SALINITY TOLERANCE AND RECLAMATION POTENTIAL OF TWO WIDELY DISTRIBUTED SUBTROPICAL TREE SPECIES

ARSLAN ARIF1*, IRFAN ASHRAF1, MUHAMMAD SAFDAR HUSSAIN2 AND RASHID IQBAL3

¹Department of Forestry, Range and Wildlife Management, Faculty of Agriculture and Environment,

The Islamia University of Bahawalpur, Pakistan

²Department of Forestry & Range Management, Kohsar University Murree, Pakistan

³ Department of Agronomy, Faculty of Agriculture and Environment, The Islamia University of Bahawalpur, Pakistan

*Corresponding author's email: arslanarif.frw.iub@gmail.com, safdarhussain@kum.edu.pk

Abstract

Soil salinity threatens land productivity and food security, particularly in arid and semi-arid regions worldwide. Phytoremediation is a cost-effective and eco-friendly approach for mitigating salinity impacts in irrigated agricultural landscapes. In this regard, a pot experiment study was conducted to evaluate the salt tolerance and reclamation potential of two commonly planted tree species in subtropical regions globally, *Eucalyptus camaldulensis* and *Syzygium cumini*, under various salinity treatments: Control, 8 dS m⁻¹, and 16 dS m⁻¹. Results revealed that salinity negatively affected the growth and biomass production of both tested species, with more prominent effects on *S. cumini*. *E. camaldulensis* exhibited the lowest reduction in shoot height (22%) and dry biomass (40%), while *S. cumini* showed a 30% and 53% decrease in both traits, respectively, under 16 dS m⁻¹ salinity. Correspondingly, increased antioxidant enzyme activities became a defense mechanism in *E. camaldulensis* as peroxidase (2.51–8.41 U mg⁻¹ protein), superoxide dismutase (3.12–11.35 U mg⁻¹ protein), and catalase (38–83.21 U mg⁻¹ protein) at 16 dS m⁻¹. Moreover, *E. camaldulensis* reduced the highest levels of soil SAR and ECe by maintaining the optimum Na, Cl, and K ratios. Thus, the present study suggests that *E. camaldulensis* maintained higher growth and exhibited an effective antioxidant defense mechanism under various salinity levels, suggesting higher potential for the amelioration of saline soils. These results demonstrate a quick and powerful screening mechanism to assess potential reclamation tree species for saline environments.

Key-words: Pakistan, Food insecurity, Tree species, Afforestation, Semi-arid, Woody biomass production, Bioremediation

Introduction

Globally, the demands for food and other agricultural products are expanding with the increase in population, which is expected to reach around 8 billion in 2025 (Jesus et al., 2015). To cope with food insecurity under the Sustainable Development Goals (SDG), there is a dire need to increase the area under agricultural productivity by cultivating presently salinity-impacted unproductive lands in arid and semi-arid regions across the (Pandey & Pandey, 2023). Salinity is an ever-increasing phenomenon in irrigated agricultural landscapes and one of the most common forms of soil degradation, which is threatening land productivity, food security, and rural livelihoods, particularly in developing countries (Munns 2002; Nawar et al., 2015). Annually, about 1-2% of irrigated arable land is decreasing continuously and causing serious threats to the agroecosystem worldwide (Anon., 2017). About 10–20 million people's livelihoods are destroyed due to salinization and its devastating effects on the natural ecosystem (Vivekananda et al., 2014).

Salinity suppresses plant growth and development mechanisms (Roy et al., 2014) and plants respond to salt stress depends on through morphological and physiological adjustments (Ma et al., 2020). Under severe salt stress conditions, plants face reduced water uptake from soil resulting in osmotic stress (Flexas et al., 2004). Salt stress also causes ion toxicity and dehydration in the plant cell (Munns, 2002). Consequently, plant metabolic processes inhibition and growth retardation are often observed following cellular dehydration (Leprince et al., 2000). Soil salinity also affects photosynthesis and stomatal closure in

plants resulting in the decline of photosynthetic activity (Brugnoli & Lauteri, 1991). Reduction in plant osmotic potential during salt stress may also cause a decline in cell expansion which ultimately leads to a reduction in photosynthesis activity (Lautner, 2013).

Managing salinity is one of the biggest challenges in the Indus Basin areas of Pakistan as salinity hinders crop growth and reduces agricultural productivity of food for human consumptions. Phytoremediation is a cost-effective and eco-friendly way to reclaim salt-affected soils (Zaghloul, 2020). In this scenario, plants have been proved to serve as natural pumps to draw salts and other contaminants from the soil and, in time, restore productivity of impacted lands (Nouri et al., 2017). Tree species are preferred over ephemeral plants due to highest biomass production and reclaiming salinity to a significant level along with providing economic returns (Prasad, 2011). Therefore, in the context where reclamation of saltaffected soils becomes crucial, exploring suitable tree species for planting in high salinity landscapes becomes imperative (Kumar et al., 2021).

