ASSESSMENT OF SEEDLING EMERGENCE, GROWTH TRAITS AND PHYSIOLOGICAL INDICES OF A MEDICINAL PLANT *CALOTROPIS GIGANTEA* (L.) AITON f. UNDER NaCl INDUCED SALINITY STRESS

MD. RABIUL ISLAM^{1*}, TANJUM ARA SINTHY¹, MD. ABU HASAN¹, MD. NAJMOL HOQUE², SUBROTA KUMER PRAMANIK¹, MD. SHAZADUR RAHMAN³ AND MD. ABU SAYED⁴

¹Department of Crop Physiology and Ecology, Faculty of Agriculture, Hajee Mohammad Danesh Science and Technology University, Dinajpur-5200, Bangladesh

²Department of Biochemistry and Molecular Biology, Faculty of Agriculture, Khulna Agricultural University, Khulna 9100, Bangladesh

³Department of Agricultural Chemistry, Faculty of Agriculture, Hajee Mohammad Danesh Science and Technology University, Dinajpur-5200, Bangladesh

⁴Department of Biochemistry and Molecular Biology, Faculty of Agriculture, Hajee Mohammad Danesh Science and Technology University, Dinajpur-5200, Bangladesh

*Corresponding author's email: rabiulislam@hstu.ac.bd

Abstract

Calotropis gigantea (L.) Aiton f. giant milkweed or *Aakanda* (in Bengali) is a branched large shrub, spreading in many countries all over the world and has important medicinal and fiber production values. With this research, we evaluate the effects of NaCl salinity stress on the seedling emergence, growth and physiological traits of *Calotropis gigantea*. Following a randomized complete block design with artificial four levels of saline soils *viz*. 6, 8, 10 and 12 dS m⁻¹ alongside with control (0 dS m⁻¹) were studied with three replications. *Calotropis gigantea* seeds were tested in varying concentrations of artificial saline soil for 45 days. Analysis of variance showed that seedling emergence, growth and physiological attributes revealed to be severely impacted due to salt stress. Mean emergence time and proline content rose along with a rise in salinity level, but other characteristics declined with the upsurge of saline stress. Reduction of total chlorophyll content was more in 12 dS m⁻¹ salinity treatment compared to control condition. Principal component analysis showed that first principal component contributed 87% of the total variation and noticeably distinguished *Calotropis gigantea* genotype for seedling emergence, growth and physiological traits under all salinity treatments (6, 8, 10 and 12 dS m⁻¹) from control condition (0 dS m⁻¹). Correlation study exhibited that mean emergence time and proline content had positive correlation but both were negatively correlated with other studied traits. Overall, our present investigation brought a new output for cultivating the wild shrub *Calotropis gigantea* in saline prone soil, especially in the coastal regions of the world.

Key words: Calotropis gigantea, Chlorophyll, Proline content, NaCl salinity, Seedling emergence, Trait association.

Introduction

Calotropis gigantean (L.) Aiton f., belonging to Apocynaceae family prefers to grow in very low (closer to 0 m) to high altitudinal ranges (up to 1300 m) and has orthodox seed storage capability (Francis, 2003; Orwa et al., 2009). Calotropis is considered as a multipurpose plant and both the species C. gigantea and C. procera have similar medicinal and pharmacological properties (Ahmed et al., 2005). In Bangladesh, China, and India C. gigantea is often practiced in conventional medicine (Parhira et al., 2016; Motaleb, 2011). Bangladesh is one of the native countries of this fast-growing large shrub or small tree (Li, 2015) where it is locally called as Aakanda. Calotropis gigantea grows on fallow lands all over Bangladesh (Islam et al., 2019). The giant milkweed plant that reaches up to 4 m tall with clusters of white or lavender waxy flowers is a salt tolerant, drought resistant plant and also can grow in disturbed soil (Sharma and Tripathi, 2009). However, though Calotropis gigantea is considered a high valuable plant, but still, this is undomesticated and there is an urgent need to develop new varieties. Presently, scientists are giving their intense attention to cultivate Calotropis plant for medicine, fiber and biodiesel production (Parhira et al., 2016; Kanchan and Atreya, 2016; Muriira, 2017). However, there are limited number of investigations available under adverse environmental conditions such as salinity and drought with regards to commercial production of *Calotropis gigantea*. Therefore, considering the key significances of this plant, it is also crucial to analyze the seed germination, seedling emergence and growth traits under different environmental conditions of the globe.

Climate change has caused an increase in soil salinity, from approximately 0.83 million ha in 1973 to 1.02 million hectares in 2000, and 1.05 million ha in 2009 in Bangladesh (Anonymous, 2010). Recently, it is reported that this salinity has spread over more than 150 km upstream in the rivers of the southern coast that seriously affected agricultural crops production, environment and public health (Uddin, 2021). In Bangladesh's saline regions, cropping intensity is generally lower (62–144%) than that of the country as a whole (159%), which corresponds to typical growing circumstances (Karim *et al.*, 1990). As an emerging medicinal and fiber plant, the domestication, establishment and cultivation of *Calotropis gigantea* is very essential for effective use of the coastal saline areas of Bangladesh.

Numerous tiny seeds are produced by wild *Calotropis* species, which also reproduces asexually through root and stem suckers and by wind-dispersed seeds. The spread of seeds, germination, emergence, and the development of young seedlings are the main factors in the establishment of

a species in a new location. Germination, emergence, and early seedling growth, all of which are particularly susceptible to the salt of the substrate make up the three stages of the plant's establishment (Jamil et al., 2005). Extremely low percentage of germination and delayed germination are both effects of soil salt (Baskin & Baskin, 1997). Early seedling establishment is enhanced by rapid and homogeneous seed germination under salty circumstances. which ultimately enhances the output (Bradford, 1995). The amount of salt tolerance during germination and the level of tolerance during seedling growth are related to one another (Jhala, 1997). Therefore, the present investigation was undertaken with an aim to assess the seedling emergence, growth and physiological attributes of Calotropis gigantea under salinity stress condition.

Material and Methods

Experimental design and layout: The experiment was carried out in a randomized complete block design with three replications at the laboratory and research field of Department of Crop Physiology and Ecology, Hajee Mohammad Danesh Science and Technology University (HSTU), Dinajpur, Bangladesh during February–May, 2021. There were five NaCl treatments, used in the experiment *viz.* control (0), 6, 8, 10 and 12 dS m⁻¹. The seedlings were grown in 12 L plastic pots that had the following dimensions: 25 cm height, 27 cm opening, and 21 cm diameter base maintaining distances between pot to pot and replication to replication were 0.5 and 1.0 m, respectively.

