COMPOSITIONAL STUDIES OF LENTIL (*LENS CULINARIS* MEDIK.) CULTIVARS COMMONLY GROWN IN PAKISTAN

M. ZIA-UL-HAQ¹, S. AHMAD²*, M. ASLAM SHAD³, S. IQBAL⁴, M. QAYUM⁵, A. AHMAD⁶, D. L. LUTHRIA⁷ AND R. AMAROWICZ⁸

¹Department of Pharmacognosy, University of Karachi, Karachi-75270, Pakistan, ²Department of Agronomy, ³Department of Chemistry, Bahauddin Zakariya University, Multan-60800, Pakistan, ⁴Department of Chemistry, University of Sargodha, Sargodha-40100, Pakistan, ⁵Department of Pharmacy, University of Peshawar, Peshawar -25120, Pakistan, ⁶Department of Agronomy, Agro-Climatology Laboratory, University of Agriculture, Faisalabad- 38040, Pakistan, ⁷Food Composition and Method Development Laboratory, USDA, ARS, BHNRC, Beltsville, MD 20705, USA and ⁸Department of Chemical and Physical Properties of Food, Institute of Animal and Reproduction and Food Research of the Polish Academy of Sciences, 10-747 Olsztyn, Tuwima 10, Poland

Abstract

Four improved lentil cultivars viz., Masoor 85, Masoor 93, NIAB Masoor 2002 and NIAB Masoor 2006 grown and consumed in Pakistan have been systematically analyzed to determine and compare their nutritional and compositional properties. Proximate analysis, anti-nutritional contents, amino acid and fatty acid profiles of the oil extracted along with mineral content from all four cultivars were investigated. Mineral composition showed that sufficient amounts of Ca, P, K, Cu, Zn and Mg were present to meet the macro and micro-nutrients demand in human diets. Despite variations, potassium and manganese were noted as being present in highest and lowest concentrations, respectively, in all cultivars. The distribution patterns of various amino acids in these cultivars suggested sulfur containing amino acids as limiting amino acids. Fatty acid profile indicated unsaturated fatty acids as major fatty acids in all cultivars. The data show that, in terms of both quality and quantity, all four lentil cultivars can serve as a significant source of essential amino acids, essential fatty acids and trace minerals to meet the demand of population of Pakistan.

Introduction

Lentil (*Lens culinaris* Medik.) is predominantly grown in South East Asia and commonly consumed as thick soup made from whole grain or split pulse commonly referred to as 'dhal'. Seeds can be fried and seasoned for consumption; flour is used to make soups, stews purees, and mixed with cereals to make bread and cakes, and as a food for infants (Williams & Singh, 1988). It is used in culinary dishes in the Indo-Pakistan sub-continent and in the Middle East and incorporated into soups in Europe and North America. In Western countries, lentils may be used in casseroles and as meat substitutes in vegetarian diets. Lentil although called as a 'poor man's meat', is equally liked by all socioeconomic groups in South East Asia (Bhatty, 1988).

Lentils are excellent source of protein and also rich in important vitamins, minerals, soluble and insoluble dietary fiber. The unsaponifiable lipid fraction of lentil is a potential source of bioactive components such as phytosterols, squalene and tocopherols (Ryan *et al.*, 2007). Lentils contain saponins (triterpene glycosides), which have been implicated in hypercholesterolemia in animals (Savage, 1991) and phenolic compounds with high antioxidant activity (Amarowicz & Pegg, 2008; Amarowicz *et al.*, 2009, 2010). Besides this, it is a valuable green manure and used as a forage crop. Husks, dried leaves, and stems are used as livestock feeds (Anon., 2000). All these factors have contributed to place the cultivation of lentils at the same economic level as that of cereals with the additional value that its cultivation is more environment-friendly, as it adds to soil fertility by symbiotic nitrogen fixation.

Lentil is the second largest grown legume crop of *Rabi* season in Pakistan after chickpea (*Cicer arietinum* L.) both in quality and quantity (Ayub *et al.*, 2001). In 2006 lentil was grown on 43,4000 ha with 25,9000 tones production and average yield of 597 kg/ha (MINFAL, 2006). In recent years, lentil production in Pakistan has increased substantially. This has been brought about by the development of new lentil cultivars with higher yields, improved adaptation to local agroclimatic conditions and better acceptability through improved nutritional status such as fatty acid and ANF (Anti-

The seeds of four lentil (*Lens culinaris* Medik.) cultivars, Masoor 85, Masoor 93, NIAB Masoor 2002, and NIAB Masoor 2006 were procured from Department of Agronomy, Bahauddin Zakariya University, Multan. Seeds of all the varieties were divided into groups for storage in stainless-steel containers at 4°C prior to analysis.

consumed in Pakistan.

