MOLECULAR AND MORPHOLOGICAL CHARACTERIZATION OF SELECTED *MENTHA* SPECIES

ZABTA KHAN SHINWARI¹, SIDRA SULTAN² AND TARIQ MAHMOOD²

¹Department of Biotechnology, Faculty of Biological Sciences, Quaid-i-Azam University, Islamabad, Pakistan ²Department of Plant Sciences, Faculty of Biological Sciences, Quaid-i-Azam University, Islamabad, Pakistan

Abstract

The present study was conducted with the objective of finding out the concordance between morphological data and molecular data of *Mentha spicata* and *Mentha royleana*. Moreover the study was aimed to investigate the relationship between and within the two species. *Mentha* species were collected from QAU, Shadara, Chattar, Donga gali and Qarshi industries, Hattar. For molecular study Random Amplified Polymorphic DNA was performed which was efficient in detecting polymorphism and genetic variation within and between *Mentha* species. Random primer OPC-9 generated a total of 63 bands sharing 93.6% polymorphism in *Mentha spicata* whereas OPC-4 generated 30 bands showing 100% polymorphism in *Mentha royleana*. The morphological and molecular data were analysed using softwares NTSYS pc version 2.02. The results indicated a considerable level of morphological and molecular diversity between the species.

Introduction

Family Labiatae comprises of about 210 genera and some 3,500 species (Davidson, 1999). The family name Labiatae refers to the flowers that have typical petals fused into an upper lip and a lower lip. Although this is still considered an acceptable alternate name, most botanists now use the name "Lamiaceae" in referring to this family. The genus Mentha belongs to the family Lamiaceae (Labiatae) consisting of about 25 to 30 species mainly found in temperate regions of Eurasia, Australia, South Africa and North America (Brickell & Zuk, 1997). All mints prefer and thrive in cool, moist spots in partial shade (Bradley, 1992). Mints grow 10 -120 centimeters tall and can spread over an indeterminate sized area. In Botany, mint is the common name for any of the various herbaceous plants and perennial aromatic herbs that are cultivated for their essential oils and culinary purposes. The genus Mentha L. (Lamiaceae) produces secondary metabolites phenols, alkaloids, flavanoids, such as gummy polysaccharides. Terpens and quinines are used in food and pharmaceutical, cosmetics and pesticide industries (Khanuja et al., 2000). Some members of this genus are also used as herbal teas and condiments both in fresh and dried form due to their distinct aroma (Baser et al., 1995).

The most common species of the genus Mentha found in Pakistan are M. pulegium, M. arvensis, M. spicata, M. longifolia, M. piperita and M. royleana (Hedge & Wendelbo, 1978). However Shinwari & Chaudhri (1992) revised the genus and re- examined its taxonomy. They reported 5 species in Pakistan viz., M. arvensis, M. piperita, M. spicata, M. longifolia and M. royleana. Morphological markers (such as plant height, leaf shape, colour, etc) are among the oldest markers used in the evaluation of genetic variability. However, they are not sufficiently specific and informative because different gene expression in different environments causes wide variability of phenotypic characters in individuals. In some cases congruence between morphology and molecular phyllogenetics were reported. (Shinwari,1995). Genetic diversity refers to the variation at the level of individual gene and provides a mechanism for the plants to adapt in ever changing environment.

Random Amplified Polymorphism DNA (RAPD) markers are a modification of Polymerase Chain Reaction (PCR) used in the late 1980 (Williams *et al.*, 1990). Among PCR based molecular markers RAPD is a widely used technique in different plants (Nazar & Mahmood, 2011; Kayani *et al.*, 2011; Mahmood *et al.*, 2011; Mahmood *et al.*, 2010a; Mahmood *et al.*, 2010b). PCR technique is one of the best available DNA-based tools for scoring variations between cultivars within species (Lakshmikumaran & Bhatia, 1998).

