

EFFECTS OF SIMULATED SO₂ POLLUTION ON SUBTROPICAL FOREST SUCCESSION: TOWARD CHLOROPHYLL FLUORESCENCE CONCEPT

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Abstract

The effects of simulated SO₂ treatment (NaHSO₃ with different concentrations) on chlorophyll fluorescence in five subtropical forest plants, namely *Pinus massoniana* Lamb, *Schima superba* Gardn., and Champ, *Castanopsis fissa* R and W, *Acmena acuminatissima* Bi.merr., and Perry, and *Cryptocarya concinna* Hance, were investigated. After the leaf sections were immersed in 0, 20, 50, 100 mM NaHSO₃ for 10 h, the low temperature (77 K) chlorophyll fluorescence emission, F_{PSI}/F_{PSII} ratio and chlorophyll fluorescence induction curves of all five woody plants reduced to different extents, while an increase in low temperature fluorescence polarization was found. Short-term treatment of SO₂ damaged PSII, decreased the fluidity of thylakoid membrane, and affected the process of electron transport. Sulfite oxidase activities of five plants grown in three light environments (100, 32, and 12% of natural light intensities) were varied, which showed different resistance to SO₂ pollution. *A. acuminatissima* and *C. concinna*, the dominant species in the late succession stage of a subtropical forest in South China, was less sensitive to SO₂ pollution. And *P. massoniana*, the pioneer heliophyte species was less able to resist NaHSO₃. It is suggested that SO₂ pollution may accelerate the succession of subtropical forest.

Introduction

Sulphur dioxide (SO₂) is a widespread air pollutant, and its environmental effects include acidification of soils, lakes and rivers and damage to plants and crops. The introduction of legislation in recent years, at least in developed world, has led to very substantial reductions in emissions (Mansfield, 1999). However, Emissions of SO₂ in the area with rapidly developing economies of south and East Asia, as well as Africa, South and Central America have been increasing quickly, the southeast Asia now emits more S into the atmosphere than either Europe or North America (Percy & Ferretti, 2004). Acid deposition derived from combustion of fossil fuels may occur close to the point or area source as well as over long distances of 1000 km or more. Therefore, acid pollution becomes the world wide subject. As the world's largest consumer of primary commercial energy and emitter of SO₂, and the second largest producer of hard coal, China faces greater risks of acid pollution than any other Third World country. The emission of SO₂ in China was 22.549 million tons during the year of 2004 (State Environmental Protection Administration of China, 2005).

Chronic SO₂ exposure affects photosynthesis in forest plants, depending on exposure dose and species analyzed. Alterations were found mainly at PSII level in young spruce trees, following SO₂ exposure, particularly affecting the structure of D1 protein (Lütz *et al.*, 1992). Chloroplast is one of the main targets of SO₂ or its degradation products generated in aqueous solution, resulting in an impairment of chloroplast functionality through a loss of net CO₂ assimilation, decline in photosynthetic electron transport rate

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and inhibition of dark reactions of photosynthesis (Veljovic-Jovanovic *et al.*, 1991). SO_2 can react with water to yield bisulfite. Within the chloroplasts, bisulfite will be further converted to sulfite because of the pH value in stroma is at 8.8 (Kurkdjian & Guern, 1989). $\text{SO}_2/\text{HSO}_3^-$ may cause injury in a certain position, restrain Calvin cycle and affect electron transport rate (Veljovic-Jovanovic *et al.*, 1991). Sulfite is known to present some cytotoxic, mutagenic and antinutritional effects (Stammati *et al.*, 1992). Sulfite oxidase (E.C.1.8.3.1) catalyses the final reaction in oxidative degradation of sulfite, and its activities can be used to evaluate the resistance of plants to SO_2 pollution.

Measuring chlorophyll fluorescence is an important technique in investigating energy transportation of photosynthesis. In vivo, chlorophyll fluorescence is almost from chlorophyll a in PSII (including antenna chlorophyll a), which supplies a quick information of utilizing and dissipating excitation energy in PSII. Each step in photosynthesis is connected and can be reflected by PSII fluorescence. Fluorescence emission at 77 K demonstrated the alterations in the overall distribution of excitation energy between PSII and PSI. The balance of absorbed light energy (or excitation energy) between the two photosystems is beneficial to achieve the maximum photochemical energy conversion efficiency. Fluorescence polarization can be used to detect the alterations of biology membrane, thus to evaluate the fluidity of thylakoid membrane responding to environmental signal. Since the early 1970s, the use of fluorescence polarization to investigate membrane fluidity has gained popularity due to its many advantages. Fluorescence polarization is a very sensitive technique, being more informative than differential scanning calorimetry (DSC) and easier to use than electron spin resonance (ESR) (Borenstain & Barenholz, 1993). Alteration of membrane fluidity in vitro modified the response of isolated thylakoid membranes to photoinactivation of PSI and PSII activities at room and low temperature (Velitchkova *et al.*, 2001). The kinetics of fluorescence induction (Wang & Shen, 1989) measured under very strong actinic light is of special interest because the rate of the photochemical reactions is very high under these conditions and the photochemical and non-photochemical events are well resolved (Schreiber, 2002). Therefore, chlorophyll fluorescence is measured in our studies to detect the influences of SO_2 pollution on photosynthetic machinery.

Most data reported in literature concerned fumigations with various concentrations of SO_2 , alone or in combination with other pollutants, either for short (Sandhu *et al.*, 1992; Veljovic-Jovanovic *et al.* 1991) or long periods (Garcia *et al.*, 1998; Ranieri *et al.*, 1999). Whereas, little information is available on the effects of NaHSO_3 permeation on biochemical and physiological responses of forest plants for understanding the influence of the intermediate of SO_2 metabolism on forest ecosystem. And to our knowledge, no report in measuring the activities of sulfite oxidase has been found in analyzing the damage of SO_2 to plants.