Materials and Methods

Proximate analysis: Moisture, lipids, ash, protein and carbohydrates were determined according to AOAC methods (Anon., 1990).

nutritional factor profiles), by the expansion of export markets, and through a keener appreciation of the benefits of crop

rotation and alternative cropping systems. In perspective of

nutritional benefits and nutraceutical attributes of lentil,

characterization and compositional analysis of its seed are of great importance. Current food databases contain limited or

dated compositional data and antioxidant activity by different

assay procedures on different lentil cultivars. As part of our

studies to explore the flora of Pakistan (Ahmad et al., 2010;

Zia-Ul-Haq et al., 2007; 2008 a, b; 2009 a, b; 2010 a, b; Nisar

et al., 2010 a, b, c) we have determined the chemical composition of four lentil cultivars commonly grown and

Minerals analysis: The samples were incinerated at 450°C for 12 h in a muffle furnace and acid digest was prepared by oxidizing each sub-sample with a nitric/perchloric acid (2:1) mixture. Aliquots were used to estimate Na and K by flame photometer (Flame Photometer Model-EEL). The minerals, such as calcium, manganese, magnesium, zinc, iron and copper were determined with an atomic absorption spectrophotometer (Perkin–Elmer Model 5000) while phosphorus was determined by the phosphovanado-molybdate (yellow) method (Anon., 1990). The samples were quantified against standard solutions of known concentration that were analyzed concurrently.

*Corresponding author: shakeel.agronomy@gmail.com

Amino acid analysis: Samples (300 mg), in triplicate from each cultivar, were hydrolyzed with 6 M HCl in an evacuated test tube for 24 h at 105°C. The dried residue was dissolved in citrate buffer (pH 2.2) after flash evaporation. Aliquots were analysed in an automatic amino acid analyser (Hitachi Perkin– Elmer Model KLA 3B), using the buffer system described earlier (Khalil *et al.*, 1990). Methionine and cystine were analysed separately after performic acid treatment and subsequent hydrolysis with HC (Khalil *et al.*, 1990). Tryptophan was determined after alkali (NaOH) hydrolysis by the colorimetric method (Freidman & Finely, 1971). Essential amino acids score was calculated with reference to the FAO/WHO reference amino acid pattern (Anon., 1985).

Amino acid score =
$$\frac{\text{Test amino acid}}{\text{Reference amino acid}} \times 100$$

Fatty acid (FA) composition: Fatty acid methyl esters (FAMEs) were prepared according to the standard of IUPAC method 2.301 (Anon., 1987) and analyzed on a Shimadzu 17-A gas chromatograph with flame ionization detector (FID). Separation was done on a capillary column SP 2330 (30 m 7 0.32 mm $\overline{7}$ 0.25 μ m; Supelco; Bellefonte, Pa., U.S.A.). Nitrogen was used as a carrier gas at a flow rate of 3.0 mL/min. Column temperature was programmed from 180 to 220 °C at the rate of 3 °C/min. Initial and final temperatures were held for 2 and 10 min, respectively. Injector and detector were kept at 230 and 250 °C, respectively. A sample volume of 1.0 µL was injected with the split ratio of 1:75. FAMEs were identified by comparing their relative and absolute retention times to those of authentic standards. The quantification was done by a Chromatography Station for Windows (CSW32) data handling software (Data Apex Ltd. CZ-158 00 Prague 5, the Czech Republic). The fatty acid composition was reported as a relative percentage of the total peak area and the results were calculated as mg/100 g of dry lentil seeds.

Antinutritional factors: Trypsin inhibitor activity was estimated according to the method of Kakade *et al.*, (1969) using benzoyl-DL-arginine- *p*-nitroanalide hydrochloric as substrate.

Statistical analysis: Analyses were performed in triplicate. Data analysis was carried out using the analysis of variance and LSD test using the "MSTATC" statistical computer package.

Results and Discussion

At the national level, efforts are being made to develop higher yielding varieties of pulses to increase their production and per capita availability and to improve the nutritional status of the people. However, information is needed on the biochemical composition of cultivars to understand their nutritional profiles from production and consumption points of view.

