STUDIES ON THE EFFICACY OF CHEMICAL AND NON CHEMICAL TREATMENTS TO CONTROL MYCOFLORA ASSOCIATED WITH CHILLI SEED

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Abstract

A total 19 genera and 38 species of fungi were isolated by using standard blotter and deep freezing method from chilli seeds. Of these *Aspergillus flavus*, *A. niger*, *A. fumigatus*, *Alternaria alternata*, *Drechslera hawiinesis*, *Fusarium moniliforme*, *F. oxysporum* and *F. solani* were more frequently isolated. Seed treatments with 8 fungicides viz., Metalaxyl + Mancozeb (72% w/w), Mancozeb (80% w/w Dithiocarbamate), Aliette (80% WP Fosetyle aluminium), Derosol (60% WP Carbendazim), Ridomyl Gold (68% WP), Thiophonate methyl (70% WP), Antracol (70%WP Propineb) and Copperoxychlorite (50% WP) and four seed powders of herbicides viz., asafoetida (*Ferula assafoetida*), black cumin (*Nigella sativa*), neem (*Azadirachta indica*) and mustard (*Brassica campestris*) were used @ 0.5%, 0.15% & 0.25% . Out of these 8 fungicides; Ridomyl Gold @ 0.15 & 0.25% inhibited the growth of all fungi whereas asafetida and *Nigella sativa* powder @0.25% were found to be more efficacious however showed little fungicidal activity toward *Fusarium moniliforme*.

Introduction

In Pakistan chilli (*Capsicum annum* L.) is an important vegetable crop planted over an estimated area of 47.3 thousand hectars with an annual production of 69.5 thousand tones, with an average yield of 1902 kg/ha (Anon., 2006). Chilli is susceptible to several diseases including root and collar rot produces by *Phytophthora capsici* (Ahmed *et al.*, 1989; Saleem *et al.*, 1996; 1998; Hussain *et al.*, 1990; Than *et al.*, 2008). Anthracnose or die-back and fruit rot caused by *Collectotrichum* spp., (Khaleeque & Khan, 1991; Sultana *et al.*, 1992; Amusa, 2004). *Fusarium* spp., produces wilt, root rot and powdery mildew caused by the fungus *Leveillula taurica* (Hafeez, 1986; Mushtaq & Hashmi, 1997). Plant product with antimicrobial properties notably have obtained emphasis for a possible application in food production in order to prevent fungal and bacterial growth (Sagdic *et al.*, 2003; Sridhar *et al.*, 2003; Lanciotti *et al.*, 2004).

Thagaraja & Honso (1996) reported antifungal activity of asafoetida against *Rhizopus sporus, Mucor dimorphosphorous, Penicillium commune* and *Fusarium solani*. Siddiqui *et al.*, (1996) also studied the inhibition effect of asafetida on *A. flavus*. Antifungal activity of asafetida have been also found effective against *Microsporeum gypseum* and *Trichophyton* (Houghton *et al.*, 2006). According to Sitara *et al.*, (2008) 0.15% *Nigella sativa* oil possess a remarkable antifungal activity against *A. flavus, Fusarium moniliforme, F.oxysporum, F.nivale, F. semitectum, Drechslera hawiiensis* and *Alternaria alternata*. The aqueous extract of *N. sativa* seeds has also been shown inhibitory effect on *Candidus albican In vivo* (Khan *et al.*, 2003). Neem is valuable plant sources of medically useful compound and has antimicrobial activities. According

to Agbenin & Marley (2006) dry neem seed extract completely inhibited the mycilial growth of *F. oxysporum* at all concentration. Agbenin *et al.*, (2004) reported that using neem seed powder, also controlled *Fusarium* spp.. Niaz *et al.*, (2008) also analyzed that 1% neem seed oil inhibited the growth of *Drechslera specifera* and *D. hawiinesis*. Sitara *et al.*, (2008) showed that at 0.15% concentration mustard oil had antifungal properties towards *Fusarium oxysporum* and *F. nivale*. According to Bowers & Locke (2000) soil populations of *Fusarium oxysporum* were lowest after 3 to 7 days of incubation when the soil was treated with 5 and 10% aqueous emulsion of mustard extract. Experiments were therefore carried out to examine the mycoflora of chilli seeds and to study the comparisons between fungicides and herbicides.

Materials and Methods

For the detection of seed borne fungi of chilli seed standard blotter and deepfreezing method by ISTA techniques was used. Sodium hypochlorite (10%) was used for surface sterilization of seeds while eight fungicides viz., Metalaxyl + Mancozeb (72%), Mancozeb (72% w/w Dithiocarbamate), Aliette (80% WP Fosetyl aluminium), Derosol (60% WP Carbendazim), Ridomyl Gold (68% WP), Thiophonate methyl (70% WP), Antracol (70% WP Propineb) and Copper Oxychlorite (50% WP) and four powder of herbicides viz., neem, asafoetida, mustard and black cumin were used @ 0.5%, 0.15% & 0.25%.