South China is located in south subtropical area with rapid industrial development, and it is one of the main acid rain polluted areas. The current paper focuses on the responses of several dominant plant species in forest succession grown under different light intensities to simulated SO_2 treatment, in order to deduce the effects of acid deposition on subtropical forest structures. The differences of physiological responses and protection strategies toward the change in environmental conditions may be of major importance for the overall physiological performance and the contribution to competitive differences among species.

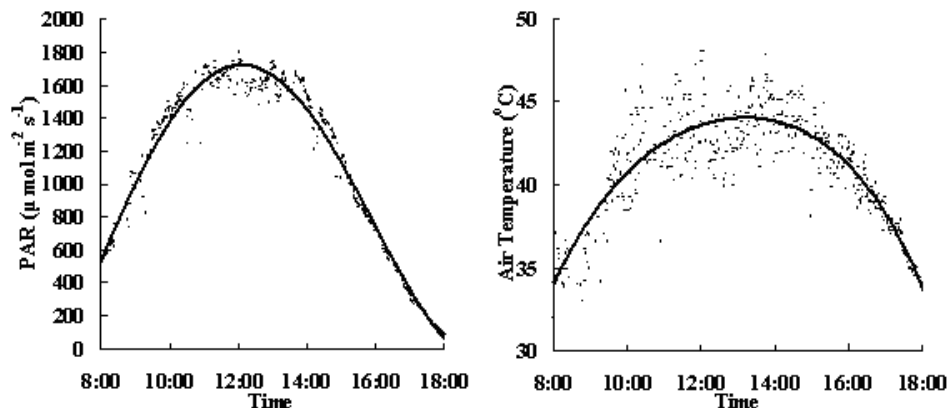


Fig. 1. Diurnal changes of photosynthetically active radiation (PAR) and air temperature in August at experimental site of South China Botanical Garden, Guangzhou, China

Materials and Methods

Field site and plant materials: The experimental site is at South China Botanical Garden, Guangzhou, Guangdong Province, China (23°35' N and 122°57' E), belonging to south subtropical monsoon climate. The annual average temperature is 21.4–21.9°C. The average temperature of July and August, the hottest months of the year, is 28.0–28.7 °C and January or February, the coldest month, with the average temperature of 12.4–13.5 °C. Average precipitation is 1623.6–1899.8 mm and average radiation is 4367.2–4597.3 MJ m⁻² during the whole year. Five typical woody plants, *Pinus massoniana* Lamb, *Schima superba* Gardn. and Champ, *Castanopsis fissa* R and W, *Acmena acuminatissima* Bi.merr. and Perry, and *Cryptocarya concinna* Hance, represent three succession periods, were chosen for studies. *P. massoniana* is the pioneer heliophyte species; *S. superba* and *C. fissa* are medium-succession species, while *A. acuminatissima* and *C. concinna* are climax species in a subtropical wildwood at Dinghu Mountain nature reserve. The potted seedlings were grown under different light intensities (100%, 32% and 12% of natural light), respectively with regular management of water and fertilizer for 21 months. Havelocks were used in setting light gradients and 12 potted plants were put under each light intensity treatment. Diurnal change of natural light intensity and air temperature in August (Fig. 1) were measured using PAR and temperature sensors (PAM-2100, Germany). 100% natural light intensities reached 1801 μmol m⁻² s⁻¹ at midday.

Sampling and simulated SO₂ treatment: Leaves from different individuals (n>3) with similar position on plants were taken for in vitro treatments and measurements. According to the rain acidity of Guangzhou city (Huang *et al.*, 2004), leaf disks (2.0 cm length for *P. massoniana* needles) were immersed, adaxial side up, in 20, 50, 100 mM NaHSO₃ (pH 5.3, 5.0, 4.8, respectively), and 0 mM NaHSO₃ (distilled water, pH 6.0) treatment as control under 20 μmol m⁻² s⁻¹ illumination for 10 h (25 °C).

Measurements of sulfite oxidase activities: According to the method described by Sezintürk & Dinçkaya (2005) with alterations, leaf samples with known weight were

ground in a mortar with addition of phosphate buffer (pH 7.5, 50 mM). The mixture was centrifuged for 15 min (10000×g, 0–4°C). Supernatant with 30 µl (equal to 1.5 mg leaf samples) was added to Na₂SO₃ solution with different concentrations (0, 0.05, 0.1, 0.15, 0.2, 0.3, 0.5, 1 M). Absorptions were measured at 240 nm due to the formation of H₂O₂ with an Ultraviolet-Visible spectrophotometer (Lambda 25, USA). Enzyme activities were expressed as $\Delta A_{240\text{ nm}} \text{ min}^{-1} \text{ mg}^{-1} \text{ FW}$.

Measurements of 77K Chlorophyll fluorescence emission spectra and polarization:

Leaf samples treated after NaHSO₃ or water were washed by distilled water, cut into strips and inserted into quartzose tubes. 77K Chlorophyll fluorescence emission spectra were recorded by a fluorescence spectrophotometer with low temperature affiliation (LS55, Perkin-Elmer Inst. USA). The excitation wavelength was at 436 nm (slit 10 nm) and the emission was between 600 to 800 nm (slit 10 nm). Chlorophyll fluorescence polarization was measured and calculated with the polarizing function of the instrument at 742 nm spontaneously. The fluorescence polarization (P) was calculated as $P = (I_w - GF \cdot I_{vh}) / (I_w + GF \cdot I_{vh})$, where I_w was the intensity with the polarizers vertical and vertical (excitation and emission), I_{vh} was the intensity with the polarizers vertical and horizontal (excitation and emission) and GF is the Grating Factor. The Grating Factor (GF) corrects for instrumental polarization which can be calculated by the ratio of vertically over horizontally polarized light ($GF = I_{hv} / I_{hh}$) when the exciting beam of light was polarized horizontally (Litman & Barenholz, 1982).