One probable disadvantage is the degree of reproducibility of these markers, sometimes which can be low (Muralidharan & Wakeland, 1993; Ellsworth et al., 1993; Skroch & Nienhis, 1995) particularly between laboratories (Penner et al., 1993; Jones et al., 1997). This is due to sensitivity of RAPD banding patterns to reaction conditions, and the difficulty in exactly replicating reaction conditions across laboratories, where different brands of thermocyclers may be used. The technique is being successfully used widely for the estimation of genetic variability as well as the cultivar identification / differentiation in various plant species, including rice (Mackill, 1995), Broccoli and cauliflower (Hu & Quiros, 1991), banana (Howel et al., 1994), Brassica (Jain et al., 1994), Triticum (Chandrashekhar & Nguyen, 1993), Medicago (Yu & Pauls, 1993), Coffea (Orozco-Castillo et al., 1994) and Lycopersicon (Williams & St Clair, 1993) etc. In the present paper, we describe the similarity and diversity in terms of RAPD profiles of two mint species which includes fifteen accessions of each.

Material and Methods

Plant materials: Various species of genus *Mentha* were collected from 5 different sites including Quaid-i-Azam University campus, Shadara, Chattar, Donga gali and Qarshi industries Hattar (Table 1). The voucher was submitted to Herbarium of QAU Islamabad.

Plant DNA extraction: Total genomic DNA was extracted from fresh leaves using CTAB method by Richards (1997).

Amplification and electrophoresis of PCR products: 10 decamer primers of OPC series were used in the present study. RAPD analysis was performed following the protocol of Williams et al., 1990 with minor modifications. The amplification mixture for each sample DNA contained 50 ng of each template, 10.5 µl of water nuclease-free, 12.5 µl of Master Mix (MBI, Fermentas), and 1µl (25pM) of primer in a 25µl of total reaction volume. PCR was performed in gradient thermal cycler (Multigene Labnet) with an initial denaturation at 94°C for 1 minute followed by 44 cycles of denaturation at 94°C for 30 seconds, annealing at 40°C for 1minute and extension at 72°C for 2 minute. Final cycle was same except extension for 7 minute at 72°C. After that PCR contents were stored at 4°C till use. Amplification was checked by running the PCR products on 1.5% agarose gel prepared in 0.5X TAE (Tris Acetate EDTA) buffer at constant voltage of 70 volts for 3 hours. The gel was stained for 30 minute with ethidium bromide and visualized using gel documentation system. The molecular weight of amplified fragments was estimated with the help of 100 bp plus (MBI, Fermentas) DNA ladder.

Table 1. Species of the genus <i>Mentha</i> collected from different sites
--

Accession No.	Species	Localities	Accession No.	Species	Localities
1	Mentha royleana	QAU	16	Mentha spicata	QAU
2	Mentha royleana	QAU	17	Mentha spicata	QAU
3	Mentha royleana	QAU	18	Mentha spicata	QAU
4	Mentha royleana	Shadara	19	Mentha spicata	Shadara
5	Mentha royleana	Shadara	20	Mentha spicata	Shadara
6	Mentha royleana	Shadara	21	Mentha spicata	Shadara
7	Mentha royleana	Chattar	22	Mentha spicata	Chattar
8	Mentha royleana	Chattar	23	Mentha spicata	Chattar
9	Mentha royleana	Chattar	24	Mentha spicata	Chattar
10	Mentha royleana	Donga gali	25	Mentha spicata	Donga gali
11	Mentha royleana	Donga gali	26	Mentha spicata	Donga gali
12	Mentha royleana	Donga gali	27	Mentha spicata	Donga gali
13	Mentha royleana	Qarshi, Hattar	28	Mentha spicata	Qarshi, Hattar
14	Mentha royleana	Qarshi, Hattar	29	Mentha spicata	Qarshi, Hattar
15	Mentha royleana	Qarshi, Hattar	30	Mentha spicata	Qarshi, Hattar

Data scoring: The amplified products were sored as 1 for presence and 0 for absence respectively. The analysis was plotted in the form of a dendrogram. All computations were carried out using the NTSYS- pc, version 2.2 package (Rohlf, 2005). A dendrogram based on similarity coefficients was generated using the unweighted pair group method of arithmetic means (UPGMA) by NTSYS software.