The results of the proximate composition are reported in Table 1. The observed range for protein was 28.80 % for Masoor 93 to 30.60% for NIAB Masoor 2006 and the results are in agreement for protein content in lentils that have been reported by several other workers. Iqbal *et al.*, (2006) reported the content of protein for lentil of 26.1 g/100 g. According to Boyle *et al.*, (2010) the contents of protein in green and red lentils were 23.03 and 25.88 g/100 g respectively. The crude fiber content ranged from 6.99% to 8.14%. The range observed for lipid content was between 1.93 and 2.15% while carbohydrates showed a range from 54.08% to 55.81%. The results are in agreement to those reported earlier (Solanki *et*

al., 1999; El-Adawy *et al.*, 2003). Seeds of NIAB Massor 2002 were characterized by the highest content of ash (5.72 mg/100g) where Massor 85 exhibited the lowest one (4.16 mg/100g). In the study of Wang *et al.*, (2009) the content of ash of lentils ranged from 2.48 to 2.84 mg/100g.

Mineral constituents of lentil seeds (Table 2), varied among the cultivars, but potassium constituted the major mineral. Potassium content ranged from 875 mg/100 g in NIAB Masoor2006 to 872mg/100 g in Masoor 85. Sodium was found in lower quantity in NIAB Masoor2002 (76mg/100 g) while NIAB Masoor2006 had the highest iron (3.2 mg/100 g) content. All cultivars contained good amounts of calcium, zinc and copper. The results correspond to those already reported for lentil in Pakistan (Amjad *et al.*, 2006). These results revealed that lentils may provide a sufficient amount of minerals to meet the human mineral requirement.

The amino acid composition of the four lentil cultivares indicated little variation in the content of essential and nonessential amino acids (Table 3). However, significant variation existed in the individual amino acids, for example the content of isoleucine varied from 3.9 (Masoor 93) to 4.4 g/16 g N (NIAB Masoor 2006); cystine from 0.5 (NIAB Masoor 2006) to 0.9 g/16 g N (Masoor 93). Glutamic acid and aspartic acid were found to be the major non-essential amino acids in the sample tested. The lowest essential to nonessential AA ratio was noted for Masoor 93 (0.78) and the highest for NIAB Masoor 2006 (0.84). The results obtained are in fair agreement with those reported for lentil by Iqbal et al., (2006) and Boyle et al., (2010). Iqbal et al., (2006) reported a value of 0.81 as an essential to nonessential AA ratio for lentil. The chemical score and amino acid index are widely used for screening potential protein foods. Essential amino acid score was computed with reference to the FAO/WHO (Anon., 1985), standard amino acid profiles established for humans. The data in Tables 3 and 4 indicate that all essential amino acids, except S-containing types and tryptophan are present in excessive amounts in all the cultivars analyzed. Amino acid profile showed methionine and cystine as the limiting amino acids. Results are comparable to those of earlier workers (Amjad et al., 2006, Boyle et al., 2010). Amino acid deficiency can be met by consuming large amounts of legumes or by taking a mixture of legumes or by employing the complementarity that exists between high sulfur amino acid cereals and legumes, especially the soybean.

Data about the qualitative and quantitative composition of fatty acids are summarized in Table 5. Fatty acid profile of all lentil cultivars reveals the lipids as a good source of the nutritionally essential linoleic and oleic acids. Linoleic acid, palmitic acid and oleic acid were the dominating fatty acids. Most of the fatty acids were unsaturated fatty acids, while saturated fatty acids mainly palmitic acid contributed little of the total fatty acids content. The fatty acid composition and high amounts of unsaturated fatty acids make lentil a special legume suitable for nutritional applications. The presence of high levels of unsaturated fatty acids in all the presently studied cultivars is nutritionally desirable and results are comparable with some edible legumes.