Measurements of chlorophyll a fluorescence induction curves: Leaf disks or needles treated after NaHSO₃ or water were washed by distilled water and dark adapted for 30 min. The fast kinetics curves (within 3.5s) were measured by a portable pulse-modulated fluorimeter PAM-2100 (Walz, Germany) with the software of PamWin. Red excitation light peaking at 650 nm (intensity of 600 µmol photons m⁻² s⁻¹) was used for fluorescence induction.

Results

Sulfite oxidase activity: Sulfite oxidase is physiologically important and its absence may even lead to death (Ganai *et al.*, 1997). The enzyme activities ($\Delta A_{240\text{ nm}} \text{ min}^{-1} \text{ mg}^{-1} \text{ FW}$) of the five forest species were increased with the elevation of Na₂SO₃ concentration gradually but became stable after 0.5 M (Fig. 2). The sulfite oxidase activity in the extract of *A. acuminatissima*, a late succession stage species, increased by 1.17–5.79 times over the control (Na₂SO₃- free) after reacting with 0.5 M Na₂SO₃, showing its better capacity in transforming and detoxifying sulfite. However, compared with the broad-leaved species, the pioneer species *P. massoniana* changed least, with 99.95, 85.49 and 87.12% increments at three natural light treatments (100, 32, 12%), respectively. The increments in sulfite oxidase activities of *S. superba*, *C. fissa* and *C. concinna* were in between, with those under 100% natural light changed little. 32% natural light treatment made these three species higher enzyme activities and alterations. Moreover, the increasing rate of sulfite oxidase activity (Na₂SO₃ concentration in the linear part of curve in Fig. 2) revealed that *P. massoniana* under three light intensities had less activity rates than the other four species, while *C. fissa* showed the highest rates. In most cases, the activity rate at the lower Na₂SO₃ concentration was repressed significantly by natural light (Table 1).

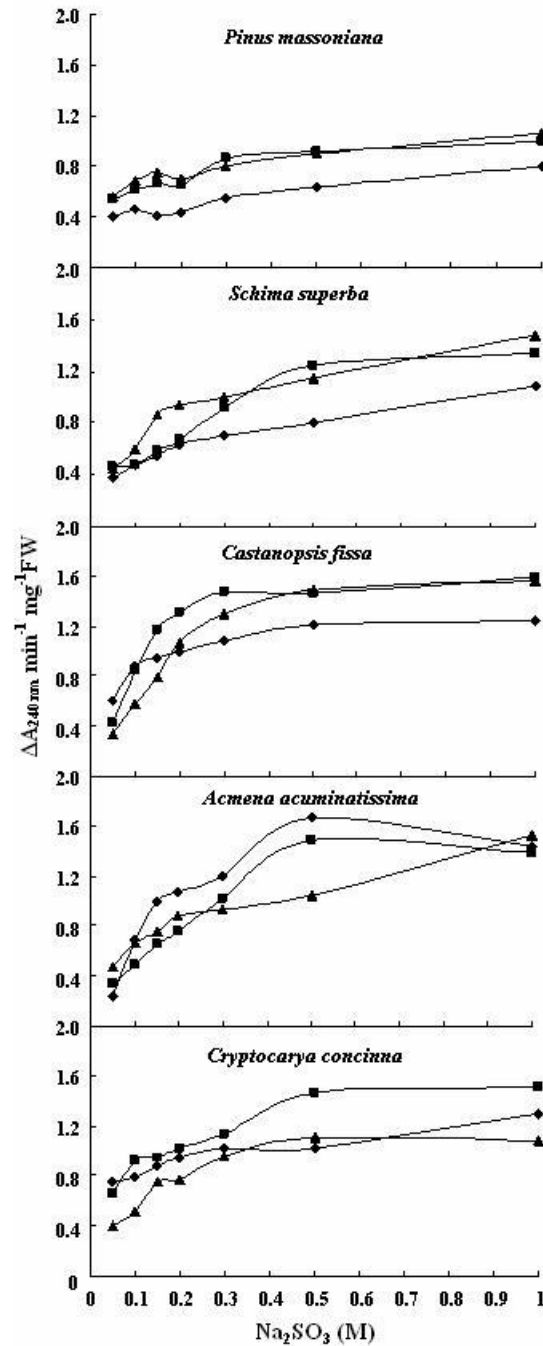


Fig. 2. Changes in sulfite oxidase activities along with Na_2SO_3 concentration in five forest plants *P. massoniana*, *S. superba*, *C. fissa*, *A. acuminatissima* and *C. concinna* by long-term light gradient treatments. (◆- 100% light intensity; ■-32% light intensity; ▲-12% light intensity=4)

Table 1. Increasing rate of sulfite oxidase activities ($\Delta A_{240 \text{ nm}} \text{ min}^{-1} \text{ mg}^{-1} \text{ FW M}^{-1} \text{ Na}_2\text{SO}_3$, the slopes in the linear part of curves in Fig. 2 between 0.05 and 0.2 M Na_2SO_3 , n=4) in five plants *P. massoniana*, *S. superba*, *C. fissa*, *A. acuminatissima* and *C. concinna* by long term light gradient treatments.