Syzygium cumini is widely used medicinal tree which is commonly used for the treatment of various diseases such as diabetes. Being a multipurpose tree species, S. cumini has significant importance of antioxidant and antibacterial properties (Banerjee et al., 2005; Shafi et al., 2002). Extracts of S. cumini dry leaves is primary used in biological control of weeds in agricultural lands (Patil et al., 1996; Shafique et al., 2005). Similarly, Eucalyptus (Sufaida) is also important tree species because of high rate of growth, environmental adaptability and superior pulp properties

2 ARSLAN ARIF ETAL.,

throughout the world (Stape *et al.*, 2004). It is an evergreen tree known to extract water from great depths relative to other species and is regarded to be salt-tolerant tree. However, the salinity tolerance potential of both tree species has not been elucidated yet for the reclamation of salinity. The present study aims to test the responses of *E. camaldulensis* and *S. cumini* to different salinity levels on growth, antioxidant enzyme activities and ion accumulation in various plant organs. Comparison of these parameters in the tested tree species may help to develop a better understanding and provide additional information on the salt tolerance mechanisms of *E. camaldulensis* and *S. cumini* for planting in salinity impaired soils.

Material and Methods

The study was carried out from September 2019 to September 2020 as a pot experiment excluding the sixmonth plant establishment period under greenhouse conditions at the experimental site of the Department of Forestry, The Islamia University Bahawalpur (IUB), Punjab Pakistan (29° 22 N, 71° 45 E; 126 m).

Preparation of soil and planting materials: Fine-grained soil consisting of silt and clay as per the Unified Soil Classification System was collected from the experimental site of the Department of Forestry. Subsequently, before being placed into polythene bags, the soil was sieved using 2 mm sieve and mixed with peat and sand in a 1:1 ratio. Thirty polythene bags were filled with prepared soil for establishing the plants. Similarly, seeds of S. cumini and E. camaldulensis were obtained from Punjab Forest Research Institute Faisalabad Gatwala. To reduce the risk of fungal and other disease attacks, seeds were sterilized in sodium hypochlorite (20%) for 20 minutes. Finally, three seeds were sown in each plastic bag and irrigated daily with a sprinkler to facilitate germination. Two weeks following seedling emergence, thinning was performed and one healthy plant in each bag was left to grow under ambient conditions in the greenhouse for 6 months.

Experimental procedure: The soil was again collected from the same site and prepared for pot filling. Collected soil was ground sieved through a 2 mm sieve to remove undesirable materials such as weeds and stones if any. Subsequently, three samples were taken from the sieved soil and examined to determine the current soil physiochemical properties as well as initial level of salinity (Table 1). Following soil analysis, salinity levels were developed with Sodium chloride (NaCl) because this salt is usually present in most of the salt-affected soils. In the control treatment, EC of the soil was maintained at the original level (1.7 dS m⁻¹) while salt was calculated to develop 8 dS m⁻¹ and 16 dS m⁻¹ EC levels following the range of EC occurring in semiarid agricultural areas of Punjab Pakistan (Hussain et al., 2021). To make the solution for treatment applications, the calculated amount of NaCl salt was liquefied (dissolved) in the water, subsequently, the prepared treatment water was sprayed on the soil which was already spread on the plastic sheet.

After -3 days, the soil was mixed again and analyzed to verify attainment of desired EC levels. Finally, 7 kg of soil was filled in each pot, and the six-month-old plants were transplanted into the pots. The pot experiment consisted of three treatments with five replications and was arranged using the Complete Randomized Design (CRD). The experiment was left for one-year duration under ambient conditions and irrigated regularly.

Data collection and measurements: After completing the experiment duration, growth parameters, biomass production, antioxidant activities and plant ionic and final soil analysis were performed.

Growth data and Biomass production: Plant height and root length were measured using the measuring tape while stem diameter was noted with the help of digital Vernier Caliper. For biomass measurement, all the individuals were harvested and tagged properly, subsequently, the fresh weight was recorded immediately after harvesting. For measuring the dry weight, the plants were oven-dried at (70°C) for 48 hours to assure attainment of constant weight. Finally, the plant root and shoot dry weight was noted using digital weight balance.