The eight fungicides and powder of four herbicides were applied on seed in conical flask separately. All treated seed were plated @ 25 seeds/ plate on 3 layers of moistened blotter in 9cm glass Petri plates, incubated at $25\pm1^{\circ}$ C in alternate cycle of 12 hours light and 12 hours darkness for 7 days. In deep-freezing method (Limonard, 1966), the treated and untreated seeds in Petri plates were incubated for one day at $25\pm1^{\circ}$ C then in deep freezer at -4°C for 24 hours. After deep freezing the Petri plates were taken out and incubated for 7 days at $25\pm1^{\circ}$ C. In both methods the growth of fungi were observed after 7 days. The fungi were identified upto species level after reference to Barnet & Hunter (1972), Booth (1971), Ellis (1971) & Nelson *et al.*, (1983).

Results and Discussion

In blotter method, a total 19 genera and 38 species of fungi were isolated. Aspergillus flavus, A. niger, A. fumigatus, Alternaria alternata, Drechslera hawiinesis, Trichoderma sp., Phoma betae, Fusarium moniliforme, Aspergillus candidus and Alternaria tenussima were more frequent in order of prevalence (Table 1). Incidence of Aspergillus species was found to be dominant on chilli seeds. These results fully supported the results obtained by Kiran et al., (2005) and Tripathi et al., (2008) on same seed whereas in deep freezing method Fusarium moniliforme, F. oxysporum, F. solani, F. nivale, A. fumigatus and Drechslera hawiinesis were more frequently isolated. Sultana et al., (1988) reported that infection percent of Fusarium moniliforme and Alternaria alternata were generally higher in the deep-freezing method. Hashmi (1989) also observed that, of the 222 samples of capsicum, 64.0% samples were infected by Fusarium moniliforme and 60.8% by Alternaria alternate.

Sr# Name of fungi 1. Aspergillus niger 2. A.flavus 3. A.candidus 4. A. terrus 5. A. sulphrus 6. A. fumigatus 7. A. terrus 8. Alternata 9. A. tenussinia 10. A. pori 11. A. solani 13. P. lingam	gi iger ternata	Blotter method Nst St Nst St 70.75 47.75 70.75 14.25 37.25 14.25 36.5 13 53.5 33.25	nethod St 46.5 47.75 14.25	Metalay	Metalaxyl + Mancozeb (72%)	ncozeb	~ 5	Mancozeb			Aliette			Derosol	
1. Aspergillus n 2. A.flavus 2. A.flavus 3. A.candidus 4. A.terrus 5. A. sulphrus 6. A. fumigatus 7. A. temari 8. Alternaria al 9. A. tenussinia 10. A. pori 11. A. solami 13. P. lingam	iger ternata	Nst 60.25 70.75 37.25 37.25 36.5 53.5	St 46.5 47.75 14.25				2	(W\W%08)	0		(80%WP)	-	J	(60%WP)	
 Aspergillus n Aspergillus n A. candidus A. candidus A. candidus A. terrus A. terrus A. terrus A. terraria al A. terrasinia A. terrussinia A. terrussinia A. terrussinia A. terrus A. terrus	iger ternata	60.25 70.75 37.25 27.25 36.5 53.5	46.5 47.75 14.25	0.50	0.15	0.25	0.50	0.15	0.25	0.50	0.15	0.25	0.50	0.15	0.25
 A.flavus A.candidus A. terrus A. suphrus A. suphrus A. fumigatus A. temari A. temari A. temaria al A. temasinia <	ternata	70.75 37.25 27.25 36.5 53.5	47.75 14.25	9.75	3.5	1.5	3	1.75	0.75	3	2	-	0	0	0
 A. candidus A. candidus A. terrus A. sulphrus A. fumigatus A. fumigatus A. ternaria al A. ternasinia A. ternasinia A. ternasinia A. ternasinia A. ternasinia A. ternasinia 	ternata	37.25 27.25 36.5 53.5	14.25	4	2	0.75	3.75	2.75	0	2.25	1.5	0	10.8	5.5	5
 A. terrus A. sulphrus A. fumigatus A. fumigatus A. tamari A	ternata	27.25 36.5 53.5		6.25	1.5	0	2	0	0	0	0	0	6	4	0
 A. sulphrus A. fumigatus A. tamari A. tamari B. Alternaria al A. tenussinia A. tenussinia A. pori A. solami Phoma beta P. lingam 	ternata	36.5 53.5	12.5	0	0	0	0	0	0	0	0	0	0	0	0
 A. fumigatus A. tamari A. tamari A. temussinia al A. temussinia A. temussinia A. temussinia A. temus al I. P. lingam 	iernata	53.5	13	0	0	0	0	0	0	0	0	0	0	0	0
 A. tamari Alternaria al A. tenussinia A. pori A. pori A. solani 11. A. solani 12. Phoma beta 13. P. lingam 	ternata		33.25	4	0	0	0	0	0	0	0	0	7.25	5.75	4.75
 8. Alternaria al 9. A. tenussinia 10. A. pori 11. A. solami 12. Phoma beta 13. P. lingam 	ternata	35.5	20.75	0	0	0	0	0	0	2	0	0	0	0	0
 A. tenussinia A. pori A. solani A. solani Phoma beta P. lingam 		50.25	35.25	10	2	0.5	2.5	0	0	0	0	0	6	4.25	5
 A. pori A. solani A. solani Phoma beta P. lingam 		36.75	31.5	6.25	0	0	0	0	0	0	0	0	0	0	0
 A. solani Phoma beta P. lingam 		25.25	15	0	0	0	0	0	0	0	0	0	0	0	0
 Phoma beta P. lingam 		31.5	19.5	0	0	0	0	0	0	0	0	0	0	0	0
13. P. lingam		39.5	23	9	0	0	0	0	0	0	0	0	4.25	0	0
		31.75	19.75	0	0	0	0	0	0	0	0	0	0	0	0
14. Phomopsis		32.25	18.25	0	0	0	0	0	0	0	0	0	5	0	0
15. Phyllosticta sp	di	16	5.5	0	0	0	0	0	0	0	0	0	0	0	0
16. Chaetomium globossom	globossom	33.75	17.75	8.25	0	0	0	0	0	0	0	0	4.75	1	0
17. C. gracile		13.75	7	0	0	0	0	0	0	0	0	0	0	0	0
18. C. distortum		12	5.75	0	0	0	0	0	0	0	0	0	0	0	0
19. Cladosporium caldosporoides		26.25	6.5	0	0	0	0	0	0	0	0	0	6.75	5	3.5