Species	Relative light intensity (%)	Na ₂ SO ₃ Concentration (M)				Increasing rate
		0.05	0.10	0.15	0.20	
<i>P. massoniana</i>	100	0.40	0.46	0.42	0.44	0.17a
	32	0.54	0.61	0.66	0.64	0.73b
	12	0.57	0.69	0.75	0.70	0.91c
<i>S. superba</i>	100	0.38	0.48	0.55	0.63	1.64a
	32	0.47	0.47	0.58	0.67	1.44b
	12	0.45	0.60	0.87	0.94	3.52b
<i>C. fissa</i>	100	0.60	0.88	0.95	0.99	2.49a
	32	0.42	0.85	1.16	1.31	5.96b
	12	0.34	0.58	0.79	1.07	4.79c
<i>A. acuminatissima</i>	100	0.24	0.69	0.10	1.06	5.54a
	32	0.34	0.49	0.66	0.75	2.75b
	12	0.48	0.67	0.75	0.89	2.60b
<i>C. concinna</i>	100	0.76	0.80	0.88	0.95	1.28a
	32	0.66	0.92	0.94	1.02	2.20b
	12	0.40	0.51	0.76	0.77	2.71b

*Within the last line, means with the same letter are not significantly different at $P < 0.05$

Low-temperature (77 K) chlorophyll fluorescence: Fluorescence emission at 77 K showed the excitation densities within LHCII, PSII and PSI determine the amplitude of emitted fluorescence at 685 nm, 695 nm (emitted by LHCII-PSII complexes) and 735 nm (emitted by PSI-LHCI) (Krause & Weis, 1991). As shown in Fig. 3, the corresponding fluorescence peaks of PSII and PSI among five species were somewhat different. Two peaks in *P. massoniana* were at 697 and 736 nm, while that in broad-leaved plant *C. fissa* were at 697 and 740 nm, also at 687 and 740 nm in other three species. With the treatment of grown light gradients and consequent NaHSO_3 , the features of low temperature fluorescence emission spectra were altered at different extents. In the control leaves (non NaHSO_3 treatment) of all tested plants, fluorescence emission, especially that from PSI reduced with the decrease in light intensity (Fig. 3 A. B. C, curve 1). NaHSO_3 treatment led to the changes in height and site of fluorescence peak. As the concentration of NaHSO_3 increased, PSI related peak was lowered down and PSII peak went up in the leaves grown at low light (12%) condition. The blue shift and reduction of PSI emission peak were observed in leaves of five species grown under 100% and 32% of natural light and treated with NaHSO_3 . It was exacerbated by the higher concentration of NaHSO_3 (50- 100 mM, Fig. 3, curve3 and 4). However, the change in peaks relative to PSII differed from plant species, which showed decrease, increase or even unchanged. A significant increase of PSII fluorescence emission was found in *P. massoniana* leaves at 100% and 32% light and *A. acuminatissima* at 32% light when treated with 100 mM of NaHSO_3 .

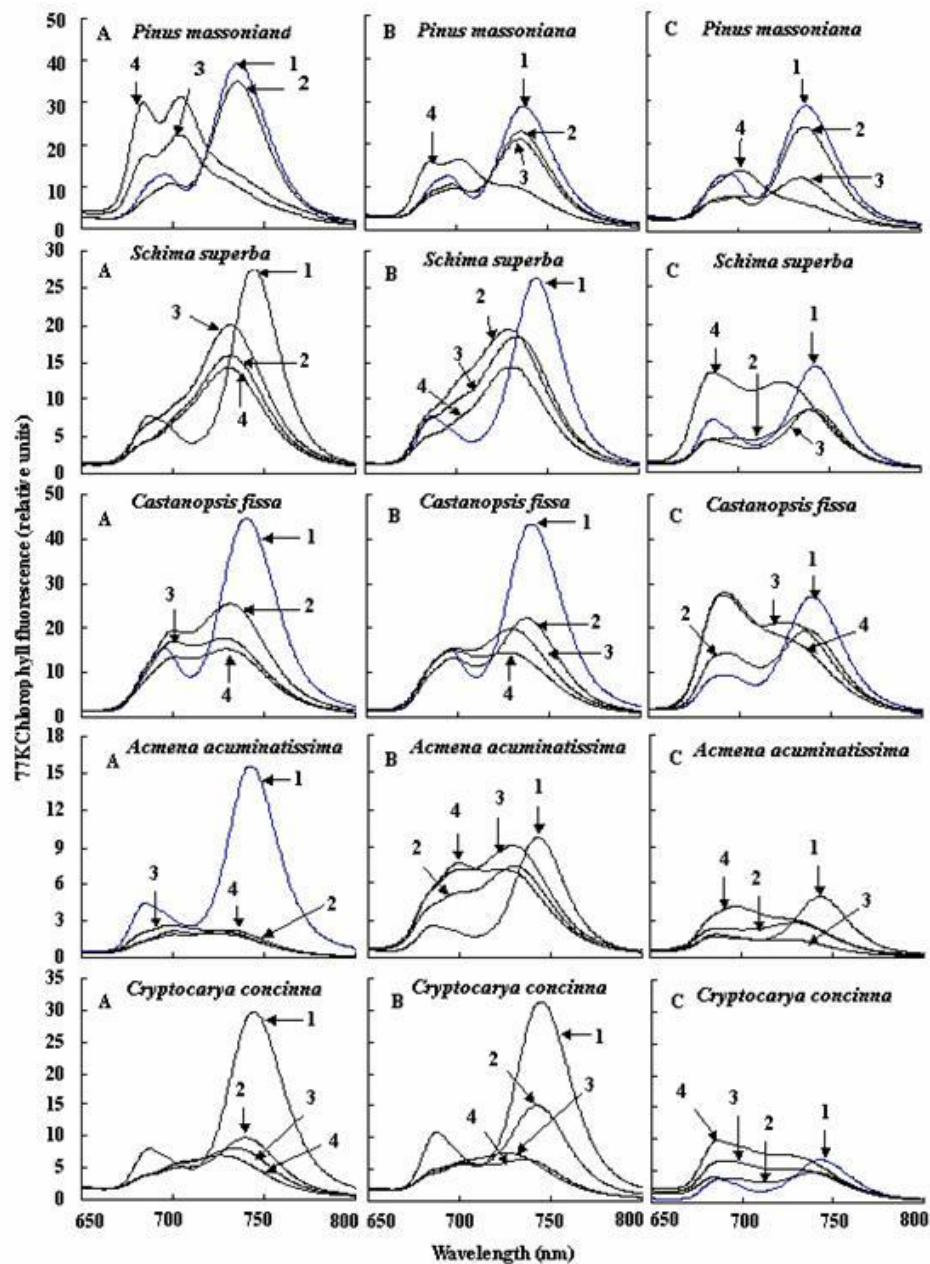


Fig. 3. 77 K fluorescence emission in five plants *P. massoniana*, *S. superba*, *C. fissa*, *A. acuminatissima* and *C. concinna* by long term light gradient and short term NaHSO₃ solution treatments. A-100% light intensity; B-32% light intensity; C-12% light intensity