Determination of antioxidant enzyme activities: For measuring the antioxidant enzyme activity, one mature leaf per plant was harvested and immediately transferred to a freezer to prevent enzyme denaturing. The leaves having weight (0.5 g) from each sample were first ground in liquid nitrogen then homogenized in 5 ml of 0.2 mol 1⁻¹ phosphate buffer (pH 7.8). This homogenate was centrifuged at 12,000 rpm under chilling condition (4°C) for 15 minutes. Finally, the supernatant was collected in r Eppendorf and photochemical procedure was used to determine the antioxidant enzyme activities. The activity of Superoxide dismutase (SOD) was determined by measuring the ability to decrease the photochemical reduction of nitroblue tetrazolium (NBT) as described by Giannopolitis & Ries (1977), while the method demonstrated by Thomas et al., (1982) was used to measure the activity of Peroxidase (POD) and Catalase (CAT).

Ionic analysis: For the ionic analysis, dried leaf powder (0.1 g) was dissolved in 2 mL of 80% perchloric acid and 10 mL of concentrated H₂SO₄ and the mixture was diluted with sterile H₂O to 100 mL. The concentrations of sodium (Na), chlorine (Cl) and potassium (K) in the plant stem, root and leaves were estimated using Flame Photometer using the methods described by Wolf (1982).

Statistical analysis

The data of morphological and physiological traits were analyzed using one-way ANOVA using Statistica (Version 8.1). The significant differences between treatment means were compared using post hoc Tukey's HSD test. All tests and correlations were considered significant at (p<0.05) and all means were expressed with their standard errors (SE).

Table 1. Chemical properties of the soil used for experiment.

on per intente			
Soil properties	Values		
SAR	7.85		
ECe (dS m ⁻¹)	1.6		
pН	7.42		
Organic matter (%)	1.29		
Infiltration rate (mm/hour)	25.7		
Saturation (%)	30		
Texture	loam		

Table 2. Effect of tree species on soil EC (treatment means ±SE).

Treatments	Initial value	E. camaldulensis	S. cumini
Control	1.8 ± 0.2	$^{a}1.28 \pm 0.5$	$^{a}1.70 \pm 0.4$
8 dS m ⁻¹	8 ± 1.1	$^{\rm b}6.25 \pm 2.4$	$^{\rm b}7.25 \pm 2.1$
16 dS m ⁻¹	16 ± 2.4	$^{\circ}14.2 \pm 3.3$	$^{c}15.65 \pm 3.4$

Table 3. Effect of tree species on soil Sodium Adsorption ratio (mmol L^{-1})^{1/2}) (treatment means $\pm SE$).

Treatments	Initial value	E. camaldulensis	S. cumini
Control	7.2 ± 1.4	$^{a}6.45 \pm 1.1$	$^{a}6.84 \pm 1.1$
8 dS m ⁻¹	18.5 ± 2.7	$^{\rm b}16.6 \pm 3.2$	$^{b}17.45 \pm 4.6$
16 dS m ⁻¹	34.12 ± 1.5	$^{\circ}30.12 \pm 5.1$	$^{\rm c}33.1 \pm 6.1$

Results

Results from the growth traits revealed that both tree species showed salt sensitivity, but to a different extent, by decreasing root shoot lengths and stem diameter. However, E. camaldulensis exhibited lower sensitivity to salt stress by maintaining the highest root shoot lengths and root collar diameter under 16 dS-1 EC (Fig. 1). The maximum reduction in shoot height and stem diameter of E. camaldulensis was observed as 22.79% and 22.69% at the highest level of salinity. On the other hand, S. cumini exhibited more sensitivity to salt stress by showing lower plant height and stem diameter along with severe leaf necrosis or complete mortality of some individuals. The highest reduction in shoot height (30.98%) and stem diameter (41.53%) was noted in S. cumini at 16 dS⁻¹ EC with respect to control (Fig. 1). Similarly, shoot fresh and dry weight decreased significantly in both species particularly under the highest salt stress treatment. However, E. camaldulensis maintained more dry biomass and appeared to be the most tolerant tree species under moderate to high salinity (Fig. 1).

The concentration of Na⁺ and Cl⁻ increased significantly in leaf, stem, and root of both *S. cumini* and *E. camaldulensis* as a function of increasing salinity. The observed response trends of Na⁺ and Cl⁻ concentrations were generally similar for both species. However, *E. camaldulensis* performed better growth than *S. cumini* perhaps by absorbing a low concentration of Na⁺ and Cl⁻ in the various plant organs (Fig. 2). On the other hand, the highest concentration of Na⁺ and Cl⁻ ion was noted in stem and root sections of *S. cumini*, particularly under 16 dS m⁻¹ (Fig. 3). Similarly, the concentration of K⁺ ions in *E. camaldulensis* was highest in leaf as compared to stem and root at 16 dS m⁻¹ EC while the low leaf concentration of K⁺ ions was observed in *S. cumini* (Fig. 3).