Statistical analysis did not show any significant differences in the content of phytic acid in four lentil varieties (Table 6). In the study of Wang *et al.*, (2006) phytic acid content in Canadian lentil seed was a bit lower and varied from 6.2 to 8.8 mg/g with a mean of 7.7 mg/g. Seeds of NIAB Masoor 2002 were characterized by the lowest content of trypsin inhibitors (29.37 TIU/mg protein). The highest content of trypsin inhibitors was found in seeds of NIAB Masoor 2006 (33.86 TIU/mg protein). The results agree with the data reported for lentil by Wang *et al.*, (2009) and Champ (2002). Germination, extrusion cooking, dehulling and hydroythermal processing are common commercial processes used to inactivate protease inhibitors in pulse crops (Roy *et al.*, 2010). Usually proximate composition of plants and crops seeds varies depending on cultivars,

agrogeoclimatological conditions, maturity and collection time of seed, water and fertilizers application as well as acceptability, selectivity and sucking up of nutrients by plants and crops. This study will pave path for future detailed investigations on this legume crop.

Table 1. Proximate chemical composition (g/100 g) of seeds.							
ComponentsMasoor 85Masoor 93NIAB Masoor 2002NIAB Masoor 2006							
Crude protein	$30.41 \pm 1.71^{\text{a}}$	28.80 ± 1.66^{a}	$29.37 \pm 1.60^{\mathrm{a}}$	$30.60 \pm 1.72^{\mathrm{a}}$			
Total lipids	$2.15\pm0.05^{\rm a}$	$2.09\pm0.05^{\rm a}$	$1.93\pm0.09^{\rm a}$	$2.08\pm0.09^{\rm a}$			
Total carbohydrates	$54.08\pm0.09^{\rm a}$	$55.43{\pm}~1.73^{a}$	$54.74 \pm 1.10^{\mathrm{a}}$	$55.81 \pm 1.75^{\mathrm{a}}$			
Crude fiber	7.74 ± 1.7^{b}	$8.14 \pm 1.6^{\rm a}$	$8.14 \pm 1.6^{\rm a}$	$6.99 \pm 1.6^{\text{b}}$			
Ash	4.16 ± 0.19^{a}	$5.54\pm.18^{ab}$	5.72 ± 0.19^{b}	$4.52\pm0.18^{\rm a}$			

Data are expressed as means \pm standard deviations on dry weight basis; values having different letters differ significantly (p<0.05)

Table 2. Content on mineral compounds (mg/100 g) of seeds.								
MineralMasoor 85Masoor 93NIAB Masoor 2002NIAB Masoor 2006								
Sodium	79 ± 2.65^{a}	79 ± 2.65^{a}	76 ± 1.33^{b}	$30.60 \pm 1.72^{\mathrm{a}}$				
Potassium	$874\pm6.43^{\rm a}$	$872\pm3.78^{\rm a}$	$873 \pm 4.08^{\mathrm{a}}$	$875\pm0.09^{\rm a}$				
Phosphorus	$294\pm3.61^{\text{a}}$	293 ± 2.13^{a}	$292\pm3.08^{\rm a}$	$294\pm2.92^{\rm a}$				
Calcium	$120\pm 6.24^{\text{a}}$	119 ± 5.48^{a}	121 ± 4.73^{a}	118 ± 5.10^{a}				
Iron	3.1 ± 0.26^{ab}	2.9 ± 0.69^{bc}	$2.7\pm0.52^{\circ}$	$3.2\pm0.19^{\rm a}$				
Cooper	$9.9\pm0.10^{\mathrm{a}}$	$8.9\pm0.07^{\rm b}$	$9.5\pm0.04^{\rm a}$	$9.6\pm0.09^{\mathrm{a}}$				
Zinc	4.4 ± 0.20^{ab}	$3.9\pm0.17^{\circ}$	4.6 ± 0.11^{a}	4.2 ± 0.07^{bc}				
Manganes	1.6 ± 0.03^{ab}	4.3 ± 0.07^{bc}	1.4 ± 0.06^{ab}	$1.7\pm0.05^{\rm a}$				
Na:K ratio	0.09	0.08	0.08	0.09				
Ca:P ratio	0.40	0.41	0.40	0.40				

Data are expressed as means \pm standard deviations on dry weight basis; values having different letters differ significantly (p<0.05)

TII 3 4 5 51	• . •	6 1 61	4.1 14.	1	1 ()
Table 3. Amino acid	COMDOSILION	VI SECUS VI ICI	uun cunuvais	12/	102111