Results and Discussion

The present study was conducted with the objective of finding out the morphological and molecular diversity to investigate the relationship within and among two species of Mentha i.e., Mentha spicata and Mentha royleana. The amplification profiles produced by five primers gave a total of 215 bands in M. royleana, out of which 150 bands were monomorphic while 59 were polymorphic. The percentage polymorphism observed in all M. royleana species was 27.44%. Maximum number of bands was produced by primer OPC- 5 (66) and minimum by primer OPC- 6 (16). In Mentha spicata, total 277 bands were produced by five primers. Out of which 135 bands were monomorphic while 131 were polymorphic. The percentage polymorphism observed in all M. spicata species was 47.2%. In this study OPC-5 generated maximum number of bands (73) while OPC-6 amplified minimum number of bands (41). It was observed that OPC 2 and OPC 9 generated maximum number of bands while OPC 5 amplified minimum number of bands across all the genotypes. The monomorphic bands are constant bands and cannot be used to study diversity while polymorphic bands revealed differences and can be used to examine and establish systematic relationships (Hadrys et al., 1992). The variation in the number of bands amplified by different primers are influenced by variable factors such as primer structure, template quantity and less number of annealing sites in the genome (Kernodle et al., 1993).

Morphological studies were carried out based on 31 morphological characters including stem, leaf size and leaf colour, inflorescence and other floret characteristics. The data was analyzed by UPGMA cluster analysis based on similarity level and using software NTSYS. The 30 accessions belonging to two different species formed 2 major clusters (Fig. 1). *M. royleana* showed 84% similarity while *M. spicata* showed 88% similarity. On the basis of morphological data *M. royleana* showed 39% similarity while *M. spicata* showed 49% similarity. Fig. 1 shows dendrogram showing morphological relationship among two species i.e., *Mentha*

spicata and Mentha royleana. At 35% similarity coefficient, two distinct groups were observed i.e., Group 1 represents *M. spicata* and Group 2 represents *M. royleana*. It was intresting to note that each intact larger group contained samples of single species. This indicated that the status of the species is valid. Group 1 contained all samples from *Mentha spicata* and group 2 contained all the samples of *Mentha royleana*.

In Group 1 there were 2 major Sub groups.Sub group 1 contained accessions 26, 24, 30, 23, 25, 27, 20, 28, 29, 19, 18 showing more genetic similarity while Subgroup 2 included accessions 22, 21, 17, 16, were genetically close to each other. In Group 2, Sub group 3 included only 2 accessions i.e., 6 and 13 while all the rest accessions 10, 4, 3, 8, 11, 5, 12, 7, 15, 2, 9, 14, 1 were clustered in sub group 4.

According to molecular data, UPGMA cluster analysis revealed two major clusters, cluster 1 of M. spicata and cluster 2 of *M. royleana* independently (Fig. 2). Cluster 1 included 2 accessions 20 and 28 which showed 88% similarity. There were two distinct sub-clusters in cluster 1: (1) within sub-cluster 1, two subgroups are clearly defined: (1) 19, 24, 29, 22 27, 23. (2) 18, 26, 21, 25 showed 95% similarity. Sub-cluster 2 included 16, 30 and 17. In the sub-cluster 1, maximum similarity was observed between M. spicata 19 and M. spicata 24 (98%). Cluster 2 included all accessions of M. royleana showed greater divergence as compared to M. spicata. Cluster 2 divided into two major sub clusters. Cluster 2 included accession 9 which remained unresolved due to greater diversity as compared to other accessions. At 91% similarity coefficient, sub cluster 1 was further divided into two subgroups. Subgroup 1 included accessions 6, 13, 8 and 14. It was observed that 6 and 13 showed 95% similarity with each other while 8 and 14 showed 92% similarity. Subgroup 2 included 2 accessions 10 and 15 which also showed 92%. Sub cluster 2 showed greater diversity as compared to sub cluster 1. Sub cluster 2 showed 86% similarity and included 2 subgroups. Subgroup 1 included two accessions i.e., 2 and 12. They showed 93% similarity with each other. Subgroup 2 included accessions 1, 4, 11, 5 and 3. Accessions 2 and 15 of Mentha royleana collected from Quaid-i-Azam University Islamabad and Qarshi industries showed 100% morphological similarity with eachother. Accessions 4 and 11 of Mentha royleana collected from Shadara & Donga Gali showed 100% genetic similarity. This suggested that geographical origin is not always a good predictor of genetic structure among populations. The present work is consistent with that of Soltis & Soltis (1991). Study on genetic diversity of Mentha species showed that the taxa maintained high levels of genetic