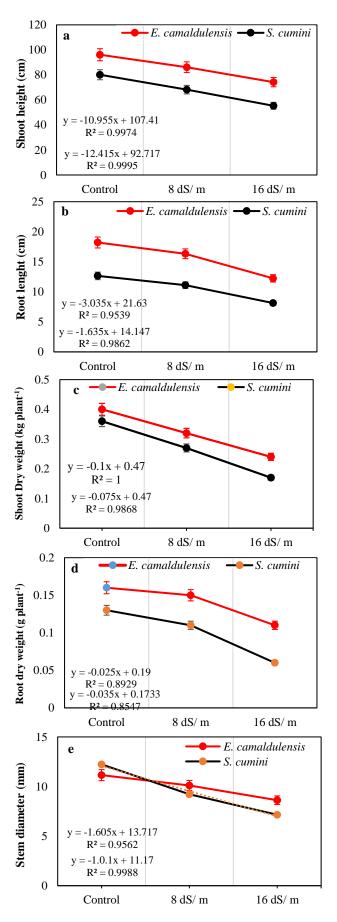


Fig. 1. Effect of salinity on growth traits and biomass production of *E. camaldulensis* and *S. cumini* (Fig; a= Shoot height, b= Root length, c= Shoot dry weight, d= Root dry weight and e= Stem diameter).

4 ARSLAN ARIF ETAL.,

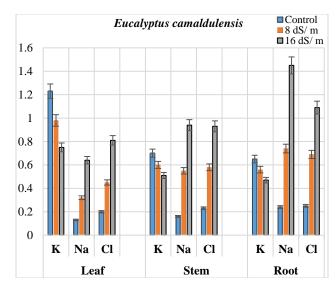


Fig. 2. Effect of salinity on ionic concentrations of leaf, stem and root in *E. camaldulensis*.

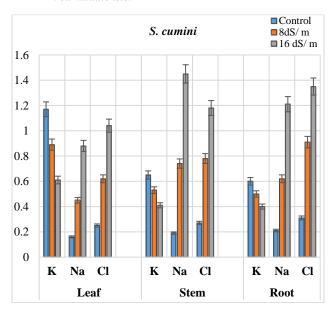


Fig. 3. Effect of salinity on ionic concentrations of leaf, stem and root in *S. cumini*.

Antioxidant enzyme activities increased significantly in both tree species with higher production of POD in *E. camaldulensis*, particularly under the high saline concentration (Fig. 4). In contrast, a low level of POD activity was observed in *S. cumini* at 16 dS m⁻¹ EC. Likewise, the activity of SOD also increased simultaneously with an increase in salt stress, where *E. camaldulensis* and *S. cumini* produced 11.35 and 8.56 U mg⁻¹ protein SOD activity, respectively. In the same way, a significant increase in CAT activity was noted in both tree species with greater increase in *E. camaldulensis*. and proved to be the first line of defense mechanism against salinity.

E. camaldulensis and *S. cumini* accumulated soil salts and significantly reduced levels of soil EC and SAR from their initial values showing phytoremedial potentials of these plants with *E. camaldulensis* showed the highest reduction in both soil parameters (Tables 2 and 3). It might be due to the strategy of *E. camaldulensis* allocating the extra salts to different plant organs.

Discussion

Plant growth under salt stress: Early plant growth features are known as the important indicators to assess the salt tolerance potential of tree species for planting in saline lands. In the present study, the effects of salinity were found more detrimental to the growth of S. cumini that started wilting from the aerial parts apparently due to sudden osmotic stress. This negative effect of salinity on growth and development has also been reported in many other tree species such as olive (Aragues et al., 2004) and Australian tree species (Van der Moezel et al., 1988). The results of the current study are supported by several previous studies indicating that the reduction in plant growth under salinity stress throughout the life cycle was closely related with decrease water potential and net CO₂ assimilation rate (Okhovatian-Ardakani et al., 2010), specific ion toxicity (Mansour, 2023), nutritional disorder and osmotic stress (Karimi & Tari, 2017). Salt sensitivity of plant species can also be attributed to the production of reactive oxygen species (ROS) under extreme saline conditions that lead to cellular damage and oxidative stress (Vaidyanathan et al., 2003). However, salt-tolerant plants also produce hydrogen peroxide (H₂O₂) that plays a significant role in initiating a series of physiological mechanisms that increase plant salt stress tolerance (Hao et al., 2021). This trend was noted in E. camaldulensis as this tree species showed a significant salt tolerance mechanism by maintaining optimum growth under extreme salinity. Our results are in line with a recent study ranking E. camaldulensis as one of the most salt-tolerant species among five multipurpose tree species (Hussain et al., 2021).