Soil analysis: At first, the soil was collected from the land of the HSTU research farm. Weeds and stubble were removed, cleaned and crushed properly. For physiochemical analysis of used soil, three random samples were tested in Soil Resource Development Institute (SRDI), Dinajpur and the following mean results were found: texture sandy loam, pH 5.50, salinity 0.370 dS m⁻¹, organic carbon 0.240%, organic matter 0.413%, total N 0.021%, available P 29.22 μ g/g and exchangeable K 0.117 meq/100g.

Development of artificial saline soil using standard curve: Artificial saline soil of different treatments (0, 6, 8, 10 and 12 dS m⁻¹) was carried out utilizing a specified quantity of NaCl in soil using standard curve (Fig. 1). The salts were fully combined with the air-dried soil after being dissolved in tap water. For every 10 kg soil, we applied NaCl as 0.0, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9 and 1.0% solution in each dish (Capacity 15 L) which replicated three times followed completely randomized design (Table 1). After proper mixing, the dishes of soil were covered with polythene sheet for 7 days and everyday soil was mixed carefully. After passing seven days the dishes were uncovered and continued proper mixing of soil every day till it was completely dried. Rainfall was also controlled with proper management using polythene sheet. When the soils were dried (almost 15 days required), 100 g of soil from each bowl kept in a separate place. Then the soils were cleaned and sieved properly. The soil samples were packed in different polybags, tagged and sent to SRDI for determining the level of salinity. From the results (Table

1) a regression linear (r) graph was established using developed salinity (dS m⁻¹) *versus* added salt (g kg⁻¹ soil) (Fig. 1). Further, the linear regression (r) was tested and it was found significant which was considered as a standard curve (y = 1.4598x + 0.387, r = 0.985, p < 0.001). Using this significant standard curve, artificial soil salinity of 6, 8, 10 and 12 dS m⁻¹ was developed at four different trays following mentioned procedures (Hussain *et al.*, 1989) and finally, again affirmed the salinity level by testing three soil samples from each tray.



Fig. 1. Linear standard curve for developing various level of artificial salinity in studied soil for pot experiment.

Seed collection and sowing: The fruit color of C. gigantea is changed from dark green to green or yellowish green on maturation (El-Tantawy, 2000). Physiologically fruit riped with open capsules and brown seeds, was collected from fallow land of Nayanpur, Dinajpur Sadar, Bangladesh (25°39'12'' N latitude and 88°39'14'' E longitude) during August 2020. Ten pods were collected from three plants maintaining ≤ 2 m plant to plant distance. Previously, the distance between two sampling individuals of C. gigantea was studied as 20-50 m within a population and revealed the homogenous genetic diversity of the individuals within this distance (Islam et al., 2019). Calotropis gigantea is a cross pollinated plant, bearing wind dispersal seeds with prominent vegetative propagation traits, hence the plants within $\leq 2m$ might be having homogenous genetic characters and mixing of all seeds from three plants was reasonable for conducting present investigation. Seeds were manually removed from the pod, mixed properly and put in the shade to dry. The dried seeds were preserved in a refrigerator at 5°C temperature until sowing at 1st February 2021. Pots were filled with 10 kg prepared saline soil (6, 8, 10 and 12 dS m⁻¹) from the respective trays along with the control. The soil was mixed properly before seed sowing and twenty seeds of C. gigantea were sown in each pot. In accordance with the suggestions from the chemical and physical analyses, soil was fertilized, 5% organic matter and 2.5 g of triple super phosphate applied at foundation, 10 g of potassium chloride and 10 g of urea applied at 30 days after sowing (DAS) of seed in each pot. Plants were irrigated by tap water in every 3 days interval, based on moisture availability of the pot soil. A Thiamethoxam-based product (8 g/ 20 L) was applied to control aphid infestation at 45 DAS.

NaCl solution (%) applied in soil	Amount of soil (kg)	Applied salt (g)	Developed salinity (dS m ⁻¹)
0.0	10	0	0.36
0.1	10	10	1.69
0.2	10	20	4.05
0.3	10	30	5.14
0.4	10	40	6.57
0.5	10	50	7.43
0.6	10	60	8.47
0.7	10	70	9.33
0.8	10	80	10.92
0.9	10	90	14.57
1.0	10	100	16.00

 Table 1. Development of artificial soil salinity in 10 kg soil at dishes for pot experiment.

Data collection: Seedling emergence and its associate traits were estimated till 30th days after seed placement into pots. After emergency evaluation, C. gigantea plants were thinned from each pot keeping 5 healthy seedlings and allowed to grow upto 45 days for further assessment of seedling growth and the physiological attributes. Seedling emergence percentage (SEP), one of the characteristics of seedling emergence, was estimated by subtracting the number of emerged seedlings from the total number of seeds put in each pot, then multiplying the result by 100 (Mitchell and Vogel, 2012). The emergence vigor index was calculated as: Emergence Vigor Index = $\Sigma(Et/Dt)$, Where Et is the number of seedlings emerged, Dt is the number of days after planting when the seedlings were counted (Yang et al., 2005; Copeland, 1976). Mean germination time (MGT) by Al-Ansari and Ksiksi (2016) was used to estimate the mean emergence time (MET): $\Sigma Fx/\Sigma F$, where F is number of seedlings emerged on day x.

Among seedling growth traits: total leaf area (TLA), number of leaves (NL), stem diameter (SD), shoot length (SL), root length (RL) and total dry mass (TDM) were collected at 45 DAS. Individual leaf area was calculated using the equation Leaf Area (cm^2) = W × L × 0.75, proposed by Moreira *et al.* (2007) for *C. procera*, a sister species of *C. gigantea*. Total Leaf Area was the sum of all leaf area measurements for each leaf (Almeida *et al.*, 2019). A digital caliper was used to measure Stem Diameter (cm) at the base of the stem, corresponding to the lap of the plants. After 45 DAS three seedlings were sampled from each pot for taking shoot length and root length manually and finally, dried separately at 70°C for 72 h in an electric oven and weighed on an electrical balance for recorded total dry mass.