polymorphism among species but not among populations. The polymorphism within populations depicted genotype richness, recombination and gene flow. Clustering of populations based on UPGMA cluster analysis showed some unresolved accessions which were not clustered together. Polymorphisms revealed through RAPD technique may be due to deletion, elimination of primer binding site, an insertion making a fragment too large for polymerization and nucleotide substitutions in the primer annealing site (Fritsch & Rieseberg, 1992). The analysis of genetic variation both within and among plant materials is of fundamental interest to plant breeders. The genetic diversity of *Mentha* species is imprecise and of heterozygote nature. This obscures the determination of genetic diversity patterns based on morphological and phonological observations (Campos-de-Quiroz & Ortega-Klose, 2001).



Fig. 1. UPGMA cluster analysis based on morphological data.





Fewer studies have been made on this genus, so a need was felt to explore and study this economically important genus. Considerable morphological and genetic variation was observed among Mentha species also showed close affinities with each other which might be due to sharing of almost similar habitat and ecology. The study was a starting point to explore Mentha of Further hilly areas. studies need to be done on different aspects including more species ecology, medicinal importance and further molecular studies.

References

- Baser, K.H.C. 1995. Essential oils from aromatic Antibacterial and antifungal activity of the essential plants which are used as herbal tea in Turkey. In: Fla- oils of Thymus revolutus Celak from Turkey. J. Ethvours, Fragrance and Essential Proceedings Oils. of noph.-Pharmacol., 76: 183-186.
- Bradley, F.M. 1992. Rodale's All-new Encyclopedia of Organic Gardening. Emmaus.
- Brickell, C. and J.D. Zuk. 1997. The American Horticultural Society: A-Z Encyclopedia of *Garden Plants*. New York.
- Campos-de–Quiroz, H and F. Ortega-Klose. 2001. Genetic variability among elite red clover (*Trifolium pretense* L. parents used in chile as revealed by RAPD markers. *Euphytica*, 122: 61-67.

- Chandrashekhar, P.J. and H.T. Nguyen. 1993. Application of RAPD technique for the detection of polymorphisms among wild and cultivated tetraploid wheats. *Theor. Appl. Genet.*, 36: 602-609.
- Davidson, A. 1999. *The Oxford Companion to Food*. Oxford: Oxford University Press.
- Ellsworth, D.L.K. Rittenhouse and R.L. Honeycutt. 1993. Artificial variation in random amplified polymorphic DNA banding patterns. *Biotech.*, 14: 214-218.
- Fritsch, P and L.H. Rieseberg. 1992. High out crossing rates maintain male and hermaphrodite individuals in populations of the flowering plant *Datisca glomerata*. *Nature*, 359: 633-636.
- Hadrys, H., M. Balick and B. Schierwater. 1992. Applications of Random Amplified Polymorphic DNA (RAPD) in molecular ecology. *Mol. Ecol.*, 1: 35-63.
- Hedge, I.C. and P. Wendelbo. 1978. Patterns of distribution and endemism in Iran. *Notes from Royal* Botanic Garden, Edinburgh, 36: 441-464
- Howell, E.C., H.J. Newbury, R.L. Swennen, L.A. Withers and B.V. Ford- Lloyd. 1994. The use of RAPD for identifying and classifying *Musa* germplasm. *Genome*, 37: 328-332.
- Hu, J. and L.F. Quiros. 1991. Identification of Broccoli and Cauliflower Cultivar with RAPD markers. *Plant Cell Rep.*, 10: 505-511.
- Jain, A., S. Bhitia., S.S. Banga., S. Prakash and M. Laxmikumaran. 1994. Potential use of RAPD technique to study the genetic diversity in Indian mustard (*Brassica juncea*) and its relationship to heterosis. *Theor. Appl. Genet.*, 88: 116-122.
- Jones, N., H. Ougham and H. Thomas. 1997. Markers and mapping: we are all geneticists now. *New Phytologist*, 137: 165-177.
- Kernodle, S.P., R.E. Cannon and J.G. Scandalios. 1993. Cincenteration of primer and template qualitatively affects product in RAPD – PCR. *Biotechniques*, 1: 362-364.
- Khanuja, S.P.S., A.K.A. Shasany, A. Srivastava and S. Kumar. 2000. Assessment of genetic relationships in *Mentha* species. *Euphytica*, 111: 121-125.
- Lakshmikumaran, M. and S. Bhatia. 1998. DNA fingerprinting of medicinal plants. In: *Intellectual Property Rights*. (Ed.): B. Debry. Rajiv Gandhi Institute for Contemporary studies B.R. Publishing Corporation, Delhi. pp. 293-331.
- Mackill, D.J. 1995. Classifying japonica rice cultivars with RAPD markers. Crop Sci., 35: 889- 894.
- Mahmood, T., A. Siddiqua, A. Rasheed and N. Nazar. 2011. Evaluation of genetic diversity in different Pakistani wheat land races. *Pak. J. Bot.*, 43(2): 1233-1239.
- Mahmood, T., N. Nazar, B.H. Abbasi, M.A. Khan, M. Ahmad and M. Zafar. 2010a. Detection of somaclonal variations using RAPD fingerprinting in *Silybum marianum* L. J. Med. Plant Res., 4(17): 1822-1824.