CONTROL OF MYCOFLORA ASSOCIATED WITH CHILLI SEED

						Table 1. (Cont'd.).	Cont'd.).								
#IS	Sr# Name of fungi	Blotter	Blotter method	Metala	Metalaxyl + Mancozeb (72%)	ncozeb	- 3	Mancozeb (80%W\W)			Aliette (80%WP)			Derosol (60%WP)	
		Nst	St	0.50	0.15	0.25	0.50	0.15	0.25	0.50	0.15	0.25	0.50	0.15	0.25
20.	20. Curvularia pallescens	14	6.75	0	0	0	0	0	0	0	0	0	0	0	0
21.	21. C. lunata	Ш	4.25	0	0	0	0	0	0	0	0	0	0	0	0
22.	22. C. robusta	18	55	0	0	0	0	0	0	0	0	0	0	0	0
23.	23. Drechslera hawiinesis	50.5	26	8.75	1.5	0	2	1.5	0	0	0	0	7.75	0	0
24.	24. D. specifera	32.75	19.5	0	0	0	1.5	0	0	0	0	0	0	0	0
25.	D. poae	29.5	10.5	8.25	0	0	0	0	0	0	0	0	0	0	0
26.	26. Rhizopus sp	32.5	21.75	0	0	0	0	0	0	0	0	0	0	0	0
27.	27. Rhizoctonia solani	30.5	19	0	0	0	0	0	0	0	0	0	0	0	0
28.	F. monilifrme	38.5	16	12.3	б	0	3.5	0	0	1	0	0	6.5	4.75	3.5
29.	F. oxysporum	29.75	10.75	9.5	0	0	4.25	0	0	0	0	0	0	0	0
30.	F. nivale	0	0	0	0	0	0	0	0	0	0	0	0	0	0
31.	F. solani	0	0	7	1	0	0	0	0	0	0	0	6.25	4.75	3.75
32.	32. Helminthosporium valutinum	15.5	6.25	4	2	0	0	0	0	0	0	0	0	0	0
33.	33. Nigrospora sp	19	9.5	0	0	0	0	0	0	0	0	0	1.5	0	0
34.	Cercospora sp	26.5	14.75	0	0	0	0	0	0	0	0	0	0	0	0
35.	35. Cephaliophora irreglaris	14.25	8	0	0	0	0	0	0	0	0	0	0	0	0
36.	36. Myrothecium straiatispor	20	9	0	0	0	0	0	0	0	0	0	0	0	0
37.	37. M. brachysporum	19.5	8	0	0	0	0	0	0	0	0	0	0	0	0
38.	38. Collectotrichum capsci	27	12.25	9.5	1.75	0	0	0	0	0	0	0	4.25	0	0
39.	39. Trichoderma	39.75	20.5	10	-	0	0	0	0	0	0	0	2.5	0	0

Table 1. (Cont'd.).

					I able 1. (Cont d.).	(- n 1110-)							
Sr#	Name of fungi	Ri	Ridomyl Gold (68%WP)	р	Thio	Thiophonate methyl (70%WP)	ethyl		Antracol (70%WP)		Copp	Copper Oxychlorite (50%WP)	orite
		0.50	0.15	0.25	0.50	0.15	0.25	0.50	0.15	0.25	0.50	0.15	0.25
-I	Aspergillus niger	1.5	0.5	0	8.25	4.5	2.25	9.25	6.25	4.25	11.3	7.8	3.75
2.	A.flavus	0.5	0	0	4.5	2	0	8.5	4	2.75	9.5	4.8	4
3.	A.candidus	0	0	0	0	0	0	0	0	0	8	6.8	0
4.	A. terrus	0	0	0	0	0	0	0	0	0	0	0	0
5.	A. sulphrus	0	0	0	0	0	0	0	0	0	0	0	0
.9	A. fumigatus	0	0	0	3.5	2	0	0	0	0	0	0	0
7.	A. tamari	0	0	0	0	0	0	7	0	0	0	0	0
8	Alternaria alternata	0	0	0	4	2.25	0	8.25	3.75	0	6.75	5.8	3
9.	A. tenussinia	0	0	0	0	0	0	0	0	0	0	0	0
10.	A. pori	0	0	0	0	0	0	7.5	0	0	0	0	0
11.	A. solani	0	0	0	0	0	0	0	0	0	0	0	0
12.	Phoma beta	0	0	0	0	0	0	0	0	0	5	3	0
13.	P. lingam	0	0	0	0	0	0	0	0	0	0	0	0
14.	Phomopsis	0	0	0	0	0	0	0	0	0	0	0	0
15.	Phyllosticta sp	0	0	0	0	0	0	0	0	0	0	0	0
16.	Chaetomium globossom	0	0	0	0	0	0	0	0	0	7.75	0	0
17.	C. gracile	0	0	0	0	0	0	0	0	0	0	0	0
18.	C. distortum	0	0	0	0	0	0	0	0	0	0	0	0
19.	Cladosporium caldosporoides	0	0	0	7.25	0	0	0	0	0	10	0	0