Table 5. Amino acid composition of seeds of lentil cultivars (g/ 16 gN).					
Amino acid	Masoor 85	Masoor 93	NIAB Masoor 2002	NIAB Masoor 2006	
Essential AA					
Isoleucine	$4.1{\pm}0.05^{bc}$	$3.9\pm0.07^{\rm a}$	4.3 ± 0.05^{ab}	$4.4\pm0.07^{\rm a}$	
Leucine	7.8 ± 0.05^{ab}	$7.3\pm0.03^{\rm c}$	$7.5\pm0.04^{\rm bc}$	$7.9\pm0.01^{\rm a}$	
Lysine	$7.0\pm0.03^{\rm a}$	$6.9\pm0.01^{\rm a}$	$6.8\pm0.08^{\rm a}$	$7.2\pm0.03^{\rm a}$	
Methionine	$0.8\pm0.02^{\rm a}$	$0.9\pm0.05^{\rm a}$	$0.9\pm0.09^{\rm b}$	$0.6\pm0.02^{\text{b}}$	
Phenylaniline	5.0 ± 0.12^{ab}	4.8 ± 0.06^{ab}	4.3 ± 0.07^{b}	$5.0\pm0.08^{\rm a}$	
Threonine	$3.5ab \pm 0.04^{ab}$	$3.2b\pm0.04^{b}$	$3.7a\pm0.03^{a}$	$3.4ab\pm0.04^{ab}$	
Tryptophan	$0.7ab \pm 0.03^{ab}$	$0.7ab \pm 0.03^{ab}$	$0.8b\pm0.02^{b}$	$0.8a\pm0.05^{a}$	
Valine	$5.0ab\pm0.05^{ab}$	$4.8b{\pm}0.08^{b}$	$5.3a \pm 0.04^{a}$	$4.9ab{\pm}~0.07^{ab}$	
Arginine	$7.8\pm0.03^{\rm a}$	$7.5\pm0.04^{\rm a}$	$7.6\pm0.03^{\rm a}$	$7.6\pm0.03^{\rm a}$	
Histidine	2.2 ± 0.05^{ab}	2.3 ± 0.02^{ab}	$1.9\pm0.01^{\rm b}$	$2.5\pm0.02^{\rm a}$	
Non-essential AA					
Alanine	$4.3\pm0.03^{\rm a}$	$4.2\pm0.07^{\rm a}$	$4.6\pm0.05^{\rm a}$	4.0 ± 0.01^{a}	
Aspartic acid	$11.2\pm0.07^{\rm a}$	$11.8\pm0.08^{\rm a}$	$11.4\pm0.07^{\rm a}$	$11.4\pm0.07^{\rm a}$	
Cystine	0.7 ± 0.08^{ab}	$0.9\pm0.04^{\rm a}$	$0.5\pm0.03^{\circ}$	0.5 ± 0.08^{bc}	
Glutamic acid	$22.0\pm0.05^{\rm a}$	$21.5\pm0.07^{\rm a}$	20.9 ± 0.09^{a}	$21.3\pm0.09^{\rm a}$	
Glycine	3.2 ± 0.04^{a}	$3.6\pm0.05^{\rm a}$	$3.7 \pm 0.04^{\mathrm{a}}$	3.0 ± 0.04^{a}	
Proline	$3.9\pm0.02^{\rm a}$	3.5 ± 0.03^{ab}	3.1 ± 0.01^{b}	3.8 ± 0.07^{ab}	
Serine	$4.9\pm0.03^{\rm a}$	$5.2\pm0.05^{\rm a}$	$5.4\pm0.08^{\rm a}$	$5.0\pm0.03^{\rm a}$	
Tyrosine	3.0 ± 0.01^{a}	$3.2\pm0.06^{\rm a}$	$3.3\pm0.02^{\rm a}$	$3.27\pm0.05^{\rm a}$	
Essential to nonessential AA ratio	0.82	0.78	0.81	0.84	

Data are expressed as means \pm standard deviations; values having different letters differ significantly (p<0.05)

Table 4. Essential amino acid score of protein of lentil cultivars.							
Amino acidMasoor 85Masoor 93NIAB Masoor 2002NIAB Masoor 200							
Isoleucine	146	139	153	157			
Leucine	118	110	113	119			
Lysine	120	118	117	124			
Methionine	60	72	56	56			
Phenylaniline + tyrosine	126	126	120	131			
Threonine	102	94	108	100			
Tryptophan	63	81	72	72			
Valine	142	137	151	140			
Histidine	115	121	100	131			
Limiting amino acid	Sulfur amino acids	Sulfur amino acids	Sulfur amino acids	Sulfur amino acids			