The number of 1, 2, 3, 4 represents the treatment with 0, 20, 50, 100 mM NaHSO₃, respectively. Each curve was the mean of 3 or 4 individual experiments.

A concomitant redistribution of excitation energy in favor of PSI relative to PSII can be detected as an increase in 77 K fluorescence emission associated with PSI relative to PSII (Bruce *et al.*, 1989). From the ratios of the two peak values (Table. 2), gradient changes of F_{PSI}/F_{PSII} were found according to different natural light treatments except *A. acuminatissima*, which demonstrated that weak light is beneficial to distribute more excitation energy from PSII to PSI, whereas strong light may increase the fraction of excitation energy at PSII but PSI. It is apparent from most of the data in Table 2 that NaHSO_3 treatment resulted in the decrease in F_{PSI}/F_{PSII} ratios regardless of the difference between either species or light intensity. Compared with the other four species, F_{PSI}/F_{PSII} of *P. massoniana* under 12% natural light decreased most (by 84.4%) after immersing in 100 mM NaHSO_3 . On the contrary, changes of *C. concinna* were relative small, with the alteration of 53.6%, 39.4% and 48.2%, under three light intensities, respectively, which showed higher tolerance ability to simulated SO_2 pollution.

Low-temperature (77 K) Chl fluorescence polarization: Membrane fluidity is inversely related to the probe polarization ratio. As membrane fluidity decreases, the polarization ratio increases and vice versa (Litman & Barenholz, 1982). In general, membrane fluidity is detected by the polarization of membrane lipids with an exogenous fluorescence probe such as DPH (1,6-diphenyl-1,3,5-hexatriene). In the present study, the polarization of chlorophyll fluorescence was monitored directly for understanding the perturbation of membrane properties because chlorophylls are bounded with proteins as complexes which locate in thylakoid membrane. Table 3 showed that fluorescence polarization was elevated obviously by the increasing NaHSO_3 concentration, reflecting that thylakoid membrane of these five forest plants were impacted with their fluidity decreasing to different extents. Among the examined plants, the levels of polarization in *P. massoniana* and *S. superba* treated with 100 mM NaHSO_3 increased gradually with decreasing light intensity. A 4.64 and 6.16 times increment of polarization under 12% natural light was found compared with the corresponding controls. However, less change was presented in *C. concinna*, with the polarization increasing of 226.8, 82.5 and 105.6%, respectively. The susceptivities of this parameter in *C. fissa* and *A. acuminatissima* were in between.

Chlorophyll a fluorescence induction curves: The fluorescence induction curves in Fig. 4 were similar to former studies, with O, I, D, P and S states appeared (Widell *et al.*, 1983). After dark adaptation, fluorescence came out immediately with F_0 level (O on the curve) when illuminated by excitation light. Thereafter, increases (or decreases at S state) in most curves were found including I, D, P and S points. Five forest species had similar induction curves in shapes but were different in fluorescence emissions after treatments of light gradients and NaHSO_3 , with those grown under 12% natural light higher than 100% natural light. Fluorescence emission (especially F_0 and F_p) of the other four species declined with the elevated NaHSO_3 concentration gradually except *C. concinna*, which showed less sensitive to SO_2 pollution with crossed curves appeared. On the contrary, the induction curves of *P. massoniana* became smooth with P peaks disappeared after immersed in 100 mM NaHSO_3 . *S. superba* and *A. acuminatissima* grown under 100% natural light shifted similar as *P. massoniana*. Meanwhile, 1- (F_0/F_p) values were calculated in Table 4, which expressed variable fluorescence of plant leaves. Similar with other fluorescence parameters, the lower values of 1- (F_0/F_p) in *P. massoniana* again demonstrated that it responded sharply to elevated concentrations of NaHSO_3 .

Table 2. Changes in F_{PSI}/F_{PSII} at 77 K in five plants *P. massoniana*, *S. superba*, *C. fissa*, *A. acuminatissima* and *C. concinna* by long term light gradient and short term NaHSO₃ solution treatments.

Species	Relative light intensity (%)	NaHSO ₃ concentration (mM)			
		0	20	50	100
<i>P. massoniana</i>	100	2.20a±0.09	3.00a±0.21	2.49a±0.30	0.53b±0.13
	32	2.09a±0.27	1.80a±0.60	2.51a±0.41	0.64b±0.22
	12	3.15a±0.19	3.13a±0.05	0.50b±0.05	0.49b±0.07
<i>S. superba</i>	100	2.06a±0.24	1.95a±0.07	1.83a±0.14	0.90b±0.07
	32	3.40a±0.55	2.54b±0.09	2.12b±0.22	1.70c±0.39
	12	3.75a±0.88	3.24a±0.40	2.08b±0.05	1.45c±0.75
<i>C. fissa</i>	100	2.92a±0.16	1.37b±0.04	0.79c±0.03	0.60c±0.14
	32	2.88a±0.12	1.70b±0.91	1.37b±0.06	1.05b±0.01
	12	2.93a±0.14	1.32b±0.07	1.04b±0.03	1.06b±0.01
<i>A. acuminatissima</i>	100	2.59a±0.20	1.17b±0.19	0.86b±0.07	0.82b±0.08
	32	3.66a±0.20	1.74b±0.15	1.29c±0.13	0.92c±0.07
	12	2.35a±0.17	1.50b±0.01	0.88c±0.03	0.85c±0.02
<i>C. concinna</i>	100	1.83a±0.21	1.23a±0.17	0.92b±0.08	0.85b±0.06
	32	2.82a±0.72	3.01a±0.78	1.74b±0.05	1.71b±0.12
	12	3.40a±0.17	2.44b±0.85	2.25b±0.57	1.76c±0.21