Physiological trait SPAD (Soil Plant Analyses Development) value was taken from middle portion of the youngest fully expanded leaf of the three tagged plants at 45 DAS using self-calibrating Minolta chlorophyll meter and the average of the three values were recorded. Total chlorophyll content of the apex leaf at 45 DAS of *C. gigantea* was estimated according to Witham *et al.* (1986) as Total Chlorophyll (mg g⁻¹ FW) = [20.2(D₆₄₅) + $8.02(D_{663})] \times [V/(1000 \times W)]$; Where, V = Final volume of filtrated extract, W = Weight of fresh leaf, D₆₄₅ =

Table 2. Analysis of variance table for the studied seedling emergence, growth and physiological traits of *Calotronis gigantea* genotype.

Culou opis gigunicu genotype.								
Traits	MS							
Seedling emergence percentage	3040.00**							
Emergence vigor index	230.03**							
Mean emergence time	4.73**							
Total leaf area	39.88**							
Number of leaves	18.51**							
Stem diameter	0.192**							
Shoot length	53.22**							
Root length	51.17**							
Total dry mass	1.10**							
SPAD value	208.46**							
Total chlorophyll content	0.175**							
Proline content	3.34**							
MC Manager **Indianter indf	- + + 10/ 11 - f1 - h :1:4							

MS = Mean squares, **Indicates significant at 1% level of probability

Absorbance at 645 nm wavelength, D_{663} = Absorbance at 663 nm wavelength. The proline content was estimated at 45 DAS by following the protocol of Bates *et al.*, (1973) as fresh weight basis using the formula below:

 μ moles proline / g of fresh plant material = {(μ g proline / ml × ml toluene) / 115.5 μ g / μ moles} / (g sample / 5).

Statistical analysis of data: Collected data were statistically analysed to find out the level of significance using Statistix 10 program (https://www.statistix.com/free-trial/). The mean differences were compared by Tukey's HSD Test at $p \le 5\%$ level. Relative changes in stress condition over control for all studied parameters were measured using the formula as Relative Change = [(Treatment - Control) / Control] ×100. Minitab 17 statistical software package (Minitab Inc., State College, PA, USA) was used for principal component (PC) analysis and Pearson correlation analysis. ANOVA of the PC scores was performed for treatments following one way ANOVA model.

Results

Effects of different salinity treatment on seedling emergence traits of Calotropis gigantea: Analysis of variance revealed that several artificially developed saline soils $(0, 6, 8, 10 \text{ and } 12 \text{ dS m}^{-1})$ had a major impact on seedling emergence percentage, emergence vigor index and mean emergence time of Calotropis gigantea genotype at 30 days after sowing (Table 2). Due to salt stress condition emergence percentage and emergence vigor index of C. gigantea seedling were reduced vigorously in all salinity levels, whereas mean germination time of C. gigantea seedling was increased at salinity stress condition compared to control situation (Table 3). Highest emergence percentage was found at control (98.33) followed by 6 dS m^{-1} (65%) and 8 dS m^{-1} (60%). This reduction was not the same for all developed salinity for emergence percentage. The reduction was more pronounced in 12 dS m^{-1} (81.36%) and 10 dS m⁻¹ (71.19%) than that of 8 dS m⁻¹ (38.98%) and 6 dS m⁻¹ (33.89%). For emergence vigor index, highest vigor index was detected at control (24.65) followed by 6 dS m^{-1} (11.88) and 8 dS m^{-1} (10.60). The reduction in

relative changes over control were lower in 6 dS m⁻¹ (51.81%) and 8 dS m⁻¹ (56.99%) than 10 dS m⁻¹ (81.99%) and 12 dS m⁻¹ (90.91%). In case of 12 dS m⁻¹ mean emergence, time was highest (23.09 days), which was followed by 10 dS m⁻¹ (22.10 days). In control condition mean emergence time was decreased by 19.66 days. The increment for mean germination time was greater in 12 dS m⁻¹ (17.45%) and 10 dS m⁻¹ (12.41%) than 8 dS m⁻¹ (9.16%) and 6 dS m⁻¹ (8.44%).

Effects of different salinity treatment on seedling growth traits of Calotropis gigantea: Analysis of variance revealed that all the studied seedling growth traits like total leaf area, number of leaves, shoot length, stem diameter, root length and total dry mass were were extremely affected by the impact of various salinity levels (0, 6, 8, 10 and 12 dS m⁻¹) in soil culture (Table 2). It was observed that the studied growth traits in C. gigantea seedling declined with the increment of salt concentration in soil (Table 4), but the degree of reduction was not equal in different salinity stress. Greater leaf area degradation had been seen when exposed to salt stress in 12 dS m^{-1} (87.05%) and 10 dS m^{-1} (84.64%) than 8 dS m^{-1} (58.55%) and 6 dS m⁻¹ (50.05%). The reduction in number of leaves over control were lower in 6 dS m⁻¹ (37.05%), medium in 8 dS m^{-1} (51.10%) and 10 dS m^{-1} (53.11%) and higher in 12 dS m^{-1} (64.36%). The reduction of stem diameter was more pronounced in 12 dS m⁻¹ (53.50%) and 10 dS m⁻¹ (45.83%) than that of 8 dS m⁻¹ (43.75%) and 6 dS m⁻¹ (30.75%). The reduction for shot length was more pronounced in 12 dS m⁻¹ (65.04%) and 10 dS m⁻¹ (64.59%) than that of 8 dS m⁻¹ (55.74%) and 6 dS m⁻¹ (48.04%). Root length was decreased over control condition at 6, 8, 10 and 12 dS m⁻¹ salinity levels by 46.91, 60.32, 63.64 and 65.56%, respectively. The reduction for total dry mass was more prominent in 12 dS m⁻¹ (93.68%) and 10 dS m⁻¹ (91.12%) than 8 dS m⁻¹ (89.74%) and 6 dS m⁻¹ (77.43%).

т

Effects of diverse salinity treatment on physiological traits of Calotropis gigantea: Analysis of variance exhibited that SPAD value, proline and total chlorophyll content of Calotropis gigantea seedling at 45 DAS were significantly influenced under various salinity levels (0, 6, 8, 10 and 12 dS m⁻¹) (Table 2). Due to salt stress condition SPAD value, total chlorophyll content of C. gigantea seedling was reduced dynamically in all salinity levels but the rising salt levels resulted in an increase in proline content (Table 5). The reduction of SPAD value was more pronounced in 12 dS m⁻¹ (32.64%) and 10 dS m⁻¹ (18.36%) than that of 8 dS m⁻¹ (3.86%) and 6 dS m⁻¹ (1.51%). The reduction in total chlorophyll content over control were lower in 6 dS m⁻¹ (16.89%), medium in 8 dS m⁻¹ (48.25%) and 10 dS m^{-1} (49.51%) and higher in 12 dS m^{-1} (51.65%). Under salt stress at 12 dS m⁻¹ the highest amount of proline was found (2.70 μ mole g⁻¹ FW) and the control condition provided the lowest proline content (0.37 μ mole g⁻¹ FW). The degree of enhancement was more in 12 dS m⁻¹ (629.73%) than 10 dS m⁻¹ (624.32%), 8 dS m⁻¹ (502.70%) and 6 dS m⁻¹ (170.27%).