Effect of salinity on ionic balance: Plant ions particularly potassium (K+) and sodium (Na+) play a key role in regulating the plant enzyme activities (Blumwald, 2000). Owing to similar physio-chemical properties, both ions compete for the active site of enzymes. In this study, the accumulation of Cl⁻ and Na⁺ ions were high in roots and stem whereas K+ was accumulated more in leaves of E. camaldulensis. The higher rates of root Cl- and Na+ accumulation in S. cumini and the necrosis of some plants suggest this species has less long-term tolerance to salinity than E. camaldulensis. It has been supported from other studies that salt-tolerant tree species exhibit more Na+ and Cl⁻ sequestration in roots to control Na⁺ entry into leaf tissues (Apse & Blumwald, 2007; Møller et al., 2009). Based on these findings, it can be concluded that a parallel mechanism is possibly operating in the roots of E. camaldulensis to restrict excessive sequestration of Na+ in the leaf tissues, which is important to protect the photosynthetic activity of plant species from ion toxicity. This association between Na⁺ and Cl⁻ exclusion (salt tolerance) was also reported by (Van der Moezel et al., 1989) in various tree species like Eucalyptus and Casuarina. Similarly, Aswathappa & Bachelard (1986) found that the two salt-tolerant Casuarina species stored low level of Na⁺ and Cl⁻ in shoots as compared to the salt sensitive species indicating a higher level of salt tolerance. This salt exclusion strategy is mainly important in perennial or woody plants in which long term contact to salts can increase the absorption of toxic ions in the leaves (Munns & Tester, 2008). Moreover, Aragues et al., (2005) observed that after three years salt tolerance of olive plant suddenly declined due to exposure to salinity and became quite sensitive by increasing the level of toxic ions in the

leaves such as Na+. Andrés et al., (2014) indicated that accumulation of K+ in the plant guard cells could adjust normal stomatal behavior which is helpful to regulate gas exchange activities such as photosynthetic and transpiration rates. Therefore, it is essential to moderate the K+ homeostasis under salt stress in order to optimize photosynthetic rate, water potential, and cell turgidity (Marriboina et al., 2017). In the present study, S. cumini showed higher Na⁺, Cl⁻ and less K⁺ in aerial plant parts such as shoot and leaves. This suggests that S. cumini showed higher affinity towards Sodium and Chloride ion uptake under moderate to highly saline conditions. Therefore, this tree species cannot be grown in the salt-affected soils where the objective is the reclamation of such soils. However, the performance of E. under high salinity due to lower accumulation of Na, particularly due compartmentation which controls the mobility of sodium ions throughout the plant.

Effect of salinity on antioxidant enzyme activities: Tree species that flourish in salty environments have a variety of physiological adaptations (Flowers & Colmer, 2008). To combat oxidative stress, these plants produce many types of enzymes (Zulfiqar & Ashraf, 2023). However, the first line of defense against salinity is thought to be the synthesis of antioxidant enzymes in reaction to salts (Numan et al., 2018). Different types of antioxidant enzymes are produced by salt-tolerant tree species, but the three most significant ones are catalase (CAT), peroxidase (POD), and superoxide dismutase (SOD). Our results indicate that both tree species displayed enhanced enzymatic activity to varying degrees when exposed to extremely salty circumstances (Fig. 4). But in E. camaldulensis, this response was more dramatic, with considerable increases in SOD, CAT, and POD, especially below 16 dS m⁻¹. Our findings concur with those of Hussain et al., (2021) who identified E. camaldulensis as a salttolerant tree species with elevated SOD and POD activity up to 30 dS m⁻¹ EC. SOD, one of these antioxidant enzymes, is crucial for the detoxification of reactive oxygen species (ROS), which it does by converting them into H₂O₂ (Apel & Hirt, 2004). Similar to this, CAT is essential to the detoxification of H₂O₂. In plants under stress, a rise in POD activity was observed in stems higher than roots and leaves, according to (Zhang et al., 2013). In line with earlier research, E. camaldulensis showed a considerable increase in the antioxidant enzyme activities, which may have been caused by the ability to manufacture more antioxidant enzymes in response to the 16 dS m⁻¹ salt treatment.