*Within each row, means with the same letter (a for 0 mM NaHSO₃) are not significantly different at $P < 0.05$, $n=3$

Table 3 Changes in chlorophyll fluorescence polarization ($\times 10^{-2}$) at 77 K in five plants *P. massoniana*, *S. superba*, *C. fissa*, *A. acuminatissima* and *C. concinna* by long term light gradient and short term NaHSO₃ solution treatments.

Species	Relative light intensity (%)	NaHSO ₃ concentration (mM)			
		0	20	50	100
<i>P. massoniana</i>	100	2.69a±0.70	4.76b±0.35	5.47b±0.28	2.36a±0.19
	32	2.18a±0.11	2.06a±0.08	3.34b±0.14	7.11c±0.51
	12	1.92a±0.83	1.88a±0.35	4.94b±0.35	10.82c±0.41
<i>S. superba</i>	100	4.41a±0.74	5.76a±0.11	8.87b±0.12	7.42b±0.02
	32	2.09a±0.93	4.62b±0.18	5.92c±0.34	5.98c±0.16
	12	1.12a±0.39	3.61b±0.07	8.77c±0.66	8.02c±0.64
<i>C. fissa</i>	100	1.19a±0.34	2.36a±0.83	5.35b±0.63	6.22b±0.37
	32	1.59a±0.46	3.37b±0.92	4.14b±0.20	4.40b±0.04
	12	1.54a±0.19	2.29b±0.13	3.21c±0.20	3.95c±0.47
<i>A. acuminatissima</i>	100	5.92a±0.13	6.33a±0.29	9.11b±0.10	11.58c±0.21
	32	4.69a±0.69	6.28b±0.55	7.80b±0.89	8.67c±0.29
	12	1.61a±0.55	4.42b±0.53	4.32b±0.11	4.91b±0.73
<i>C. concinna</i>	100	2.84a±0.78	7.81b±0.47	8.25b±0.82	9.28c±0.22
	32	2.51a±0.83	4.75b±0.96	4.38b±0.71	4.58b±0.25
	12	2.49a±0.10	3.32b±0.83	5.05c±0.75	5.12c±0.15

*Within each row, means with the same letter (a for 0 mM NaHSO₃) are not significantly different at $P < 0.05$, $n=4$

