

DISCOVERY OF A NOVEL CRYSTAL PROTEIN FROM PAKISTANI *BACILLUS THURINGIENSIS* STRAIN TOXIC TO *TRIBOLIUM CASTENEUM* HERBST (COLEOPTERA:TENEBRIONIDAE)

KAUSAR MALIK AND SHEIKH RIAZUDDIN

*National Center of Excellence in Molecular Biology,
University of the Punjab 87-West Canal Bank Road,
Thokar Niaz Baig Lahore-53700, Pakistan*

Abstract

The biotoxicity analysis of crystal protein of some *Bacillus thuringiensis* strains has been carried out against the larvae of red flour beetle *Tribolium castaneum* Hebst. Seven isolates were found highly active against *T. castaneum*. The most toxic isolate SG31.11 has calculated LC₅₀ value of 0.2 ug/mg of artificial diet. Presently, the active protein of isolate SG31.11 was sequenced and data showed that it resembles with a novel Cry3 protein.

Introduction

Tribolium castaneum, is a serious pest of stored grains throughout the world and also a genetic model for the Coleoptera. It not only affects the quantity but also the quality of stored grains. The quantitative estimation of the loss incurred by red flour beetle is difficult because this insect is found in flourmills, godowns and warehouses with other associated stored grain pest complex. To control the infestation of this insect, many synthetic pesticides have been used for several years. However, these pesticides produce several adverse effects, which include accumulation of lethal chemicals in food chain and environment, lack of selectivity towards beneficial insects and evolution of resistance. These factors have directed the attention of scientists from traditional chemical pesticides to biopesticides.

Microbial control of insect pest of crops using entomopathogens is an ecologically sound pest management strategy. Although insect viruses and fungal pathogens are used as microbial control agents, but *Bacillus thuringiensis* Berliner (Bt) appears to have the greatest potential for this purpose. This gram-positive, spore forming crystalliferous bacterium synthesizes a proteinaceous parasporal crystalline inclusion (δ -endotoxins) during the sporulation phase. These crystalline proteins are highly specific against different insect orders, and non-target organisms like parasitoids, predators and vertebrates are not affected by their use (Aronson *et al.*, 1986: Whiteley & Schnepf., 1986). A promising variety of crystal proteins (Cry proteins) have been recognized in different Bt strains. Of these crystal proteins Cry3 are reported to be toxic against coleoptera. Our previous study presented initial efforts to assess the potential of Bt strains isolated from different environmental samples, as a biological control agent of *T. castaneum* (Malik & Riazuddin., 2000).

In the present study, the active protein of isolate SG31.11 was sequenced and data showed that it resembles with a novel Cry3 protein of *Bacillus thuringiensis* serovar japonensis strain Buibui, (toxic to larvae of the cupreous chafer, *Anomala cuprea* reported by Sato *et al.*, (1994). Sequence of this gene is the same but our locally isolated Bt and target pest (*T. castaneum*) is different.

Materials and Methods

All the organic and inorganic chemicals used were from Sigma Chemical Company. Molecular weight protein markers, polyvinylidene difluoride (PVDF) membrane and Bradford protein assay reagents were from Bio-Rad Laboratories. *Tribolium castaneum* larvae were obtained from the insectory of the CEMB. All protein concentrations were measured by Bio-Rad protein assay with bovine serum albumin as standard (Bradford, 1976). Cry3A clones were obtained from culture collection lab of CEMB.

Purification of insecticidal crystal proteins: Cry3A and SG31.11 proteins were purified by the procedure described by Lee *et al.*, (1992). Purified proteins were solubilized in 50mM Na₂CO₃ pH 9.5, containing 10mM dithiothreitol treated with 5% trypsin at 37°C for 4-hours and stored at 4°C.

Biotoxicity assay: Biotoxicity assay was determined as reported by Malik & Riazuddin (2000). The most toxic isolate SG31.11 has calculated LC₅₀ value of 0.2 ug/mg of artificial diet.