Table 5. Content	of individual fatty	acids (mg/100 g o	f dry matter) of seeds

Table 5. Content of multilular fatty actus (mg/100 g of ury matter) of secus.						
Masoor 85	Masoor 93	NIAB Masoor 2002	NIAB Masoor 2006			
$14.57\pm0.03^{\rm a}$	$13.67\pm0.05^{\rm a}$	13.67 ± 0.05^{a}	$13.67\pm0.05^{\rm a}$			
$0.09\pm0.02^{\rm a}$	$0.03\pm0.04^{\rm a}$	$0.07\pm0.03^{\rm a}$	$0.05\pm0.09^{\rm a}$			
0.13 ± 0.09^{ab}	0.09 ± 0.02^{bc}	$0.17\pm0.04^{\rm a}$	$1.17\pm0.01^{\rm b}$			
$1.17\pm0.01^{\text{b}}$	$1.32\pm0.08^{\rm a}$	$1.32\pm0.08^{\rm a}$	$1.17\pm0.01^{\rm b}$			
$22.65\pm0.08^{\mathrm{a}}$	$22.65\pm0.08^{\rm a}$	$21.87\pm0.08^{\rm a}$	22.11 ± 0.07^{a}			
$47.21\pm0.05^{\rm a}$	$46.98\pm0.03^{\mathrm{a}}$	$47.01\pm0.05^{\rm a}$	$46.89\pm0.05^{\rm a}$			
$11.77\pm0.07^{\rm a}$	$11.21\pm0.02^{\rm c}$	$10.99\pm0.01^{\text{d}}$	$11.43\pm0.06^{\text{b}}$			
$0.44\pm0.04^{\rm a}$	$0.19\pm0.01^{\rm c}$	$0.31\pm0.05^{\rm b}$	0.27 ± 0.04^{bc}			
$0.70\pm0.01^{\rm a}$	0.51 ± 0.03^{b}	0.44 ± 0.07^{b}	$0.65\pm0.08^{\rm a}$			
$0.28\pm0.07^{\rm a}$	$0.31\pm0.05^{\rm a}$	$0.19\pm0.06^{\text{b}}$	0.13 ± 0.09^{b}			
	$\begin{array}{c} 14.57 \pm 0.03^{a} \\ 0.09 \pm 0.02^{a} \\ 0.13 \pm 0.09^{ab} \\ 1.17 \pm 0.01^{b} \\ 22.65 \pm 0.08^{a} \\ 47.21 \pm 0.05^{a} \\ 11.77 \pm 0.07^{a} \\ 0.44 \pm 0.04^{a} \\ 0.70 \pm 0.01^{a} \end{array}$	$\begin{array}{ccccc} 14.57\pm 0.03^{a} & 13.67\pm 0.05^{a} \\ 0.09\pm 0.02^{a} & 0.03\pm 0.04^{a} \\ 0.13\pm 0.09^{ab} & 0.09\pm 0.02^{bc} \\ 1.17\pm 0.01^{b} & 1.32\pm 0.08^{a} \\ 22.65\pm 0.08^{a} & 22.65\pm 0.08^{a} \\ 47.21\pm 0.05^{a} & 46.98\pm 0.03^{a} \\ 11.77\pm 0.07^{a} & 11.21\pm 0.02^{c} \\ 0.44\pm 0.04^{a} & 0.19\pm 0.01^{c} \\ 0.70\pm 0.01^{a} & 0.51\pm 0.03^{b} \end{array}$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$			

Data are expressed as means \pm standard deviations on dry weight basis; values having different letters differ significantly (p<0.05)

Compounds	Masoor 85	Masoor 93	NIAB Masoor 2002	NIAB Masoor 2006
Phytic acid (mg/g dry matter)	11.45 ± 0.31^{a}	$10.99\pm0.32^{\rm a}$	$11.18\pm0.28^{\rm a}$	$11.31\pm0.19^{\rm a}$
Trypsin inhibitors (TIU/mg protein)	31.30 ± 1.25^{ab}	27.93 ± 0.33^{b}	29.37 ± 0.35^{b}	33.86 ± 0.09^{a}

Data are expressed as means \pm standard deviations; values having different letters differ significantly (p<0.05)