Phenotypic correlation among studied seedling emergence, growth and physiological traits: Correlation study revealed the mutual relationship among the seedling emergence, growth and physiological traits (Table 6). The traits under study showed both strong positive and negative correlations. Proline content indicated substantial negative association with all other features excluding mean emergence time, and mean emergence time also exhibited strong negative correlation with proline content. Mean emergence time displayed significant positive correlation with proline content (r = 0.826). Remaining attributes showed significant positive correlation with each other.

Salinity level	S	EP	E	VI	MET		
(dS m ⁻¹)	%	RC (%)	Values	RC (%)	Days	RC (%)	
0	98.33 a	—	24.65 a	—	19.66 c	—	
6	65.00 b	-33.89	11.88 b	-51.81	21.32 b	8.44	
8	60.00 b	-38.98	10.60 b	-56.99	21.46 b	9.16	
10	28.33 c	-71.19	4.44 c	-81.99	22.10 ab	12.41	
12	18.33 c	-81.36	2.24 c	-90.91	23.09 a	17.45	
CV (%)	12.93		17	7.61	2.00		

Table 3. Effect of different salinity levels on seedling emergence traits of *Calotropis gigantea* genotype.

Mean followed by same letter(s) in column did not significant at 5% level of probability under Tukey HSD, RC (%) = Relative changes over control (%), SEP= Seedling emergence percentage (%), EVI= Emergence vigor index, MET= Mean emergence time

Fable 4. Effect of different salini	y levels on growth traits of	Calotropis gigantea genotype.

	Table 4. Effect of uniferent samily levels on growth traits of <i>Caloropis giganiea</i> genotype.											
Salinity level	Salinity level TLA		NL		S	SD		SL		RL		DM
(dS m ⁻¹)	cm ²	RC (%)	No.	RC (%)	cm	RC (%)	cm	RC (%)	cm	RC (%)	g	RC (%)
0	10.35 a	-	9.96 a	-	1.20 a	-	15.59 a	-	15.07 a	_	1.52 a	-
6	5.17 b	-50.05	6.27 b	-37.05	0.831 b	-30.75	8.10 b	-48.04	8.00 b	-46.91	0.343 b	-77.43
8	4.29 b	-58.55	4.87 bc	-51.10	0.675 bc	-43.75	6.90 bc	-55.74	5.98 bc	-60.32	0.156 c	-89.74
10	1.59 c	-84.64	4.67 bc	-53.11	0.650 bc	-45.83	5.52 c	-64.59	5.48 c	-63.64	0.135 c	-91.12
12	1.34 c	-87.05	3.55 c	-64.36	0.558 c	-53.50	5.45 c	-65.04	5.19 c	-65.56	0.096 c	-93.68
CV (%)	CV (%) 15.61		11	11.50		9.75 7.8		7.80		.87	13.09	

Mean followed by same letter(s) in column did not significant at 5% level of probability under Tukey HSD, RC (%) = Relative changes over control (%), TLA = Total leaf area, NL = Number of leaves, SD = Stem diameter, SL = Shoot length, RL = Root length, TDM = Total dry mass

Salinity level	SPAD	value	Total chloro	ohyll content	Proline content		
$(dS m^{-1})$	SPAD unit	RC (%)	mg g ⁻¹ FW	RC (%)	µmole g ⁻¹ FW	RC (%)	
0	59.53 a	_	1.030 a	-	0.37 c	170.27	
6	58.63 a	-1.51	0.856 a	-16.89	1.00 bc	502.70	
8	57.23 a	-3.86	0.533 b	-48.25	2.23 ab	624.32	
10	48.60 ab	-18.36	0.520 b	-49.51	2.68 a	629.73	
12	40.10 b	-32.64	0.498 b	-51.65	2.70 a	170.27	
CV (%)	CV (%) 8.23		9.4	41	31.98		

Table 5. Effect of different salinity levels on physiological traits of *Calotropis gigantea* genotype.

Mean followed by same letter(s) in column did not significant at 5% level of probability under Tukey HSD, RC (%) = Relative changes over control (%)

Table 6. Correlation coefficients among seedling emergence, growth and physiological traits of <i>Calotropis gigantea</i> genotype.												
Traits	SEP	EVI	MET	TLA	NL	SD	SL	RL	TDM	SPAD	TC	Proline

-	SEP EVI	1.00 0.972***	1.00										
	MET	- 0.920***	- 0.954***	1.00									
	TLA	0.935***	0.969***	- 0.937***	1.00								
	NL	0.888***	0.925***	- 0.893***	0.940***	1.00							
	SD	0.847***	0.896***	- 0.872***	0.928***	0.954***	1.00						
	SL	0.894***	0.961***	- 0.892***	0.951***	0.926***	0.924***	1.00					
	RL	0.850***	0.908***	- 0.839***	0.939***	0.940***	0.924***	0.969***	1.00				
				0.002					0.969***	1.00			
				0.020					0.546*		1.00		
	TC	0.834***	0.851***	- 0.813***	0.893***	0.886***	0.896***	0.874***	0.895***	0.858***	0.634*	1.00	
	D 1	-	-	0.00(***	-	-	-	-	0.001***	0 700***	0 (77**	0.000***	1 00

Proline 0.794*** 0.809*** 0.826*** 0.873*** 0.805*** 0.823*** 0.804*** -0.821*** -0.722*** -0.677** -0.899*** *, ** and *** = significant at $P \le 5$, $\le 1\%$ and $\le 0.1\%$ level of probability, SEP = Seedling emergence percentage (%), EVI = Emergence vigor index, MET = Mean emergence time (days), TLA = Total leaf area (cm²), NL = Number of leaves (No.), SD = Stem diameter (cm), SL = Shoot length (cm), RL = Root length (cm), TDM = Total dry mass (g), SPAD = SPAD value (SPAD unit), TC = Total chlorophyll content (mg g⁻¹ fresh weight), Proline = Proline content (μ mole g⁻¹ fresh weight).