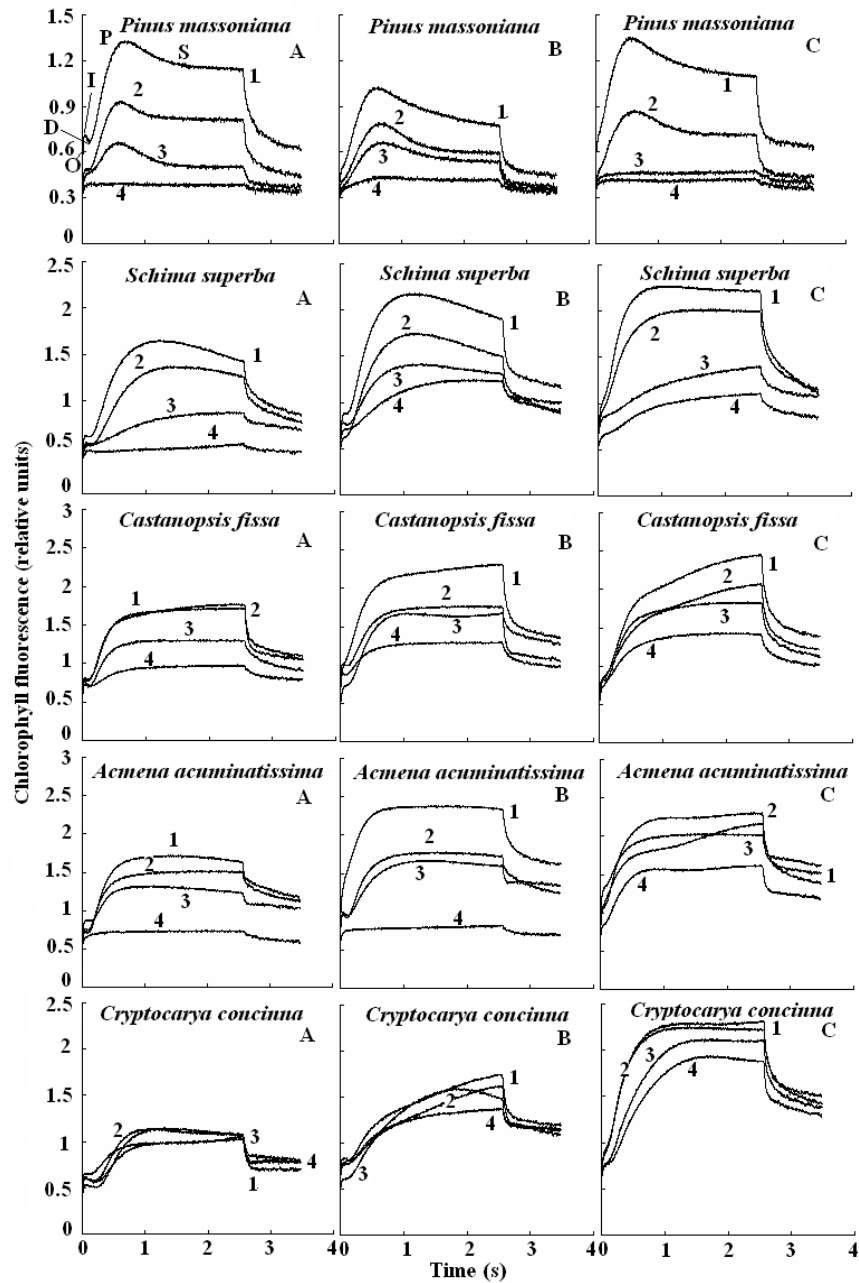


Fig. 4. Chlorophyll a fluorescence induction curves in five plants *P. massoniana*, *S. superba*, *C. fissa*, *A. acuminatissima* and *C. concinna* by long term light gradient and short term NaHSO_3 solution treatments. A-100% light intensity; B-32% light intensity; C-12% light intensity. The number of 1, 2, 3, 4 represents the treatment with 0, 20, 50, 100 mM NaHSO_3 , respectively. Each curve was the mean of 3 or 4 individual experiments.

Table 4 Changes of 1- (Fo/Fp) in chlorophyll a fluorescence in five plants *P. massoniana*, *S. superba*, *C. fissa*, *A. acuminatissima* and *C. concinna* by long term light gradient and short term NaHSO₃ solution treatments.

Species	Relative light intensity (%)	NaHSO ₃ concentration (mM)			
		0	20	50	100
<i>P. massoniana</i>	100	0.63a	0.61a	0.48b	0.18c
	32	0.64a	0.57b	0.54b	0.32c
	12	0.59a	0.56a	0.22b	0.18b
<i>S. superba</i>	100	0.72a	0.70a	0.46b	0.25c
	32	0.72a	0.72a	0.56b	0.50b
	12	0.70a	0.69a	0.46b	0.47b
<i>C. fissa</i>	100	0.70a	0.67a	0.54b	0.38c
	32	0.70a	0.56b	0.71a	0.48b
	12	0.69a	0.69a	0.63a	0.52b
<i>A. acuminatissima</i>	100	0.68a	0.62a	0.51b	0.24c
	32	0.64a	0.61a	0.53b	0.24c
	12	0.56a	0.68a	0.58a	0.60a
<i>C. concinna</i>	100	0.46a	0.60b	0.64b	0.52a
	32	0.55a	0.52a	0.62b	0.42c
	12	0.70a	0.71a	0.71a	0.68a

*Within each row, means with the same letter (a for 0 mM NaHSO₃) are not significantly different at $P < 0.05$, $n = 4$

Discussion

Acidic precipitation arises from the oxidation of sulfur dioxide and nitrogen dioxide in the atmosphere to form H₂SO₄ and HNO₃ acids deposited onto forests via rain, fog, cloud or by dry deposition of gases and vapour (Percy & Ferretti, 2004). Thus the use of buffered H₂SO₃ to exposed cells to different concentrations of SO₂ is arguably realistic (Taylor *et al.*, 1981). The effects of SO₂ on vegetation and agriculture, and its role in the formation of acid rain, continues to be controversial. However, it is reported that, after penetration through the stomata, SO₂ rapidly dissolves in the aqueous phase of the cell wall and, at apoplastic pH value, it reacts with water to form toxic molecular species such as bisulfite and sulfite, which, in turn, can rapidly be converted, through a series of reactions, to a non-toxic sulfate (Rennenberg & Polle, 1994; Ranieri *et al.*, 1999). Rapid metabolic conversion of sulfite to sulfate may be achieved by apoplastic sulfite oxidase activity (Pfanzen *et al.*, 1990). Rennenberg & Polle (1994) calculated that the capacity of apoplastic fluids for enzymatic conversion of sulfite to sulfate is three orders of magnitude higher than the influx of atmospheric SO₂ into the leaf via the stomata at 30ppb SO₂. Therefore, apoplastic sulfite oxidase activity may be able to convert the bulk of the sulfite produced from SO₂ influx into the leaves at the atmospheric SO₂ concentrations which plants experience even in polluted environments. The finding that sulfite accumulates in xylem sap of red spruce trees fumigated in the winter with low SO₂ concentrations is consistent with this assumption (Wolfenden *et al.*, 1991; Rennenberg & Herschbach, 1996). In our experiments, extractions of different plant species reacted with Na₂SO₃ in creating H₂O₂, which demonstrated that five examined forest plants were able to catalyze sulfite into sulfate so as to avoid further damages in bioplasm (Fig. 2, Table 1). However, the sulfite oxidase activities of plant leaves along the natural light gradient were different (lowest under 100% natural light), which implies that leaves grown with long term of low light intensity possessed the better tolerant ability to SO₂. In this way,

activities of this enzyme were mostly consistent to chlorophyll fluorescence emission according to natural light gradient (Table 1, 2). Among the five forest plants examined, *P. massoniana* was more sensitive to high concentration of Na_2SO_3 , with smallest alterations in absorption at 240 nm. Hence, it is deduced that SO_2 and SO_3^{2-} might easily infiltrated into *P. massoniana* cells, yielding and accumulating more highly oxidant molecular species in bioplasm, which resulted in the damage of lipids, proteins and subcellular structures. Contrarily, *A. acuminatissima* could detoxify more sulfite by the enzyme systems to avoid further destructions of plant cells.