Protein purification and sequencing: Protein from *Bacillus thuringiensis* SG31.11 strain solubilized in alkaline buffer was resolved on non-denaturing protein gel. Protein band was excised from the gel to elute the protein. The eluted protein was transferred onto a PVDF membrane using semidry transblot apparatus, stained with Coomassie brilliant blue and used in amino acid sequencing by Edman degradation.

Results

Screening microbial collections to search for novel Bt proteins: Seven Bt were found toxic against *T. castaneum* during screening of Bt in the Pakistani environment to search for novel Bt proteins. Locally isolated Bt SG31.11 is highly toxic to the larvae in biotoxicity assays.

Amino acid sequencing: Purified protein sequenced by Edman degradation method of amino acid sequencing.

Homology search: When the protein was sequenced by Edman degradation, yielded amino acid sequence which was searched for homology with other sequences in GenBank using Blastx, at <http://www.ncbi.nlm.nih.gov/BLAST/>.

There was significant sequence homology to a novel 130-kDa crystal protein antigen of *Bacillus thuringiensis* serovar japonensis strain Buibui in the database, under the Accession number U04366.

Amino acids sequence

MSPNNQNEYEIIDALSPTSVDNSIRYPLANDQTNLQNMNYKDYLKMTTESTNA
ELSRNPGTFISAQDAVGTGIDIVSTIISGLGIPVLGEVFSILGSLIGLLWPSNNEVW
QIFMNRVEELIDQKILDSVRSRAIADLANSRIAVEYYQNALEDWRKNPHSTRSAA
LVKERFGNAEAILRTNMGSFSQTNYETPLLPTYAQAASLHLLVMRDVQIYGKEW
GYPQNDIDLFYKEQVSYTARYSDHCVQWYNAGLNKLRGTGAKQWVDYNRFR

EMNVMVLDLVALFPNYDARIYPLETNAELTREIFTPVGSYVTGQSSTLISWYDM
 IPAALPSFSTLENLLRKPdffTLLQEIRMYTSFRQNGTIEYYNYWGGQRLTLSYIY
 GSSFNKYSGVLAGAEDIIPVGQNDIYRVVWYIIGRYTNSLLGVNPVTFYFSNNTQ
 KTYSKPKQFAGGIKTIDSGEELTYENYQSYSHRVSYITSFEIKSTGGTVLGVVPIFG
 WTHSSASRNNFIYATKISQIPINKASRTSGGAVWNFQEGLYNGGPVMKLSGSGSQ
 VINLRVATDAKGASQRYRIRIRYASDRAGKFTISSRSPENPATYSASIAYTNTMST
 NASLTYSTFAYAESGPINLGISGSSRTFDISITKEAGAANLYIDRIEFIPVNTLFEAEE
 DLDVAKKAVNGLFTNEDALQTSVTDYQVNQAANLIECLSDELYPNEKRMLWDA
 VKEAKRLVQARNLLQDTGFNRINGENGWTGSTGIEVVEGDVLFKDRSLRLTSAR
 EIDTETYPTYLYQQIDESLLKPYTRYKLKGFIGSSQDLEIKLIRHRANQIVKNVDPN
 LLPDVRPVNSCGGVDRCSQQYVDANLALENNGENGMSSDSHAFSFHIDTGEI
 DLNENTGIWIVFKIPTTNGNATLGNLEFVEEGPLSGETLEWAQQQEQQWQDKMA
 RKRAASEKTYAAKQAIDRLFADYQDQKLNSGVEMSDLLAAQNLVQSIPYVYN
 DALPEIPGMNYTSFTEL TNRLQQA WNL YDLQNAIPNGDFRNGLSNWNATSDVN
 VQQLSDTSVLVIPNWSQVSQQFTVQPNYRYVLRVTARKEGVGDGYVIIRDGAN
 QTETLTFNICDDDTGVLSTDQTSYITKTVEFTPSTEQVWIDMSETEGVFNIESVEL
 VLEEE"