Table 7. Coefficients of PCs from PCA and mean PC scores with standard deviation of *Calotropis gigantea* genotype.

with standard deviation of Caloiropis gigun	1
Variable	PC1
Seedling emergence percentage (%)	0.292
Emergence vigor index	3.02
Mean emergence time (days)	-0.294
Total leaf area (cm ²)	3.05
Number of leaves (No.)	0.297
Stem diameter (cm)	0.294
Shoot length (cm)	0.299
Root length (cm)	0.299
Total dry mass (g)	0.292
SPAD value (SPAD unit)	0.224
Total chlorophyll content (mg g ⁻¹ fresh weight)	0.286
Proline (µmole g ⁻¹ fresh weight)	-0.274
% Variation explained	87.00
P value	< 0.001
Mean PC scores with standard deviation	
Treatment	PC1
0 dS m^{-1}	$5.55\pm0.14~a$
6 dS m^{-1}	$0.86\pm0.56\ b$
8 dS m ⁻¹	$\textbf{-0.80} \pm 0.45~c$
10 dS m^{-1}	$\textbf{-2.30}\pm0.15~d$
12 dS m ⁻¹	$3.30\pm0.30\;e$

Principal component analysis (PCA): PCA is a dimensionality-reduction method that lessens the dimensionality of the large data sets. In this experiment, first principal component (PC1) explained 87.00% of the total data variation for the studied seedling emergence, growth and physiological traits of genotype Calotropis gigantea (Table 7). This PC was acknowledged on the basis of an eigen value higher than unity. Positive coefficients of total leaf area (3.05), emergence vigor index (3.02) and negative coefficient of mean emergence time (-0.294), proline content (-0.274) were mostly contributed to the variation explained by PC1. PC scores clearly separated all salinity treatments (6, 8, 10 and 12 dS m⁻¹) from control (0 dS m^{-1}) condition for *C. gigantea* genotype by their opposing scenery in PCA-biplot (Fig. 2) and by their differing mean PC scores (Table 7). The PCA-biplot clearly revealed that PC1 scores for control (0 dS m^{-1}) were completely separated from the PC1 scores of other salinity treatments (6, 8, 10 and 12 dS m⁻¹) used in this experiment (Fig. 2, Table 7).

1.00



Fig. 2. Biplot for studied seedling emergence, growth and physiological traits of *Calotropis gigantea* genotype under different salinity (NaCl) levels; SEP = Seedling emergence percentage (%), EVI = Emergence vigor index, MET = Mean emergence time (days), TLA = Total leaf area (cm²), NL = Number of leaves (No.), SD = Stem diameter (cm), SL = Shoot length (cm), RL = Root length (cm), TDM = Total dry mass (g), SPAD = SPAD value (SPAD unit), TC = Total chlorophyll content (mg g⁻¹ fresh weight), Proline = Proline content (µmole g⁻¹ fresh weight). 1, 2 and 3 = T0 = 0 dS m⁻¹ salinity treatment, 4, 5 and 6 = T1 = 6 dS m⁻¹ salinity treatment, 7, 8 and 9 = T2 = 8 dS m⁻¹ salinity treatment, 10, 11 and 12 = T3 = 10 dS m⁻¹ salinity treatment.

Discussion

Seedling emergence evaluation under salinity stress: The stages of emergence and germination are essential for plant establishment (Song *et al.*, 2008). The majority of plant species are extremely vulnerable to high salt during the germination and seedling stages of development (Gulzar *et al.*, 2003). Most plants are vulnerable to ion stress during germination and seedling emergence (Catalan *et al.*, 1994, Chauhan and Johnson, 2009). In this experiment, all salinity levels vigorously reduced the

emergence percentage and emergence vigor index of C. gigantea seedling, whereas an increase in mean germination time of C. gigantea seedling was observed at different salinity levels over control (Table 3). More distinct reduction of emergence percentage and emergence vigor index was noticed at 12 and 10 dS m⁻¹ salinity treatments compared to 6 and 8 dS m⁻¹ salinity treatments (Table 3). Similarly increase of mean emergence time was also more prominent in 12 dS m⁻¹ compared to 6 dS m⁻¹ (Table 3). This result of emergence percentage showed conformity with the findings of Guo et al., (2020) who reported that greater salt concentrations decreased the rate of emergence of seedlings, with a drop of 26.00 % (when cultivated in 200 mM NaCl) and a fall of 46.00 % (in 400 mM NaCl) for third generation seeds of Suaeda salsa. Again Dehnavi et al., (2020) also found that seedling vigor index of different genotypes of sorghum was decreased 55.5-88.4% at 200 mM NaCl saline stress. Gill et al., (2003) found that salt stress delayed the germination of seeds due to developing osmotically enforced dormancy. It might be a mechanism used by seeds to avoid germination under challenging conditions and ensure optimal seedling establishment. These outputs were in support with the present findings.

Seedling growth evaluation under salinity stress: Plants are predominantly sensitive in early stages of seedling development by elevated salinity (Jamil et al., 2005). Salinity stress also tempts the incidence of water shortage stress through reduction of osmotic potential of soil solutes thus crop roots face difficulties for water uptake from soil (Heidarpour et al., 2009). This causes harmful effects on the metabolism of plants (James et al., 1983). Salinity stress decreased root length, callus size, coleoptile length, and seedling growth (Agnihotri et al., 2006; Bera et al., 2006). Numerous plants are susceptible to ion stress caused by salinity condition during early seedling growth (Alom et al., 2016) Additionally, a high Na+ concentration prevents the absorption of K+ ions, a crucial component for growth and development, which lowers productivity and may even cause plant death (James et al., 1983). In present experiment, all the studied growth traits in C. gigantea seedling were decreased when the percentage of salt rises in soil (Table 4). The reduction of total leaf area, number of leaves, stem diameter, shoot length and root length were more noticeable in higher salinity level compared to control condition (Table 4). When the impact of saline water stress on Calotropis procera seedling growth and characteristics was examined in a greenhouse setting, it was discovered that the control treatment had the highest rates of root length, shoot and root dry weight, seedling height, collar diameter, and number of leaves (Bahmani and Kartoolinejad, 2017). Bilkis et al., (2016) established that shoot height, root length, shoot and root dry weight were greatly diminished as a result of an increase in salt concentrations in wheat. Matsuura et al., (2005) found that the leaf area was reduced by the salt treatment to 72% and 52% is under control in Tsushima and Pontivy buckwheat, respectively. These outputs were in parallel with the present findings. Reduction in the number of leaves per plant over control was found in polyembryonic mango