Practical implementation: Even though *E. camaldulensis* gathered the least amount of Na in its leaves, it greatly decreased the amount of Na in the pot soil. This might be because plants use a technique to distribute salts to various organs, including the stem and roots, as was previously mentioned. These results, which are in line with earlier research, show that salt-tolerant tree species have the greatest potential for Na soil extraction (Hussain *et al.*, 2021). Harvesting the aboveground biomass enables the efficient removal of Na from the soil since Na accumulates mostly in the above-ground mass of woody plants. Similar findings were made by Tanvir & Siddiqui (2010), who looked into how *Populus. deltoides'* phytoextraction technique ended up being the primary driver of the removal of soil contaminants.

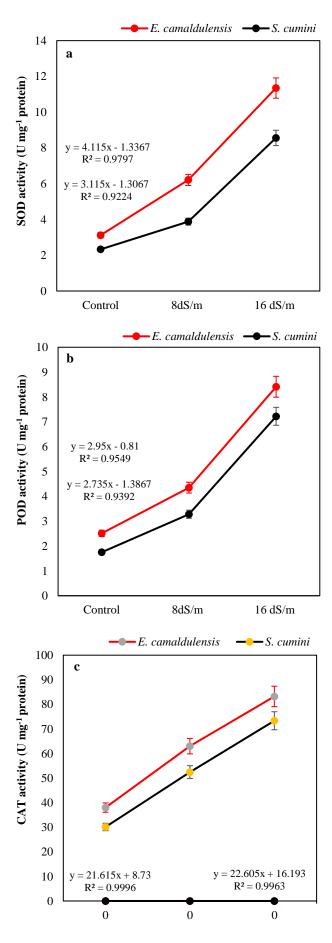


Fig. 4. Effect of salinity on antioxidant enzymes activities (Fig; a= SOD, b= POD and c=CAT).

ARSLAN ARIF ET AL.,

Conclusions

In the present study, different salt treatments negatively impacted the growth and biomass production of both tree species. However, E. camaldulensis showed the highest plant growth indicating the most salt-tolerant tree species under 8 dS⁻¹ while S. cumini showed significant decrease in growth with severe dieback and appeared to be the less salt-tolerant tree species. In both tree species, antioxidant enzymes (SOD, POD and CAT) activity increased by increasing salinity levels, but the significant increase was noted in E. camaldulensis indicating its defense mechanism against higher level of salinity. Furthermore, E. camaldulensis exhibited less accumulation of Na and Cl while more retention of K in different plant organs, suggesting more K ratio to Na and Cl was the important salt tolerance mechanism. On the other hand, S. cumini, accumulated more Na and Cl contents in leaves and stem with low K, indicating the salt sensitivity of this tree species. This comparative study identifies that plantation of E. camaldulensis is better than, S. cumini for restoration of salt-affected soils and biomass production. The expansion of agroforestry systems by planting such tolerant species is a sensible strategy would benefit the productivity of saline agriculture landscapes in order to cope with deficiency of staple foods, particularly in the countries facing severe food crises like Pakistan.

Acknowledgments

The authors would like to extend their sincere appreciation to Dr. John W. Groninger, Southern Illinois University, U.S.A. for his volunteer and enormous support to improve the language and overall quality of the manuscript.

References

- Andrés, Z., J. Pérez-Hormaeche, E.O. Leidi, K. Schlücking, L. Steinhorst, D.H. McLachlan, K. Schumacher, A.M. Hetherington, J. Kudla and B.J. Cubero. 2014. Control of vacuolar dynamics and regulation of stomatal aperture by tonoplast potassium uptake. *Proc. Natl. Acad. Sci. U. S. A.*, 111: E1806-E1814.
- Anonymous. 2017. The state of the world's land and water resources for food and agriculture. http://www.fao.org/3/a-i1688e.pdf
- Apel, K. and H. Hirt. 2004. Reactive oxygen species: metabolism, oxidative stress, and signal transduction. *PNAS.*, 55: 373-399.
- Apse, M.P. and E. Blumwald. 2007. Na+ transport in plants. *Annu. Rev. Plant Biol.*, 581: 2247-2254.
- Aragüés, R., J. Puy and D. Isidoro. 2004. Vegetative growth response of young olive trees (*Olea europaea L.*, cv. *Arbequina*) to soil salinity and waterlogging. *Plant Soil*, 258: 69-80.
- Aragüés, R., J. Puy, A. Royo and J.J.P. Espada. 2005. Three-year field response of young olive trees (*Olea europaea L.*, cv. *Arbequina*) to soil salinity: Trunk growth and leaf ion accumulation. *Plant Soil*, 271: 265-273.
- Aswathappa, N. and E.P. Bachelard. 1986. Ion regulation in the organs of *Casuarina* species differing in salt tolerance. *Aust. J. Plant Physiol.*, 13: 533-545.
- Banerjee, A., N. Dasgupta and B. De. 2005. *In vitro* study of antioxidant activity of *Syzygium cumini* fruit. *Food Chem.*, 90: 727-733.