CONTROL OF MYCOFLORA ASSOCIATED WITH CHILLI SEED

					Table 1.	Table 1. (Cont'd.).							
\$r#	Name of fungi	R	Ridomyl Gold (68%WP)	p	Thiol	Thiophonate methyl (70%WP)	ethyl		Antracol (70%WP)		Copp	Copper Oxychlorite (50%WP)	orite
		0.50	0.15	0.25	0.50	0.15	0.25	0.50	0.15	0.25	0.50	0.15	0.25
20.	Curvularia pallescens	0	0	0	0	0	0	0	0	0	0	0	0
21.	C. lunata	0	0	0	0	0	0	0	0	0	0	0	0
22.	C. robusta	0	0	0	0	0	0	0	0	0	0	0	0
23.	Drechslera hawiinesis	0	0	0	0	0	0	0	0	0	5.25	5.3	2.25
24.	D. specifera	0	0	0	0	0	0	0	0	0	5.25	0	0
25.	D. poae	0	0	0	0	0	0	0	0	0	0	0	0
26.	Rhizopus sp	0	0	0	0	0	0	0	0	0	0	0	0
27.	Rhizoctonia solani	0	0	0	0	0	0	0	0	0	0	0	0
28.	F. monilifrme	1.5	0	0	9.75	0	0	11.3	10	8	14.3	7.5	3.75
29.	F. oxysporum	0	0	0	0	0	0	0	0	0	8.25	5.5	4.25
30.	F. nivale	0	0	0	0	0	0	0	0	0	0	0	0
31.	F. solani	0	0	0	0	0	0	10.5	7.25	0	7.25	0	0
32.	Helminthosporium valutinum	0	0	0	0	0	0	0	0	0	3.75	0	0
33.	Nigrospora sp	0	0	0	4.25	0	0	0	0	0	0	0	0
34.	Cercospora sp	0	0	0	0	0	0	0	0	0	0	0	0
35.	Cephaliophora irreglaris	0	0	0	0	0	0	0	0	0	0	0	0
36.	Myrothecium straiatispor	0	0	0	0	0	0	0	0	0	0	0	0
37.	M. brachysporum	0	0	0	0	0	0	0	0	0	0	0	0
38.	Collectotrichum capsci	0	0	0	0	0	0	0	0	0	7.5	4.5	0
39.	Trichoderma	0	0	0	0	0	0	0	0	0	5.75	2.5	0

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Eight fungicides and four herbicides were used to control the fungus associated with the seed of chilli @ 0.5%, 0.15% & 0.25% concentration. Results showed that out of all fungicides, Ridomyl Gold (68% WP) @ 0.25% concentration completely controlled the fungi; however, 0.15% dose also suppressed the growth of all fungi whereas *A. niger* showed only 0.5% growth. This result is in close conformity with the findings of Sitara & Shahida (2007) who found that Ridomyl Gold was effective @ 0.3% concentration against all fungi except *A. niger*. Fungicides Mancozeb, Aliette, Thiophonate methyl @ 0.25% also controlled all isolated fungi whereas *A. niger* gave 0.75%, 1% & 2.25% mycelial growth respectively. Aliette @0.15% showed better result as compare to Mancozeb and Thiophonate methyl at same doses. It is also interesting to note that Derosol (60% WP) completely inhibited the growth of fungi at all doses in blotter and deep freezing method.

In deep freezing method three fungicides viz., Ridomyl Gold, Thiophonate methyl and Mancozeb @ 0.25% concentration completely controlled all fungi whereas Thiophonate methyl and Aliette were effective for all fungi nevertheless the growth of *A. niger* and *A. flavus* increased to some extent (Table 2). It is also noted that fungi viz., *Phoma lingam, Phomopsis* sp., *Phyllosticta* sp., *Cheatomium gracile, C. distortum, Curvularia pallescens, C. lunata, C. robusta, Rhizopus* sp., *Rhizoctonia solani, Cercospora* sp., *Cephalophora irregularis, Myrothecium straiatispor* and *M. brachysporum* were completely inhibited by all fungicides in blotter and deep freezing methods. The growth of *Fusarium nivale* and *Nigrospora* sp., promoted only in deep freezing method.