(*Mangifera indica* L.) genotypes at saline stress (Nimbolkar *et al.*, 2018). Guo *et al.*, (2020) revealed that stem diameter was reduced due to salinity stress in *S. salsa*. Dehnavi *et al.*, (2020) found that shoot length of different genotypes of sorghum was decreased over control 40– 70.4% whereas root length was decreased over control 48.8–73.7% at saline stress. Dehnavi *et al.*, (2020) also found that dry weight of PAYAM genotypes of sorghum was decreased over control 72% at saline stress. Similar declines in growth metrics were recorded in *Catharanthus roseus* (Jaleel *et al.*, 2008a, b), *Withania somnifera* (Jaleel *et al.*, 2008c) and in *Salvodora persica* (Dagar and Kumar, 2004) under saline conditions. All these outputs displayed conformity with the present findings.

Physiological traits evaluation under salinity stress: Soil salinity became a critical problem worldwide among various environmental stresses because of its intense effects on physiology and enactments (Flowers et al., 1997). All environmental stresses directly or indirectly effect the osmotic pressure of the plants (Hannan et al., 2020; Nupur et al., 2020). Reduced photosynthetic activity and a reduced stomatal aperture are two physiological changes brought on by osmotic stress at the early stages of salinity stress (Rahnama et al., 2010). For revealing the salinity impact on photosynthetic responses, researchers quantify the amount of chlorophyll in the leaf. The use of Soil and plant analysis development (SPAD) meter is also used for determination of greenness (indicated chlorophyll content indirectly) for leaf under stress condition (Negrão et al., 2017). Salinity hampers photosynthesis by destroying chlorophyll pigments and hindering the activity of photosystem II (Saddiq et al., 2021). Within the current study, SPAD value and total chlorophyll content of C. gigantea seedling were reduced dynamically in all salinity levels but proline content was increased with increasing salinity levels (0, 6, 8, 10 and 12 dS m⁻¹) (Table 5). Like all the studied seedling emergence and growth traits, reduction of SPAD value and total chlorophyll content were more in 12 dS m⁻¹ salinity treatment compared to control. However, with the increase of saline level, proline content was increased and it was observed that the highest amount of proline for C. gigantea found at 12 dS m⁻¹ compared to other saline conditions (Table 5). Proline content increases with the rise of salt concentration also reported in Salvadora persica (Dagar and Kumar, 2004), Catharanthus roseus (Osman et al., 2007), wheat (Bilkis et al., 2016) and Calotropis procera (Alaish et al., 2016). One of the most common alterations caused by salt stress in plants is accumulation of the amino acid proline, which is assumed to be involved in stress tolerance pathways, but its exact function is still controversial (Lutts et al., 1999). Ali et al., (2004) reported reduced chlorophyll concentrations in 18 modern rice genotypes with artificial salinization (EC = 8.5 dS m^{-1}) soil conditions. It was reported that the growth and chlorophyll levels of Calotropis procera plant were badly affected by NaCl stress (Al-Sobhi et al., 2006). Ibrahim (2013) also found that the salinity reduced chlorophyll content in Calotropis procera at 100% sea water as compared to control. Jaleel et al., (2008b) reported about 14% and 34% total chlorophyll content reduction under low and high salinity, respectively in Catharanthus roseus. These outputs are in agreement with the present findings.

Trait association: In this experiment, principal component analysis revealed the most suitable blend of the studied traits through the mean values, whereas the magnitude of described variation by particular attributes and treatment were displayed by vector length on PCA-biplot. A negative association of mean emergence time and proline content with other growth traits was found. PC1 clearly separated the performance of C. gigantea genotype under all salinity treatments (6, 8, 10 and 12 dS m⁻¹) from control condition (0 dS m^{-1}) (Fig. 2, Table 7). From this, it could be inferred that different salinity levels affected the growth and physiological performance of C. gigantea genotype. On the biplot, scattered position of the same treatment might be due to heterogeneity within the treatments. From correlation analysis, it was confirmed that mean emergence time and proline content was negatively correlated with other studied traits, whereas they showed positive correlation between them (Table 6). Earlier studies also reported significant correlation among germination, growth and physiological traits of various plant species (Majid et al., 2013; Saddig et al., 2021).

Conclusion

At artificially developed saline soil, seedling emergence, growth traits (shoot and root length, total dry mass, stem diameter, leaf number and area) and chlorophyll content were decreased as a result of NaCl salt stress. Mean emergence time and salinity levels rose along with a rise in proline content. This study concluded that *C. gigantea* is a salinity tolerant plant and it has a potential scope on commercial cultivation in coastal saline regions of the globe.

Acknowledgement

The financial support of this study provided by Institute of Reasearch and Training (IRT), Hajee Mohammad Danesh Science and Technology University, Dinajpur, Bangladesh (Project number 32 and Fiscal year 2020-2021) was gratefully acknowledged.

References

- Agnihotri, R.K., L.M.S. Palni and D.K. Pandey. 2006. Screening of land races of rice under cultivation in Kumaun Himalayan for salinity stress during germination and early seedling growth. *Ind. J. Plant Physiol.*, 11(30): 262-272.
- Ahmed, K.K.M., A.C. Rana and V.K. Dixit. 2005. Calotropis species (Asclepediaceae), a comprehensive review. Pharm. Mag., 1(2): 48-52.
- Alaish, F.M.Z., H.S.M. Al-Zahrani and S.Z.S. Hindi. 2016. Some metabolic activities of *Calotropis procera* habitated under high voltage-power lines. *Int. J. Innov. Res. Sci. Eng. Technol.*, 5(3): 2716-2726.
- Al-Ansari, F. and T. Ksiksi. 2016. A quantitative assessment of germination parameters: the case of *Crotalaria Persica* and *Tephrosia Apollinea. Open Ecol. J.*, 9: 13-21.
- Ali, Y., Z. Aslam, M.Y. Ashraf and G.R. Tahir. 2004. Effect of salinity on chlorophyll concentration, leaf area, yield and yield components of rice genotypes grown under saline environment. *Int. J. Environ. Sci. Technol.*, 1(3): 221-225.
- Almeida, I.V.B., M.M.D. Rêgo, F.R.C. Batista, E.R. Rêgo and R.L.A. Bruno. 2019. Phenology of *Calotropis procera* (Ait.)