The photosynthetic apparatus in plant leaves can harmonize the relationships of different parts, including state transitions, to adapt to various light environments. This phenomenon occurs via the reversible transfer of a fraction of the light harvesting complex II (LHCII) from PSII to PSI (state I to state II transition), as the consequence of its phosphorylation (Allen, 1992; Aro & Ohad, 2003). When PSII is over excited, the photosynthetic apparatus can be converted into state II. Whereas, when PSI is over excited, the photosynthetic apparatus is converted into state I. Murata (1969) reported that when state I to state II transition happens in *Porphyridium cruentum*, 77 K fluorescence emission of PSI increases, while PSII fluorescence intensity decreases. Therefore, the gradually increasing of $F_{\text{PSI}}/F_{\text{PSII}}$ responding to reduced light intensities in Table 2 reflected the long term light environment acclimation in the forest plants by adjustments of state I to state II transition. And also in the present study, the fact that altered the site and reduced emission of PSI associated with the enhancement of PSII emission by NaHSO_3 treatment (Fig. 3) illuminated a state II to state I transition. Taken together with the calculated data of $F_{\text{PSI}}/F_{\text{PSII}}$ (Table. 2), clearly, the impact of NaHSO_3 on energy transformation between two photosystems might be from the modification of their component and structure. The degrees of state transition varied among five forest plants. $F_{\text{PSI}}/F_{\text{PSII}}$ at 77 K of *P. massoniana* changed most, indicating it was damaged more than other species. On the contrary, the photosystems of *C. concinna* had superiority for excited energy distribution in resisting SO_2 pollution.

Investigating membrane fluidity by using fluorescence polarization has become popular due to its many advantages including virtual real time measurements, delay between excitation and emission is 10^{-8} s or less and detect weak signals with high quality instruments (Denich *et al.*, 2003). The fluorescent probes used in fluorescence polarization can embed into the lipid bilayer and are sensitive to alterations in membrane fluidity resulting from structural changes (Luly *et al.*, 1981). The movement of membrane affects the depolarization of the exciting light. This depolarization is used to calculate fluorescence polarization values and lifetimes. These values report on the rotational rate of the probe which reflects the microviscosity of the membrane environment surrounding the probe (Adler & Tritton, 1988). Since PSI and LHCI are imbedded in the bilayer of thylakoid membrane and when excited at 436 nm, the chlorophyll fluorescence emission (at 77 K) reached maximum at 742 nm. Therefore, it is used as probe in our experiments to investigate the possible alteration of thylakoid membrane fluidity. After treating by simulated SO_2 , the elevation of low temperature chlorophyll polarization of the five forest plants (Table. 3) was thought to elucidate the increasing thylakoid membrane mucosity and decreasing fluidity. If this is true, then the higher increment of polarization in *P. massoniana* and lower increment of polarization in *C. concinna* by the combination of short term NaHSO_3 (100 mM) and long term low light intensity again evidenced their different tolerance to SO_2 .

Two phases are clearly distinguished in the kinetics of fluorescence induction: the photochemical phase is completed within 1-2 ms and the thermal phase lasts for 200 ms after the onset of light (Schreiber & Krieger, 1996). The former is attributable to the light-induced reduction of Q_A, the primary quinone acceptor of photosystem II, whereas the thermal phase is not directly related to the redox state of Q_A (Samson *et al.*, 1999). In the curve, O→I means the separation of electron from PSII (Critchley & Smillie, 1981). Hawkins (1985) believes, if the duration of O→I→D is lengthened, the water photolysis will be hindered and the electron numbers arriving at PSII will be decreased. The changes of O→P process is due to the deoxidization of PQ pool and the slowing of corresponding electron transport speed. The appearance of P→S changes may be due to the excitation of dark reaction in photosynthesis, which consumes the electrons at PSI and results in oxidation of PQ pool (Melis & Zeiger, 1982). The induction curve changes its shape according to many environmental conditions, such as light intensity, temperature, drought or chemical influences. The calculation of several phenomenological and biophysical expressions leads to the dynamic description of a photosynthetic sample at a given physiological state. The maximal measured fluorescence intensity, F_p, which can be denoted as F_m if the excitation intensity was experimentally found, in all physiological states studied, to permit the closure of all reaction centers. The fluorescence intensity at 50 μs, considered to be the intensity when all reaction centers are open (Krüger *et al.*, 1997). Therefore, 1- (F_o/F_p) were used to estimate the impairments of electron transport pathway to PSII reaction centers of the five forest plant species to simulated SO₂ pollution (Table. 4). The results calculated from this formula in control leaves were not changed significantly by different light intensities. However, after gradient increasing of NaHSO₃ concentrations, the values of *P. massoniana* were lowered down most but remained unchanged in *C. concinna*. It is speculated that the impact of photosynthetic electron transport processes were aggravated by high NaHSO₃ concentration and the injury effect was different among species by SO₂ pollution.

In South China, the subtropical evergreen monsoonal broadleaf forest, stands alone in the world, develops in the succession process of conifer forest → coniferous and broad-leaved mixed forest with conifer dominated → coniferous and broad-leaved mixed forest with heliophyte broad-leaved species dominated → evergreen broad-leaved forest with heliophyte broad-leaved species dominated → evergreen broad-leaved forest with shade-tolerance species (Peng *et al.*, 1996). Five woody plants selected in this experiment were the dominant species in different succession periods. Studying on the responses and adaptations of these plant species grown under different light intensities to simulated SO₂ and acid precipitation are useful to find out the effects of human activities on subtropical forest structures and functions. The total results demonstrated that *P. massoniana*, the dominant species in early stage of succession, was lower in sulfite oxidase activity and its fluorescence parameters, particularly under 12% natural light illumination were sensitive to simulated SO₂ treatment compared with the other broad-leaved species. Whereas the late stage dominant species *C. concinna* exhibited better tolerance to simulated SO₂ treatment from chlorophyll fluorescence aspects. Hence, we deduced that in subtropical coniferous and broad-leaved mixed forest, acid pollution and low light will accelerate the decline of *P. massoniana* (including other heliophyte forests) and promote succession course. On the contrary, forest climax dominant species like *C. concinna* less sensitive to simulated SO₂ treatment are propitious to its development in the forest.

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