Antifungal activity of asafoetida, Nigella sativa, neem and mustard seed powder were analyzed by blotter and deep freezing method against all isolated fungi. The results revealed that asafoetida @ 0.25% showed positive response for all fungi except the growth of F. moniliforme (Table 3). Siddiqui et al., (1996) also reported antifungal activity of asafoetida oil against A. flavus. According to Sitara et al., (2008), asafoetida oil @ 0.1% & 0.15% significantly inhibited the growth all tested fungi except A. flavus. Antifungal activity of Nigella sativa was most significantly effective @ 0.25 %; however, it exhibited no fungicidal activity against A. flavus and Fusarium moniliforme. The oil extract of Nigella sativa showed antimicrobial effect In vivo towards Staphylococcus aurous, Pseudomonas aeroginosa and Candida albicans (Hanafy & Hatem, 1991; Mashhadian & Rakshandeh, 2005). Neem oil was effective @ 0.15% and 0.25%; nevertheless the growth of A. flavus, A. niger, Fusarium moniliforme and F. solani were not reduced. Ishrat et al., (2008) found that 0.1% concentration of neem oil was effective against for Macrophomina phaseolina and Rhizoctonia solani. Kazmi et al., (1995) also noted that 0.1% neem seed oil was more effective against Macrophomina phaseolina. Mustard oil @ 0.25% concentration also showed significant growth reduction in all isolated fungi, moreover, the growth of Aspergillus flavus, A. niger, Alternaria alternata, F. moniliforme and F. solani was somewhat repressed or promoted. Kazmi et al., (1993) also reported fungistatic activity of mustard oil, most significantly against Alternaria alternata as compare to other fungi.

	Table 2.	. Percent	age occur	rence of	fungi afte	er differe	Table 2. Percentage occurrence of fungi after different treatments of fungicides by deep freezing method.	ents of fu	ngicides	by deep f	reezing n	nethod.			
Sr#	Sr# Name of fungi	Blotter method	method	Metala	Metalaxyl + Mancozeb (72%)	ncozeb	N 8)	Mancozeb (80%W\W)			Aliette (80%WP)			Derosol (60%WP)	
		Nst	St	0.50	0.15	0.25	0.50	0.15	0.25	0.50	0.15	0.25	0.50	0.15	0.25
Ι.	Aspergillus niger	41.5	31.25	2.75	1.5	0.5	1.5	1	0	2.5	1.5	1.5	0	0	0
2.	A.flavus	37.25	25	2.5	1	0.5	2	1.5	0	1.25	0.5	0	7.5	4	1.5
3.	A. candidus	20.25	7.5	4.75	0.5	0	1.75	0	0	0	0	0	6.25	2	0
4.	A. terrus	18.25	8	0	0	0	0	0	0	0	0	0	0	0	0
5.	A. sulphrus	20	11.75	0	0	0	0	0	0	0	0	0	0	0	0
6.	A. fumigatus	40.5	30.25	2	0	0	0	0	0	0	0	0	4	3.75	3.5
7.	A. tamari	28.25	13.75	0	0	0	0	0	0	1.5	0	0	0	0	0
8.	Alternaria alternata	34.75	23	3.75	1.25	0.25	1.25	0	0	0	0	0	1.75	4.25	0
9.	A. tenussinia	31	24.5	4.75	0	0	0	0	0	0	0	0	0	0	0
10.	A. pori	25.75	14.25	0	0	0	0	0	0	0	0	0	0	0	0
11.	A. solani	27.25	15.75	0	0	0	0	0	0	0	0	0	0	0	0
12.	Phoma beta	36.5	18.75	3.5	0	0	0	0	0	0	0	0	3	0	0
13.	P. lingam	27.25	14	0	0	0	0	0	0	0	0	0	0	0	0
14.	Phomopsis	29.75	13.5	0	0	0	0	0	0	0	0	0	0	0	0
15.	Phyllosticta sp	10	5	0	0	0	0	0	0	0	0	0	0	0	0
16.	Chaetomium globossom	20	9.25	4.5	0	0	0	0	0	0	0	0	3	0.5	0
17.	C. gracile	9.25	4	0	0	0	0	0	0	0	0	0	0	0	0
18.	C. distortum	7	4	0	0	0	0	0	0	0	0	0	0	0	0
19.	19. Cladosporium caldosporoides	14.5	3.75	0	0	0	0	0	0	0	0	0	3.75	3.75	3