W.T. Aiton accessions based on morphophysiological characteristics. *Revista Caatinga, Mossoró*, 32(2): 543-551.

- Alom, R., M.A. Hasan, M.R. Islam and Q.F. Wang. 2016. Germination characters and early seedling growth of wheat (*Triticum aestivum* L.) genotypes under salt stress conditions. J. Crop Sci. Biotech., 19(5): 383-392.
- Al-Sobhi, O.A., H.S. Al-Zahrani and S.B. Al-Ahmadi. 2006. Effect of salinity on chlorophyll & carbohydrate contents of *Calotropis procera* seedlings. *Sci. J. King Faisal Uni.*, 7(1): 105-115.
- Anonymous. 2010. Coastal saline soils of Bangladesh. Soil Resources Development Institute. Ministry of Agriculture, Dhaka, Bangladesh. pp. 96.
- Bahmani, M. and D. Kartoolinejad. 2017. Effect of salinity stress on growth, morphological and physiological characteristics of Milkweed (*Calotropis procera* Ait.) seedlings. *Arid Biomed. Sci. Res. J.*, 8(1): 27-35.
- Baskin, C.C. and J.M. Baskin. 1997. Seeds: Ecology, biogeography and evolution of dormancy and germination. 1st edition, Academic Press, New Delhi.
- Bates, L.S., R.P. Waldren and I.D. Teare. 1973. Rapid determination of free proline for water-stress studies. *Plant Soil*, 39: 205-207.
- Bera, A.K., M.K. Pati and A. Bera. 2006. Bassionolide ameliorates adverse effect on salt stress on germination and seedling growth of rice. *Ind. J. Plant Physiol.*, 11(2): 182-189.
- Bilkis, A., M.R. Islam, M.H.R. Hafiz and M.A. Hasan. 2016. Effect of NaCl induced salinity on some physiological and agronomic traits of wheat. *Pak. J. Bot.*, 48(2): 455-460.
- Bradford, K.J. 1995. Water relations in seed germination. In: (Eds.): Kigel, J. and G. Galili. Seed development and germination, Marcel Dekker, New York, pp. 351-396.
- Catalan, L., M. Balzarini, E. Taleisnik, R. Sereno and U. Karlin. 1994. Effects of salinity on germination and seedling growth of *Prosopis flexuosa* (D.C.). *Forest Ecol. Manag.*, 63: 347-357.
- Chauhan, B.S. and D.E. Johnson. 2009. Seed germination and seedling emergence of Synedrella (*Synedrella nodiflora*) in a tropical environment. *Weed Sci.*, 57(1): 36-42.
- Copeland, L.O. 1976. *Principles of seed science and technology*. Burgress Publishing Company, Minneapolis, Minnesota.
- Dagar, J.C. and H.B.Y. Kumar. 2004. Effect on growth performance and biochemical contents of *Salvadora persica* when irrigated with water of different salinity. *Ind. J. Plant Physiol.*, 9(3): 234-238.
- Dehnavi, A.R., M. Zahedi, A. Ludwiczak, S.C. Perez and A. Piernik. 2020. Effect of salinity on seed germination and seedling development of sorghum *(Sorghum bicolor (L.))* Moench) genotypes. *Agronomy*, 10(6): 859.
- El-Tantawy, H. 2000. Flowering and Fruiting Eco-physiology of *Calotropis procera* (Ait.) W.T. Ait, and importance of Gas in Fruit dehiscence. *Taeckholmia*, 20(1): 69-80.
- Flowers, T.J., A. Garcia, M. Koyama and A.R. Yeo. 1997. Breeding or salt tolerance in crop plants-role of molecular biology. *Acta Physiol. Plant.*, 19: 427-433.
- Francis, J.K. 2003. Calotropis procera. U.S. Department of Agriculture, Forest Service, International Institute of Tropical Forestry, Puerto Rico.
- Gill, P.K., A.D. Sharma, P. Singh and S.S. Bhullar. 2003. Changes in germination, growth and soluble sugar contents of *Sorghum bicolor* L. Moench seeds under various abiotic stresses. *Plant Growth Regul.*, 40: 157-162.
- Gulzar, S., M.A. Khan and I.A. Ungar. 2003. Salt tolerance of a coastal salt marsh grass. *Comm. Soil Sci. Plant Ana.*, 34(18): 2595-2605.
- Guo, J., M. Du, H. Tian and B. Wang. 2020. Exposure to high salinity during seed development markedly enhances seedling emergence and fitness of the progeny of the extreme Halophyte Suaeda salsa. Front. Plant Sci., 11: 1291.