Nucleotide sequence

atgagtcaa ataatacaaaa tgagtatgaa attatagatg cttatcacc cactctgta tccgataatt ctattagata
 tectttagca aacgatcaaa cgaacacatt acaaaacatg aattataaag attatctgaa aatgaccgaa tcaacaatg
 ctgaattgtc tcgaaatccc gggacattta ttagtgcgca ggatgcggtt ggaactggaa ttgatattgt tagtactata
 ataagtgggt tagggattcc agtgcttggg gaagtctct caattctggg tcattaatt ggcttattgt ggccgtcaaa
 taatgaaaat gtatggcaaa tatttatgaa tcgagtggaa gagctaattg atcaaaaaat attagattct gtaagatcaa
 gagccattgc agatttagct aattctagaa tagctgtaga gtactatcaa aatgcactg aagactggag aaaaaacca
 cacagtacac gaagcgcagc acttgtaaag gaaagattg gaaatgcaga agcaattta cgtactaaca tgggttcatt
 ttctcaaag aattatgaga ctccactctt accacatat gcacaggcgc cctctctgca ttgcttga atgagggatg
 tcaaattta cgggaaggaa tggggatc ctcaaatga tattgacctt tttataaag aacaagtac ttatacggct
 agatattccg atcattgcgt ccaatgttac aatgctggtt taataaatt aagaggaacg ggtgctaagc aatgggtgga
 ttataatcgt ttccgaagag aaatgaatgt gatggtattg gatctagttg cattattcc aaactacgat gcgcgtat
 atccactgga acaaatgca gaacttaca gagaaattt cacagatcct gttggaagt acgtaactgg acaatcgagt
 acccttat ctggtacga tatgattcca gcagctctt cttcatttc aacgctcgag aacctacta gaaaacctga
 tttcttact ttgctgcaag aaattagaat gtatacaagt tttagacaaa acggtacgat tgaatattat aattattggg
 gaggacaaag gtaaccctt tcttatct atggttctc attcaataa tatagtggg ttctgcccgg tgctgaggat
 attattctg tgggtcaaaa tgatattac agagtgtat ggacttat aggaaggtac acgaatagc tgctaggagt
 aatccagt acttttact tcagtaataa tacacaaaaa acttattcga agccaaaaca atcgcgggt ggaataaaaa
 caattgattc cggcgaagaa ttaacttac aaaattatca atcttatagt cacagggtaa gttacattac atctttgaa
 ataaaaagta cgggtgttac agtattagga gtatgtccta tatttggtg gacgcatagt agtgccagc gcaataact
 tattacgca acaaaaatct cacaaatccc aatcaataa gcaagtagaa ctacgggtgg agcggttgg aattccaag
 aaggtctata taatggagga cctgtaataa aattatctgg gtctggttcc caagtaataa acttaagggt cgcaacagat
 gcaaaggag caagtcaag atactgtatt agaactcagat atgcctctga tagagcgggt aaatttacga tatctccag
 atctccagag aatcctgca cctattcagc ttctattgct tatacaata ctatgtctac aatgctct ctaactgata
 gtactttgc atatgcagaa tctggccta taaacttagg gatttcggga agttcaagga ctttgatat atctattaca
 aaagaagcag gtgctgctaa cctttatatt gatagaattg aattattcc agttaatagc ttatttgaag cagaagaaga
 cctagatgtg gcaaagaaag ctgtgaatgg cttgtttac aatgaaaag atgccttaca gacaagtga acggattatc
 aagtcaatca agcggcaaac ttaatagaat gcctatccga tgagttatc ccaatgaaa aacgaatgt atgggatgca
 gtgaaagagg cgaacgact tgttcaggca cgtaacttac tccaagatagc aggctttaat aggattaatg gagaaaacgg