					L	Table 2. (Cont'd.).	Cont'd.).								
Sr#	Sr# Name of fungi	Blotter method	method	Metala	Metalaxyl + Mancozeb (72%)	ncozeb	N 8)	Mancozeb (80%W\W)		2	Aliette (80%WP)			Derosol (60%WP)	
		Nst	St	0.50	0.15	0.25	0.50	0.15	0.25	0.50	0.15	0.25	0.50	0.15	0.25
20.	Curvularia pallescens	8.75	4.25	0	0	0	0	0	0	0	0	0	0	0	0
21.	C. lunata	6.25	3.75	0	0	0	0	0	0	0	0	0	0	0	0
22.	C. robusta	10.75	3.75	0	0	0	0	0	0	0	0	0	0	0	0
23.	Drechslera hawiinesis	38	22.25	4.5	0.75	0	0.75	0.5	0	0	0	0	6.25	0	0
24.	D. specifera	33	20	0	0	0	1.25	0	0	0	0	0	0	0	0
25.	D. poae	18.75	8.25	8.25	0	0	0	0	0	0	0	0	0	0	0
26.	Rhizopus sp	23	13.75	0	0	0	0	0	0	0	0	0	0	0	0
27.	Rhizoctonia solani	25	16	0	0	0	0	0	0	0	0	0	0	0	0
28.	F. monilifrme	72.5	46.5	6.5	1	0	2	0	0	1	0	0	3.75	3	3.75
29.	F. oxysporum	68.75	46	4.5	0	0	1.75	0	0	0	0	0	0	0	0
30.	F. nivale	43.25	22.5	0	0	0	0	0	0	0	0	0	0	0	0
31.	F. solani	52	31	3.5	0.5	0	0	0	0	0	0	0	3.75	0	4.75
32.	Helminthosporium valutinum	10	4.75	2	0.75	0	0	0	0	0	0	0	0	0	0
33.	Nigrospora sp	13.5	6.75	0	0	0	0	0	0	0	0	0	0	0	0
34.	Cercospora sp	21	11	0	0	0	0	0	0	0	0	0	0	0	0
35.	Cephaliophora irreglaris	8	2.5	0	0	0	0	0	0	0	0	0	0	0	0
36.	Myrothecium straiatispor	10.5	5	0	0	0	0	0	0	0	0	0	0	0	0
37.	M. brachysporum	13	5	0	0	0	0	0	0	0	0	0	0	0	0
38.	Collectotrichum capsci	17.5	10.5	3.25	0.5	0	0	0	0	0	0	0	0	0	0
39.	Trichoderma	29.5	21.25	4.5	0.75	0	0	0	0	0	0	0	8	0	0

					Table 2.	Table 2. (Cont'd.).							
#IS	Name of fungi	Я	Ridomyl Gold (68%WP)	PI	Thio	Thiophonate methyl (70%WP)	ethyl		Antracol (70%WP)		Copp	Copper Oxychlorite (50%WP)	orite
		0.50	0.15	0.25	0.50	0.15	0.25	0.50	0.15	0.25	0.50	0.15	0.25
	Aspergillus niger	0.75	0.25	0	5	3.75	0	10.3	5	3	7.25	3.5	2.5
2.	A flavus	0	0	0	2.25	0.75	0	3.5	2.75	1.75	5.25	3	С
3.	A.candidus	0	0	0	0	0	0	0	0	0	4.25	4	0
4.	A. terrus	0	0	0	0	0	0	0	0	0	0	0	0
5.	A. sulphrus	0	0	0	0	0	0	0	0	0	0	0	0
.9	A. fumigatus	0	0	0	2	0	0	0	0	0	0	0	0
7.	A. tamari	0	0	0	0	0	0	0	0	0	0	0	0
8.	Alternaria alternata	0	0	0	3.25	0	0	4.5	3.25	0	3.75	3.5	1.75
9.	A. tenussinia	0	0	0	0	0	0	0	0	0	0	0	0
10.	A. pori	0	0	0	0	0	0	0	0	0	0	0	0
11.	A. solani	0	0	0	0	0	0	0	0	0	0	0	0
12.	Phoma beta	0	0	0	0	0	0	0	0	0	0	0	0
13.	P. lingam	0	0	0	0	0	0	0	0	0	0	0	0
14.	Phomopsis	0	0	0	0	0	0	0	0	0	0	0	0
15.	Phyllosticta sp	0	0	0	0	0	0	0	0	0	0	0	0
16.	Chaetomium globossom	0	0	0	0	0	0	0	0	0	0	0	0
17.	C. gracile	0	0	0	0	0	0	0	0	0	0	0	0
18.	C. distortum	0	0	0	0	0	0	0	0	0	0	0	0
19.	Cladosporium caldosporoides	12	0	0	2.5	0	0	0	0	0	0	0	0

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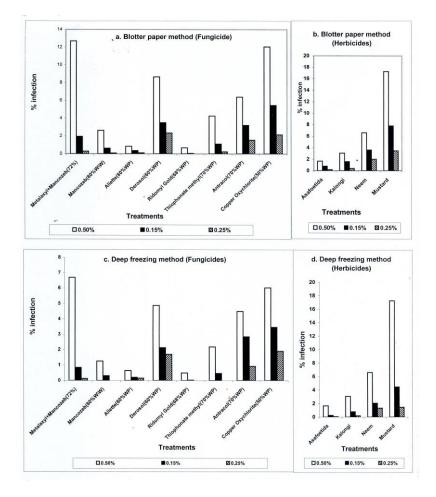
					Table 2.	Table 2. (Cont'd.).							
\$r#	Name of fungi	R	Ridomyl Gold (68%WP)	ld	Thio	Thiophonate methyl (70%WP)	ethyl		Antracol (70%WP)		Copp	Copper Oxychlorite (50%WP)	orite
3		0.50	0.15	0.25	0.50	0.15	0.25	0.50	0.15	0.25	0.50	0.15	0.25
20.	Curvularia pallescens	0	0	0	0	0	0	0	0	0	0	0	0
21.	C. lunata	0	0	0	0	0	0	0	0	0	0	0	0
22.	C. robusta	0	0	0	0	0	0	0	0	0	0	0	0
23.	Drechslera hawiinesis	0	0	0	0	0	0	0	0	0	4.5	4.5	1.5
24.	D. specifera	0	0	0	0	0	0	0	0	0	0	0	0
25.	D. poae	0	0	0	0	0	0	0	0	0	0	0	0
26.	Rhizopus sp	0	0	0	0	0	0	0	0	0	0	0	0
27.	Rhizoctonia solani	0	0	0	0	0	0	0	0	0	0	0	0
28.	F. monilifrme	1	0	0	6.25	0	0	11	9.75	4.25	14.3	9.3	5
29.	F. oxysporum	0	0	0	0	0	0	0	0	0	11	3.5	4.75
30.	F. nivale	0	0	0	0	0	0	0	0	0	0	0	0
31.	F. solani	0	0	0	0	0	0	14.5	L	0	0	0	0
32.	Helminthosporium valutinum	0	0	0	0	0	0	0	0	0	0	0	0
33.	Nigrospora sp	0	0	0	0	0	0	0	0	0	0	0	0
34.	Cercospora sp	0	0	0	0	0	0	0	0	0	0	0	0
35.	Cephaliophora irreglaris	0	0	0	0	0	0	0	0	0	0	0	0
36.	Myrothecium straiatispor	0	0	0	0	0	0	0	0	0	0	0	0
37.	M. brachysporum	0	0	0	0	0	0	0	0	0	0	0	0
38.	Collectotrichum capsci	0	0	0	0	0	0	0	0	0	3.75	2.5	0
39.	Trichoderma	0	0	0	0	0	0	0	0	0	2.75	0	0