References

- Ahmad, S., M. Akhter, M. Zia-ul-Haq, Mehjabeen and S. Ahmed .2010. Antifungal and nematicidal activity of selected legumes of Pakistan. *Pak. J. Bot.*, 42: 1327-1331.
- Amarowicz, R. and R.B. Pegg. 2008. Legumes as a source of natural antioxidants *Eur. J. Lipid Sci. Technol.*, 110: 865-878.
- Amarowicz, R., I. Estrella, T. Hernández, M. Dueñas, A. Troszyńska, A. Kosińska and R.B. Pegg. 2009. Antioxidant activity of a red lentil extract and its fractions. *Int. J. Mol. Sci.*, 10: 5513-5527.
- Amarowicz, R., I. Estrella, T. Hernández, S. Robredo, A. Troszyńska A. Kosińska and R.B. Pegg. 2010. Free radical-scavenging capacity, antioxidant activity, and phenolic composition of green lentil (*Lens culinaris*). *Food Chem.*, 121: 705-711.
- Amjad, L., A.L. Khalil, N. Ateeq and M.S. Khan. 2006. Nutritional quality of important food legumes. *Food Chem.*, 97: 331-335.
- Anonymous. 1985 Energy and protein requirements. WHO Technical Report Series No. 724, Geneva.
- Anonymous. 1987. Standard methods for the analysis of oils, fats and derivatives. International Union of Pure and Applied Chemistry. 7th rev. enlarged (Ed.): C. Paquot, A. Hautfenne, London, U.K.: Blackwell Scientific.
- Anonymous. 1990. *Official Methods of Analysis*. 14th ed. Association of Official Agricultural, Chemists. Washington DC, USA.
- Anonymous. 2000. Processing and Utilization of Legumes. Asian Productivity Organization <u>http://www.apo-tokyo.org/00e-books/AG-12 Legumes.htm</u>
- Anonymous. 2004-2005. MINFAL., Agricultural Statistics of Pakistan, Govt. of Pakistan, Islamabad. (2006) p. 50-51.

- Aurand, L.W., A.E. Woods and M.R. Wells. 1987. Food composition and analysis. Van Nostrand Reinhold Company, New York, USA.
- Ayub K, M. Rahim and A. Khan. 2001. Performance of exotic lentil varieties under rainfed conditions in Mingora (NWFP) Pakistan. *J. Bio. Sci.*, 1: 343-344.
- Bhatty, R.S. 1988. Composition and quality of lentil (*Lens culinaris* Medik): a review. *Can. Inst. Food Sci. Technol. J.*, 21: 144-160.
- Boyle, J.L., S. Aksay, S. Roufik, S. Ribereau, M. Mondor, M. Mondor, E. Farnworth and S.H. Rajamohamed. 2010. Comparison of the functional peoperties of pea, chickpea and lentil protein concentrates processed using ultrafiltration and isoelectric precipitation techniques. *Food Res. Int.*, 43: 537-546.
- Champ, M.M.J. 2002. Non-nutrient bioactive substances of pulse. Brit. J. Nutr., 88: S307-S319.
- El-Adawy, T.A., E.H. Rahma, A.A. Eel-Bedawey and A.E. El-Beltagy. 2003. Nutritional potential and functional properties of germinated mung bean, pea and lentil seeds. *Plant Foods Hum. Nutr.*, 58: 1-13.
- Freidman, M. and J.W. Finely. 1971. Methods of tryptophan analysis. *J. Agric. Food Chem.*, 19: 626-631.
- Iqbal, A., I.A. Khalil, N. Ateeq and M.S. Khan. 2006. Nutritional quality of important food legumes. *Food Chem.*, 97: 331-335.
- Kakade, M.L., N. Simons and I.E. Liener. 1969. An evaluation of natural vs synthetic substrates for measuring the antitryptic activity of soybean samples. *Cereal Chem.* 46: 518-528.
- Khalil, L.A. and F.R. Durani. 1990. Haulm and hull of peas as a protein source in animal feed. *Sarhad J. Agric.*, 6: 219-225.
- Nisar, M., M. Qayum, M.R. Shah, H.L. Siddiqui, W.A. Kaleem

and M. Zia-ul-Haq. 2010b. Biological screening of *Impatiens* bicolor royle. Pak. J. Bot., 42(3): 1903-1907.