- Mahmood, T., S. Muhammad and Z.K. Shinwari. 2010b. Molecular and morphological characterization of *Caralluma* species. *Pak. J. Bot.*, 42(2): 1163-1171.
- Muralidharan, K. and E.K. Wakeland. 1993. Concentration of primer and template qualitatively affects products in random amplified polymorphic DNA PCR. *Biotech.*, 14: 362-364.
- Nazar, N. and T. Mahmood. 2011. Morphological and molecular characterization of selected *Artemisia* species from Rawalakot, Azad Jammu and Kashmir. *Acta Physiol. Plantarum*, 33: 625-633.
- Orozco-Castillo, C., K.J. Chalmers, R. Waugh and W. Powell. 1994. Detection of genetic diversity and selective gene introgression in coffee using RAPD markers. *Theor. Appl. Genet.*, 87: 934- 940.
- Penner, G.A., A. Bush, R. Wisw, W. Kim, L. Domier, K. Kasha, A. Laroche, G. Scoles, S.J. Molnar and G. Fedek. 1993. Reproducibility of random amplified polymorphic DNA (RAPD) analysis among laboratories. *PCR Methods and Applications*, 2: 341-345.
- Richard, E.J. 1997. Preparation of plant DNA using CTAB. In: Short protocols in Molecular biology. (Eds.): F. Ausubel, R. Brent, R.E. Kingston, D.D. Moore, J.G. Siedman, J.A. Smith and K. Struhl. Wiley, New York: 2.10-2.11.
- Rohlf, F. J. NTSYSpc (Numerical Taxonomy and Mutivarate Analysis System). Version 2.2, 2005. Exeter Software. Applied Biostatistics Inc. New York. USA.
- Shinwari, Z.K. and M.N. Chaudheri. 1992. Taxonomic revision of genus Mentha from Pakistan Acta Phytotax. Geobot., 43(2): 97-110.
- Shinwari, Z.K.1995. Congruence between morphology and molecular phylogeneties in Prosartes (Liliaceae). *Pak. J. Bot.*, 27(2): 361-369.
- Skroch, P. and J. Nienhuis. 1995. Impact of scoring error and reproducibility of RAPD data on RAPD-based estimates of genetic distance. *Theor. Appl. Genet.*, 91:1086-1091.
- Soltis, P.S. and D.E. Soltis. 1991. Genetic variation in endemic and widespread plant species, examples from Saxifragaceae and Polystiochum (Dryopteridaceae). *Aliso*, 13: 215-223.
- Williams, C.E. and D.A. St Clair. 1993. Phenetic relationship and levels of variability detected by RFLP and Random amplified DNA analysis of cultivated and wild accessions of *Lycopersicon esculentum. Genome*, 36: 619-630.
- Williams, J.G.K., A.R.K. Kubelik., T. Livak., J.A. Rafalski and S.V. Tingey. 1990. Nucleic Acids Research. 18: 6531-6539.
- Yu, K.F. and K.P. Pauls, 1993. Rapid estimation of genetic relatedness among heterogenous populations of alfa- alfa by random amplification of bulked genomic DNA samples. *Theor. Appl. Genet.*, 86: 788-794.

(Received for publication 23 June 2010)