	Table 3.	% Occu	Table 3. % Occurrence of fungi after different treatments (herbicides) by blotter method	fungi aft	er differe	ent treatn	nents (her	bicides) l	by blotter	method.			
C M.O		Ł	Asafoetida	-		Kalongi			Neem			Mustard	
ONTO	S.NO Name of fungi	0.50%	0.15%	0.25%	0.50%	0.15%	0.25%	0.50%	0.15%	0.25%	0.50%	0.15%	0.25%
I.	Aspergillus flavus	2	-	0	3.25	1.75	0.5	3.75	3	2.25	7	5.5	2.5
2.	A.niger	1.5	0.5	0	2.5	1.75	0	5	4	3	6.25	5.75	4
З.	A.fumigatus	0	0	0	0	0	0	2.5	1.5	0	4.5	3	0
4.	A. candidus	0	0	0	0	0	0	0	0	0	4.5	0	0
5.	Alternaria alternata	0.5	0	0	1.5	1	0	2	1.25	0	9	4	2
6.	A. tennius	0	0	0	0	0	0	2.5	0	0	2.5	0	0
7.	A.solani	0	0	0	0	0	0	0	0	0	4.75	0	0
8.	A. tenussinia	0	0	0	1.75	0	0	0	0	0	4	0	0
9.	F. monilifrme	2.25	1.5	0.75	2.5	1.5	1	4.5	1.75	1	5.75	4.5	2.25
10.	F. solani	0	0	0	0	0	0	2.5	7	1.25	4	2.25	-
11.	F. oxysporum	0	0	0	0	0	0	2	0	0	4.5	2.5	0
12.	Cladosporium cladosporidi	0	0	0	0	0	0	0	0	0	2	0	0
13.	collectotrichum	0	0	0	0	0	0	0	0	0	3.5	0	0

C N.S		ł	Asafoetida	a		Kalongi			Neem			Mustard	
01.0	S.No Name of fungi	0.50%	0.15%	0.25%	0.50%	0.15%	0.25%	0.50%	0.15%	0.25%	0.50%	0.15%	0.25%
1.	Aspergillus flavus	1.5	0.5	0	2	1.25	0	3	2	1.75	4.5	4	1.5
2.	A.niger	1	0	0	1.75	0.75	0	4	3	2.25	9	4.75	2.25
3.	A.fumigatus	0	0	0	0	0	0	0	0	0	3	1.75	0
4.	A. candidus	0	0	0	0	0	0	0	0	0	3.5	0	0
5.	Alternaria alternata	0	0	0	1	0	0	1.5	0	0	3	2.25	0
6.	A.solani	0	0	0	0	0	0	0	0	0	3	0	0
7.	A. tenussinia	0	0	0	1	0	0	0	0	0	0	0	0
8.	F. monilifrme	1.75	0.5	0.25	2.25	1	0.75	3.5	1.5	0.5	4.5	2.75	1.25
9.	F. solani	0	0	0	0	0	0	2	1.25	0.5	3	1.25	0.5
10.	F. oxysporum	0	0	0	0	0	0	0	0	0	3.5	0	0
11.	collectotrichum	0	0	0	0	0	0	0	0	0	2.25	0	0
12.	Drechslera hawiinesis	0	0	0	0	0	0	0	0	0	2	0	0

Table 4. % Occurrence of fungi at different treatments (deep freezing method).