Blumwald, E. 2000. Sodium transport and salt tolerance in plants. *Curr. Opin. Cell Biol.*, 12: 431-434.

- Brugnoli, E. and M. Lauteri. 1991. Effects of salinity on stomatal conductance, photosynthetic capacity, and carbon isotope discrimination of salt-tolerant (*Gossypium hirsutum* L.) and salt-sensitive (*Phaseolus vulgaris* L.) C3 non-halophytes. *J. Plant Physiol.*, 95: 628-635.
- Flexas J., J. Bota, F. Loreto, G. Cornic and T. Sharkey. 2004. Diffusive and metabolic limitations to photosynthesis under drought and salinity in C3 plants. *Plant Biol.*, 6: 269-279.
- Flowers, T. and T. Colmer. 2008. Salinity tolerance in halophytes. Abstract. *New Phytol.*, 179: 945-963.
- Giannopolitis, C.N. and S.K. Ries. 1977. Superoxide dismutases: I. Occurrence in higher plants. *Plant Physiol.*, 59: 309-314.
- Hao, S., Y. Wang, Y. Yan, Y. Liu, J. Wang and S. Chen. 2021. A review on plant responses to salt stress and their mechanisms of salt resistance. *Horticul.*, 7: 132.
- Hussain, M.S., M.S. Naeem, M.A. Tanvir, M.F. Nawaz and A. Abd-Elrahman. 2021. Eco-physiological evaluation of multipurpose tree species to ameliorate saline soils. *Int. J. Phytoremed.*, 23: 969-981.
- Jesus, J.M., A.S. Danko, A. Fiúza and M.T. Borges. 2015. Phytoremediation of salt-affected soils: A review of processes, applicability, and the impact of climate change. *Environ. Sci. Pollut. Res.*, 22: 6511-6525.
- Karimi, H. and F.E. Tari. 2017. Effects of NaHCO₃ on photosynthetic characteristics, and iron and sodium transfer in pomegranate. J. Plant Nutr., 40: 11-22.
- Kumar, A., S. Singh, A. Mukherjee, R.P. Rastogi and J.P. Verma. 2021. Salt-tolerant plant growth-promoting *Bacillus pumilus* strain JPVS11 to enhance plant growth attributes of rice and improve soil health under salinity stress. *Microbiol. Res.*, 242: 126616.
- Lautner, S. 2013. Wood formation under drought stress and salinity. In: Cellular aspects of wood formation. Springer., 187-202.
- Leprince, O., F.J. Harren, J. Buitink, M. Alberda and F.A. Hoekstra. 2000. Metabolic dysfunction and unabated respiration precede the loss of membrane integrity during dehydration of germinating radicles. *Plant Physiol.*, 122: 597-608.
- Ma, Y., M.C. Dias and H. Freitas. 2020. Drought and salinity stress responses and microbe-induced tolerance in plants. Front. Plant Sci., 11: 591911.
- Mansour, M.M. 2023. Role of vacuolar membrane transport systems in plant salinity tolerance. *J. Plant Growth Regul.*, 42: 1364-1401.
- Marriboina, S., D. Sengupta, S. Kumar and A.R. Reddy. 2017. Physiological and molecular insights into the high salinity tolerance of *Pongamia pinnata* (L.) pierre, a potential biofuel tree species. *Plant Sci.*, 258: 102-111.
- Møller, I.S., M. Gilliham, D. Jha, G.M. Mayo, S.J. Roy, J.C. Coates, J. Haseloff and M. Tester. 2009. Shoot Na+exclusion and increased salinity tolerance engineered by cell type–specific alteration of Na+ transport in Arabidopsis. *The Plant Cell.*, 21: 2163-2178.
- Munns, R. 2002. Comparative physiology of salt and water stress. *Plant Cell Environ.*, 25: 239-250.
- Munns, R. and M. Tester. 2008. Mechanisms of salinity tolerance. *Ann. Rev. Plant Biol.*, 59: 651-681.
- Nawar, S. H. Buddenbaum and J. Hill. 2015. Digital mapping of soil properties using multivariate statistical analysis and ASTER data in an arid region. *Remote Sens.*, 7: 1181-1205.
- Nouri, H., S.C. Borujeni, R. Nirola, A. Hassanli, S. Beecham, S. Alaghmand, C. Saint and D. Mulcahy. 2017. Application of green remediation on soil salinity treatment: A review on halophytoremediation. *Proc. Saf. Environ. Prot.*, 107: 94-107.