- Hannan, A., M.N. Hoque, L. Hassan and A.H.K. Robin. 2020. Adaptive mechanisms of root system of rice for withstanding osmotic stress. In: *Recent Advances in Rice Research*, M.U.R. Ansari, 1-19.
- Heidarpour, M., B.M. Fard, A. Arzani, A. Aghakhani and M. Feizi. 2009. Effects of irrigation water salinity and leaching fraction on yield and evapotranspiration in spring wheat. *Comm. Soil Sci. Plant Ana.*, 40(15-16): 2521-2535.
- Hussain G., M.A. Qayyum and A.N. Alajaji. 1989. Calculation for the preparation of salt mixtures to develop artificial salinity for use in soils and waters. *Arid Soil Res. Rehabil.*, 3(1): 85-89.
- Ibrahim, A.H. 2013. Tolerance and avoidance responses to salinity and water stresses in *Calotropis procera* and *Suaeda aegyptiaca*. *Turk. J. Agric. For.*, 37: 352-360.
- Islam, M.R., Z.Z. Li, A.W. Gichira, M.N. Alam and P.C. Fu. 2019. Population genetics of *Calotropis gigantea*, a medicinal and fiber resource plant, as inferred from microsatellite marker variation in two native countries. *Biochem. Genet.*, 57: 522-539.
- Jaleel, C.A., B. Sankar, R. Sridharan and R. Panneerselvam. 2008b. Soil salinity alters growth, chlorophyll content, and secondary metabolite accumulation in *Catharanthus roseus*. *Turk. J. Biol.*, 32: 79-83.
- Jaleel, C.A., G.M.A. Lakshmanan, M. Gomathinayagam and R. Panneerselvam. 2008c. Triadimefon induced salt stress tolerance in *Withania somnifera* and its relationship to antioxidant defense system. S. Afr. J. Bot., 74 (1): 126-132.
- Jaleel, C.A., R. Gopi, P. Manivannan and R. Panneerselvam. 2008a. Soil salinity alters the morphology in *Catharanthus roseus* and its effects on endogenous mineral constituents. *Eur. J. Biosci.*, 2: 18-25.
- James, D.W., R.J. Hawks and J.J. Jurinak. 1983. *Modern Irrigated Soils*. John Wiley and Sons, New York.
- Jamil, M., C.C. Lee, S.U. Rehman, D.B. Lee, M. Ashraf and E.S. Rha. 2005. Salinity (NaCl) tolerance of brassica species at germination and early seedling growth. *Elect. J. Environ. Agri. Food Chem.*, 4 (4): 970-976.
- Jhala, Y.V. 1997. Seasonal effects on the nutritional ecology of blackbuck Antilope cervicapra. J. App. Ecol., 34(6): 1348-1358.
- Kanchan, T. and A. Atreya. 2016. *Calotropis gigantea*. Wilder. *Environ. Med.*, 27(2): 350-351.
- Karim, Z., S.G. Hussain and M. Ahmed. 1990. Salinity problems and crop intensification in the coastal regions of Bangladesh. *Soil Pub.*, 33: 1-63.
- Li, B., M.G. Gilbert and W.D. Stevens. 2015. *Calotropis gigantea* (Linnaeus) W. T. Aiton, Hortus Kew. Ed. 2. 2: 78.
- Lutts, S., V. Majerus and J.M. Kinet. 1999. NaCl effects on proline metabolism in rice (*Oryza sativa*) seedlings. *Physiol. Plant.*, 10(3): 450-458.
- Majid, A., S. Mohsen, A. Mandana, J.H. Saeid, E. Ezatollah and S. Fariborz. 2013. The effects of different levels of salinity and indole-3-acetic acid (IAA) on early growth and germination of wheat seedling. J. Stress Physiol. Biochem., 9(4): 329-338.
- Matsuura, A., S. Inanaga and K. Murata. 2005. Differences in the Vegetative Growth between Common and Tartary Buckwheat in Saline Hydroponic Culture. *Plant Prod. Sci.*, 8(5): 533-538.
- Mitchell, R.B. and K.P. Vogel. 2012. Germination and Emergence Tests for Predicting Switchgrass. *Agron. J.*, 104(2): 458-465.
- Moreira, E.C.F., D.S. Silva, W.E. Pereira, C.R.J Cabral and M.V.M. Andrade. 2007. Estimação da área foliar da flor de seda (*Calotropis procera*) (In English: Leaf area estimation in flor de seda (*Calotropis procera*)). Archivos de Zootecnia, Cordoba, 56(214): 245-248.

- Motaleb, M.A. 2011. Selected Medicinal Plants of Chittagong Hill Tracts. IUCN (International Union for Conservation of Nature), Dhaka, Bangladesh. pp. 116.
- Muriira, N.G. 2017. Genetic and epigenetic diversity and differentiation of *Calotropis* a fiber producing shrub. PhD, Kunming Institute of Botany, Chinese Academy of Sciences, China, 1-152.
- Negrão, S., S.M. Schmöckel and M. Tester. 2017. Evaluating physiological responses of plants to salinity stress. *Ann. Bot.*, 119(1): 1-11.
- Nimbolkar, P.K., R.M. Kurian, K.K. Upreti, R.H. Laxman and L.R. Varalakshmi. 2018. Seed germination and seedling growth responses of polyembryonic mango (*Mangifera indica* L.) genotypes to salinity stress. Int. J. Chem. Stud., 6(2): 3641-3648.
- Nupur, J.A., A. Hannan, M.A.U Islam, G.H.M. Sagor and A.H.K Robin. 2020. Root development and anti-oxidative response of rice genotypes under polyethylene glycol induced osmotic stress. *Plant Breed. Biotech.*, 8(2): 151-162.
- Orwa, C., A. Mutua, A. Kindt, R. Jamnadass and A. Simons. 2009. Agroforestry database: a tree reference and selection guide version 4.0[EB/OL]. World Agroforestry Center, Kenya.
- Osman, M.E.H., S.S. Elfeky, K.A. El-Soud and A.M. Hasan. 2007. Response of *Catharanthus roseus* shoots to salinity and drought in relation to vincristine alkaloid content. *Asian J. Plant Sci.*, 6: 1223-1228.

- Parhira, S., Z.F. Yang, G.Y. Zhu, Q.L. Chen, B.X. Zhou, Y.T. Wang, L. Liu, L.P. Bai and Z.H. Jiang. 2016. In vitro anti– influenza virus activities of a new lignan glycoside from the latex of *Calotropis gigantea*. *PLoS One*, 9(8): e104544.
- Rahnama, A., R.A. James, K. Poustini and R. Munns. 2010. Stomatal conductance as a screen for osmotic stress tolerance in durum wheat growing in saline soil. *Fun. Plant Biol.*, 37(3): 255-263.
- Saddiq, M.S., S. Iqbal, M.B. Hafeez, A.M. Ibrahim, A. Raza, E.M. Fatima, H. Baloch, P. Woodrow and L.F. Ciarmiello. 2021. Effect of salinity stress on physiological changes in winter and spring wheat. *Agronomy*, 11(6): 1193.
- Sharma, A.P. and B.D. Tripathi. 2009. Assessment of atmospheric PAHs profile through Calotropis gigantea R.Br. leaves in the vicinity of an Indian coal-fired power plant. *Environ. Monitor. Assess.*, 149: 477-482.
- Song, J., H. Fan, Y, Zhao, Y. Jia and X. Du. 2008. Effect of salinity on germination, seedling emergence, seedling growth and ion accumulation of a euhalophyte *Suaeda salsa* in an intertidal zone and on saline inland. *Aqu. Bot.*, 88(4): 331-337.
- Uddin, M.J. 2021. Salinity has spread over one and a half hundred km upstream. Prothom Alo, accessed 6 May 2021.
- Witham, F.H., D.F. Blaydes and R.M. Devlin. 1971. Chlorophyll absorption spectrum and quantitative determination. *Exp. Plant Physiol.*, 167-172.
- Yang, Q., W.H. Ye, X. Deng, H. Cao, Y. Zhang and K. Xu. 2005. Seed germination ecophysiology of *Mikania micrantha* H.B.K. *Bota. Bull. Acade. Sin.*, 46: 293-299.

(Received for publication 15 August 2022)