In deep freezing method, asafoetida powder @ 0.5 & 0.15% was effective against for all isolated fungi; however Aspergillus flavus, A. niger and Fusarium moniliforme were not controled whereas 0.25% concentration also inhibited the growth of all fungi except F. moniliforme. Remarkably Nigella sativa powder @ 0.25% controlled all fungi; however Fusarium moniliforme showed little growth (Table 4). Neem seed powder also inhibited the growth of same fungi as in blotter method. Mustard powder @ 0.15% controlled F. oxysporum whereas in blotter method the growth of Fusarium oxysporum was somewhat promoted. It is also noted that 0.50% mustard powder inhibited the growth of Drechslera hawiinesis in blotter method nevertheless the growth was not much reduced in deepfreezing method. Sumbali & Mehrotra (1980) also analyzed the control of Macrophomina phaseolina by using mustard oil. Metalexyl + Mancozeb, Derosol, Copper Oxychlorite, and Antracol were not effective (a) 0.5% concentration whereas Mancozeb, Aliette and Ridomyl Gold reduced infection percent at same dose level (Fig. 1a). Infection percent is high in Mustard seed powder @ 0.5 & 0.15% concentration whereas growth inhibited in Asafoetida and Kalongi @ 0.25% concentration. All herbicides were effective at 0.25% concentration and compete well with fungicides (Fig. 1b).



Sr No	Treatments	0.5 Mean ± Std.	0.15 Mean ±	0.25 Mean ±
51.110	Treatments	Error	Std. Error	Std. Error
1.	Metalaxyl + Mancozeb (72%)	12.692 ± 2.662	1.948 ± 0.594	0.282 ± 0.175
2.	Mancozeb (80%W\W)	2.615 ± 0.864	0.615 ± 0.359	0.076 ± 0.076
3.	Aliette (80%WP)	0.846 ± 0.432	0.358 ± 0.253	0.1025 ± 0.1025
4.	Derosol (60%WP)	8.641 ± 2.094	3.487 ± 1.1931	2.333 ± 0.925
5.	Ridomyl Gold (68%WP)	0.666 ± 0.369	0.051 ± 0.051	0
6.	Thiophonate methyl (70%WP)	4.256 ± 1.6032	1.1025 ± 0.572	0.230 ± 0.230
7.	Antracol (70%WP)	6.384 ± 2.249	3.205 ± 1.4570	1.538 ± 0.951
8.	Copper Oxychlorite (50%WP)	12.052 ± 2.658	5.461 ± 1.6033	2.153 ± 0.837
9.	Asafoetida	1.666 ± 0.826	0.8 ± 0.4700	0.2 ± 0.2
10.	Kalongi	3.066 ± 1.220	1.6 ± 0.728	0.4 ± 0.2894
11.	Neem	6.6 ± 1.856	3.6 ± 1.3444	2.0 ± 0.9904
12.	Mustard	17.26 ± 1.528	7.8 ± 2.240	3.466±1.312

Fig. 1. Effect of fungicides and herbicides on the growth of fungi in blotter and deep freezing method. Table 5a. Mean and standard error of fungicides and herbicides (Blotter paper method).

Table 5b. Mean and standard error of fungicides and herbicides (Deep freezing method)

Sr.No	Treatments	0.5 Mean ± Std. Error	0.15 Mean ± Std. Error	0.25 Mean ± Std. Error
1.	Metalaxyl + Mancozeb (72%)	6.692 ± 1.456	0.8461 ± 0.2611	0.128 ± 0.075
2.	Mancozeb (80%W\W)	1.256 ± 0.418	0.307 ± 0.187	0
3.	Aliette (80%WP)	0.641 ± 0.329	0.205 ± 0.1608	0.153 ± 0.153
4.	Derosol (60%WP)	4.871 ± 1.362	2.128 ± 0.807	1.692 ± 0.757
5.	Ridomyl Gold (68%WP)	0.487 ± 0.328	0.025 ± 0.025	0
6.	Thiophonate methyl (70%WP)	2.179 ± 0.191	0.461 ± 0.390	0
7.	Antracol (70%WP)	4.487 ± 2.128	2.846 ± 1.344	0.923 ± 0.550
8.	Copper Oxychlorite (50%WP)	6.02 ± 2.150	3.461 ± 1.249	1.897 ± 0.800
9.	Asafoetida	1.666 ± 0.826	0.2666 ± 0.1817	0.066 ± 0.066
10.	Kalongi	3.066 ± 1.220	0.8 ± 0.438	0.2 ± 0.2
11.	Neem	6.6 ± 1.856	2.066 ± 0.987	1.333 ± 0.728
12.	Mustard	17.266 ± 1.528	4.466 ± 1.6814	1.466 ± 0.735

In deep freezing method fungicides @ 0.25% concentration showed low infection percent compare to blotter method. Metalaxyl + Mancozeb & Aliette also suppressed the fungal growth @ 0.15% concentration (Fig. 1c). *Asafoetida* and *Kalongi* posses strong antifungal activity at all doses levels followed by neem and mustard (Fig. 1d).

Statistical analysis of fungicides and herbicides revealed that Ridomyl Gold (68% WP) was found to be most effective at all dose levels followed by Mancozeb (80% W/W), Aliette (80% WP) and asafoetida @ 0.25% inhibited mycelial growth of fungi as compare to other treatments in blotter paper method (Table 5a) whereas in deep freezing method Mean & Std. error showed that Ridomyl Gold, Mancozeb and Thiophonate methyl controlled all fungal flora asafoetida possess strong fungicidal effect @ 0.25% (Table 5b). Analysis of variance to compare fungicides and herbicides at 0.01 level of significance; showed significant differences at all level i.e., 0.50% (p<0.01), 0.1% (p<0.01) & 0.25% (p<0.01). The results showed that fungicide Ridomyl Gold (68% WP) and herbicide asafoetida @ 0.25% were more effective and showed strong fungicidal activity towards isolated fungi.

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