- Numan, M., S. Bashir, Y. Khan, R. Mumtaz, Z.K. Shinwari, A.L. Khan, A. Khan and A.H. Ahmed. 2018. Plant growth promoting bacteria as an alternative strategy for salt tolerance in plants: a review. *Microbiol. Res.*, 209: 21-32.
- Okhovatian-Ardakani, A., M. Mehrabanian, F. Dehghani and A. Akbarzadeh. 2010. Salt tolerance evaluation and relative comparison in cuttings of different omegranate cultivar. *Plant, Soil Environ.*, 56: 176-185.
- Pandey, P.C. and M. Pandey. 2023. Highlighting the role of agriculture and geospatial technology in food security and sustainable development goals. Sustain. Dev., 31: 3175-3195.
- Patil, S., M. Hebbara and S. Devarnavadagi. 1996. Screening of multipurpose trees for saline Vertisols and their bioameliorative effects. *Ann. Arid Zone*, 35: 57-60.
- Prasad, M. 2011. A state-of-the-art report on bioremediation, its applications to contaminated sites in India. Ministry of Environment & Forests, Government of India.
- Roy, S.J., S. Negrão and M. Tester. 2014. Salt resistant crop plants. *Curr. Opin. Biotechnol.*, 26: 115-124.
- Shafi, P., M. Rosamma, K. Jamil and P. Reddy. 2002. Antibacterial activity of *Syzygium cumini* and *Syzygium travancoricum* leaf essential oils. *Fitoterapia*, 73: 414-416.
- Shafique, S., R. Bajwa, A. Javaid and S. Shafique. 2005. Biological control of parthen/um iv: Suppressive ability of aqueous leaf extracts of some allelopathic trees against germination and early seed ling grovvth. *Pak. J. Weed Sci. Res.*, 11: 75-79.
- Stape, J.L., D. Binkley and M.G. Ryan. 2004. Eucalyptus production and the supply, use and efficiency of use of water, light and nitrogen across a geographic gradient in Brazil. For. Ecol. Manag., 193: 17-31.
- Tanvir, M.A. and M.T. Siddiqui. 2010.Growth performance and cadmium (Cd) uptake by *Populus deltoides* as irrigated by urban wastewater. *Pak. J. Agric. Sci.*, 47: 235-240.

- Thomas, R. L., J.J. Jen and C. V. Morr. 1982. Changes in soluble and bound peroxidase—IAA oxidase during tomato fruit development. *J. Food Sci.*, 47: 158-161.
- Vaidyanathan, H., P. Sivakumar, R. Chakrabarty and G. Thomas. 2003. Scavenging of reactive oxygen species in NaClstressed rice (*Oryza sativa* L.)—differential response in salttolerant and sensitive varieties. *Plant Sci.*, 165: 1411-1418.
- Van der Moezel, P., C. Walton, G. Pearce-Pinto and D.T. Bell. 1989. Screening for salinity and waterlogging tolerance in five *Casuarina* species. *Landsc. Urban Plan.*, 17: 331-337.
- Van der Moezel, P., L. Watson, G. Pearce-Pinto and D.T. Bell. 1988. The response of six Eucalyptus species and *Casuarina obesa* to the combined effect of salinity and waterlogging. *Aust. J. Plant Physiol.*, 15: 465-474.
- Vivekananda, J. J. Schilling, S. Mitra and N. Pandey. 2014. On shrimp, salt and security: livelihood risks and responses in South Bangladesh and East India. *Environ. Dev. Sustain*, 16:1 141-1161.
- Wolf, B. 1982. A comprehensive system of leaf analyses and its use for diagnosing crop nutrient status. *Comm. Soil Sci. Plant Anal.*, 13: 1035-1059.
- Zaghloul, M. 2020. Phytoremediation of heavy metals principles, mechanisms, enhancements with several efficiency enhancer methods and perspectives: A Review. *Middle East J.*, 9: 186-214.
- Zhang, M., Y. Fang, Y. Ji, Z. Jiang and L. Wang. 2013. Effects of salt stress on ion content, antioxidant enzymes and protein profile in different tissues of *Broussonetia papyrifera*. S. Afr. J. Bot., 85: 1-9.
- Zulfiqar, Faisal and M. Ashraf. 2023. Proline alleviates abiotic stress induced oxidative stress in plants. *J. Plant Growth Regul.*, 42: 4629-4651.

(Received for publication 30 May 2023)