Eragrostis ciliaris</i>	Karachi Univesity Campus, near Boys Hostel.	<i>Tahmeena Siddiqui 128 (KUH)</i>
<i>Eragrostis ciliaris</i>	Karachi University Campus, Biochemistry Department.	<i>Tahmeena Siddiqui 48 (KUH)</i>
<i>Dactyloctenium aegyptium</i>	Karachi University Campus, near Main Library.	<i>Tahmeena Siddiqui 43 (KUH)</i>
<i>Dactyloctenium scindicum</i>	Ibid.	<i>Tahmeena Siddiqui 73 (KUH)</i>
<i>Eleusine indica</i>	Karachi University Campus, near Main Library.	<i>Abrar Hussain s.n. (KUH)</i>
<i>Panicum antidotale</i>	Karachi University Campus, near I.R. Department.	<i>Tahmeena Siddiqui</i>
<i>Tahmeena Siddiqui 56 (KUH)</i>		
<i>Cenchrus biflorus</i>	Karachi University Campus, Botany Department.	<i>Tahmeena Siddiqui 55 (KUH)</i>
<i>Eriochloa fatmensis</i>	Karachi University Campus, near Boys Hostel.	<i>Tahmeena Siddiqui 150 (KUH)</i>
<i>Echinochloa crus-galli</i>	Thatta Cement Factory.	<i>Tahmeena Siddiqui 111 (KUH)</i>
<i>Eracharia eruciformis</i>	Karachi University Campus, Botany Department.	<i>Tahmeena Siddiqui 118 (KUH)</i>
<i>Paspalum paspalodes</i>	Safari Park, Karachi.	<i>Tahmeena Siddiqui 90 (KUH)</i>
<i>Digitaria nodosa</i>	Karachi University Campus, Zoology Department.	<i>Tahmeena Siddiqui 87 (KUH)</i>
<i>Digitaria ciliaris</i>	Karachi University Campus, I.B.A. Department.	<i>Tahmeena Siddiqui 147 (KUH)</i>
<i>Eragrostis minor</i>	North Nazimabad.	<i>Saood Omer s.n. (KUH)</i>
<i>Octochloa compressa</i>	Karachi University Campus, Botany Department.	<i>Tahmeena Siddiqui</i>
<i>Saccharum ravennae</i>	Safari Park, Karachi.	<i>42 (KUH)</i>
		<i>Tahmeena Siddiqui 164 (KUH)</i>

<i>Chrysopogon aucheri</i>	Karachi University Campus, I.B.A. Department.	<i>Tahmeena Siddiqui</i> 66 (KUH)
<i>Cymbopogon jwarancusa</i>	Cape Monze.	<i>Tahmeena Siddiqui</i> 65 (KUH)
<i>Cymbopogon commutatus</i>	Karachi University Campus, near Girls Hostel.	<i>Abrar Hussain</i> s.n. (KUH)
<i>Chloris barbata</i>	Karachi University Campus, Botany Department.	<i>Tahmeena Siddiqui</i> 50 (KUH)
<i>Phragmites karka</i>	Safari Park, Karachi.	<i>Tahmeena Siddiqui</i> 88 (KUH)
<i>Aeluropus lagopoides</i>	Sandspit, Karachi.	<i>Tahmeena Siddiqui</i> 76 (KUH)
<i>Paspalidium geminatum</i>	Karachi University Campus, I.B.A. Department.	<i>Tahmeena Siddiqui</i> 100 (KUH)
<i>Tricholaena teneriffae</i>	Darsano Chano, Karachi.	<i>Tahmeena Siddiqui</i> 80 (KUH)
<i>Brachiaria ramosa</i>	Karachi University Campus, Botany Department.	<i>Abrar Hussain</i> s.n. (KUH)
<i>Tragus roxburghii</i>	Karachi University Campus, near Boys Hostel.	<i>Tahmeena Siddiqui</i> 125 (KUH)
<i>Cymbopogon martini</i>	Drigh Colony.	<i>Razia Ahmed</i> s.n. (KUH)
<i>Saccharum griffithii</i>	Thatta Cement Factory.	<i>Tahmeena Siddiqui</i> 82 (KUH)
<i>Sorghum halepense</i>	Karachi University Campus, Agricultural Research Institute.	<i>Abrar Hussain</i> s.n. (KUH)
<i>Dichanthium annulatum</i>	Karachi University Campus, Botany Department.	<i>Tahmeena Siddiqui</i> 44 (KUH)
<i>Dichanthium faveolatum</i>	Ibid.	<i>Tahmeena Siddiqui</i> 59 (KUH)
<i>Hackelochloa granularis</i>	Cultivated fields near Pakistan Hotel, Malir.	<i>Abrar Hussain</i> s.n. (KUH)
<i>Lasiurus scindicus</i>	Karachi University Campus, near Girls Hostel.	<i>Surraya Khatoon</i> s.n. (KUH)
<i>Mnesithea laevis</i>	Begar mori.	<i>Tahmeena Siddiqui</i> 165 (KUH)
<i>Cynodon dactylon</i>	Karachi University Campus, Biochemistry Department.	<i>Tahmeena Siddiqui</i> 40 (KUH)
<i>Phalaris minor</i>	Karachi University Campus, Agricultural Research Institute.	<i>Tahmeena Siddiqui</i> 121 (KUH)
<i>Arundo donax</i>	Safari Park, Karachi.	<i>Wajad Ahmed</i> s.n. (KUH)
<i>Sporobolus arabicus</i>	Karachi University Campus, near Girls Hostel.	<i>Tahmeena Siddiqui</i> 67 (KUH)
<i>Sporobolus coromandelianus</i>	Karachi University Campus, near Boys Hostel	<i>Tahmeena Siddiqui</i> 159 (KUH)
<i>Sporobolus helvolus</i>	Karachi University Campus, Staff Town.	<i>Saood Omer</i> s.n. (KUH)
<i>Urochondra setulosa</i>	Cape Monze.	<i>Tahmeena Siddiqui</i> 75 (KUH)
<i>Setaria intermedia</i>	Karachi University Campus, I.B.A. Department.	<i>Tahmeena Siddiqui</i> 116 (KUH)
<i>Setaria verticillata</i>	Karachi University Campus, near Main Library.	<i>Tahmeena Siddiqui</i> 114 (KUH)
<i>Panicum turgidum</i>	2 miles from Hub on way to Sonmiani.	<i>Ali et al</i> 1247 (KUH)
<i>Aristida mutabilis</i>	Karachi University Campus, Arts Faculty.	<i>Ali et al</i> 47 (KUH)
<i>Chloris quinquesetica</i>	Safari Park, Karachi.	<i>Ali et al</i> 167 (KUH)

<i>Tetrapogon tenellus</i>	Karachi University Campus, near Boys Hostel.	<i>Abrar Hussain</i> s.n. (KUH)
<i>Eragrostis pilosa</i>	Karachi University Campus, near Boys Hostel.	<i>Abrar Hussain</i> s.n. (KUH)
<i>Dinebra retroflexa</i>	Darsano Chano.	<i>Tahmeena Siddiqui</i> 84 (KUH)
<i>Polypogon monspeliensis</i>	Karachi University Campus, near Main Library.	<i>Tahmeena Siddiqui</i> 113 (KUH)
<i>Eriochloa procera</i>	North Nazimabad, Karachi.	<i>Saood Omer</i> s.n. (KUH).