- Nisar, M., M. Qayum, M.R. Shah, W.A. Kaleem, I. Ali and M. Ziaul-Haq. 2010a. Antimicrobial screening of *Impatiens bicolor royle. Pak. J. Bot.*, 42(1): 523-526.
- Nisar, M., W.A. Kaleem, M. Qayum, A. Hussain, M. Zia-ul-Haq, I. Ali and M.I. Choudhary. 2010c. Biological screening of *Zizyphus* oxyphylla edgew leaves. *Pak. J. Bot.*, 42(6): 4063-4069.
- Nisar, M., W.A. Kaleem, M. Qayum, I.K. Marwat, M. Zia-ul-Haq, I. Ali and M.I. Choudhary. 2011. Biological screening of Zizyphus oxyphylla edgew stem. *Pak. J. Bot.*, 43(1): 311-317.
- Roy, F., J.L. Boye and B.K. Simpson. 2010. Bioactive proteins and peptides in pulse crops: Pea, chickpea and lentil. *Food Res. Int.*, 43: 432-442.
- Ryan, E., K. Galvin, T.P.O. Connor, A.R. Maguire and N.M.O. Brien. 2007. Phytosterol, squalene, tocopherol content and fatty acid, profile of selected seeds, grains, and legumes. *Plant Foods Hum. Nutr.*, 62: 85-91.
- Savage, G.P. 1991. Lentils a forgotten crop. *Outlook Agric.*, 20: 109-112.
- Solanki, I.S., A.C. Kapoor and U. Singh. 1999. Nutritional parameters and yield evaluation of newly developed genotypes of lentil (*Lens culinaris* Medik.), *Plant Foods Hum. Nutr.*, 54: 79-87.
- Taussky, H.H. and E. Shorr. 1953. A microcolorimetric method for the determination of inorganic phosphorus. J. Biol. Chem., 202: 675-682.
- Wang, N., D.W. Hatcher, R. Toews and E.J. Gawalko. 2009. Influence of cooking and dehuling on nutritional composition of several varieties of lentils (*Lens culinaris*), *LWT – Food Sci. Technol.*, 42: 842-848.

- Wheeler, E.I. and R.E. Ferrel. 1971. A method for phytic acid determination in wheat and wheat fractions. *Cereal Chem.*, 48: 312-316.
- Williams, P.C. and U. Singh. 1988. Quality screening and evaluation in pulse breeding. 445-457. In: World Crops: Cool Season Food Legumes. (Ed.): R.J. Summerfield. Kluwer Academic Publishers, Dordrecht The Netherlands.
- Zia-ul-Haq, M., M. Ahmad and M. Akhter. 2010b. Nematicidal activity of indigenous flora of Pakistan. *Pak. J. Bot.*, 43: 1463-1466.
- Zia-ul-Haq, M., M. Ahmad S. Iqbal, S. Ahmad and H. Ali. 2007b. Characterization and compositional studies of oil from seeds of desi chickpea (*Cicer arietinum* L.) cultivars grown in Pakistan. *J. Am. Oil Chem. Soc.*, 84: 1143-1148.
- Zia-ul-Haq, M., S. Ahmad, E. Chiavaro, Mehjabeen and S. Ahmed. 2010a. Studies of oil from cowpea (*Vigna unguiculata* (l) walp.) cultivars commonly grown in Pakistan. *Pak. J. Bot.*, 42(2):214-220.
- Zia-ul-Haq, M., S. Ahmad, M. Ahmad, S. Iqbal and K.M. Khawar. 2009. Effects of cultivar and row spacing on tocopherol and sterol composition of chickpea (*Cicer arietinum* L) seed oil. *Tarim Bilimleri Dergisi*, 15: 25-30.
- Zia-ul-Haq, M., S. Iqbal and M. Ahmad. 2008a. Characteristics of oil from seeds of 4 mungbean (*Vigna radiate* L. wilczek) cultivars grown in Pakistan. J. Am. Oil Chem. Soc., 85: 851-856.
- Zia-ul-Haq, M., S. Iqbal, S. Ahmad, M. I. Bhanger and R. Amarowicz. 2008b. Antioxidant Potential of Desi Chickpea varieties commonly consumed in Pakistan. J. Food Lipid., 15: 26-342.
- Zia-ul-Haq, M., S. Iqbal, S. Ahmad, M. Imran, A. Niaz and M.I. Bhanger. 2007a. Nutritional and compositional study of desi chickpea (*Cicer arietinum* L.) cultivars grown in Punjab, Pakistan. Food Chem., 105: 1357-1363.

(Received for publication 15 February 2010)