atggacggga agtacgggaa tcgaggtgt ggaaggagat gttctgtta aagatcggtc gcttcgttg acaagtgcga
gagagattga tacagaaaca taccacgt atctctatca acaaatagat gaatcgcttt taaaccata tacaagatat
aaactaaaag gttttatagg aagtagtcaa gatttagaga ttaaattaat acgtcatcgg gcaaatcaaa tcgtcaaaa
tgtaccagat aatctctgc cagatgtacg ccctgtcaat tcttgggtg gagtcgatcg ctgcagtga caacagtatg
tagacgcgaa tttagcactc gaaacaatg gagaaaatgg aatatgtct tctgattccc atgcatttc ttccatatt
gatacgggtg aatatagattt gaatgaaaat acaggaattt ggatcgtatt taaaattccg acaacaatg gaaacgcaac
actaggaaat ctgaatttg tagaagaggg gccattgtca ggggaaacat tagaatgggc ccaacaaca gaacaacaat
ggcaagacaa aatggcaaga aaacgtgcag catcagaaaa aacatattat gcagcaaagc aagccattga tcgtttatc
gcagattatc aagacaaaa acttaattct ggtgtagaaa tgcagattt gttggcagcc caaaccttg tacagtccat
tccttacgta tataatgatg cgttaccgga aatccctgga atgaactata cgagttttac agagttaaca aatagactcc
aacaagcatg gaattgtat gatcttcaaa acgtatacc aatggagat ttcgaaatg gattaagtaa ttggaatgca
acatcagatg taaatgtgca acaactaagc gatacatctg tcttgtcat tccaaactgg aattctcaag tgcacaaca
attacagtt caaccgaatt atagatatgt gttacgtgc acagcgagaa aagagggagt aggagacgga tatgtgatca
tccgtgatgg tgcaaatcag acagaaacac tcacatttaa tatatgtgat gatgatacag gtgtttatc tactgatcaa
actagctata tcacaaaaac agtggaaatc actccatcta cagagcaagt ttggattgac atgagtgaga ccgaaggtgt
attcaacata gaaagtgtag aactcgtgtt agaagaagag taa

Discussion

The use of Bt in controlling insect pests has increased over the past few decades. With the expansion of biotechnology in crop sciences use of Bt toxins is becoming a common practice (Sanchis & Lereclus, 1999). New variants of Bt with interesting toxicity spectra are also appearing. The search for Bt strains with novel toxicity, coupled with a more complete understanding of the toxins and their associated proteins, is paramount to current efforts to harness fully the potential of Bt technology. Shoaib *et al.*, (2008) also revealed that *B. thuringiensis* has a great potential to prove as an alternative to methyl bromide for controlling coleopteran pests including *T. castaneum*. They concluded that there is a dire need to reach and test more and more isolates for tracing out the best to achieve maximum control of this pest.

During previous research, which was aimed at exploring the diversity of Bt in the Pakistani environment to search for novel Bt proteins, 7 Bt were found toxic against *T. castaneum*. The aim of the present study was to characterize Bt isolates toxic to coleopteran insect pests.

We report that locally isolated Bt SG31.11 is highly toxic to the larvae of *T. castaneum* and its protein sequence is identical with novel Cry3 crystal protein in the database, under the Accession number U04366. When we determined protein sequence, a report appeared by Sato *et al.*, (1994) which showed proteins sequence similarity in Pakistani *Bacillus thuringiensis* strain SG31.11 and *Bacillus thuringiensis* serovar japonensis strain Buibui.

Sato *et al.*, (1994) revealed that novel Cry3 protein of *Bacillus thuringiensis* serovar japonensis strain Buibui is toxic to larvae of the cupreous chafer (*Anomala cuprea*) which is a scarabaeid insect. However, there is homology between protein sequences of Pakistani *B. thuringiensis* strain SG31.11 and *B. thuringiensis* serovar japonensis strain Buibui, but our locally isolated *B. thuringiensis* and target pest (*T. castaneum*) is different. *T. castaneum* belongs to the family Tenebrionidae of the order Coleoptera.

The information obtained from these studies will be helpful in adopting strategies for controlling the insect pests of commercially important crops, better suited to Insect Resistance Management (IRM).

References

- Aronson, A.I., W. Beckman and P. Dunn. 1986. *Bacillus thuringiensis* and related insect pathogens. *Microbiol. Rev.*, 50: 1-24.
- Bradford, M.M. 1976. A rapid alkaline extraction procedure for screening recombinant plasmid DNA. *Anat. Biochem.*, 72: 248-254.
- Lee, M.K., R.E. Milne, A.Z. Ge and D.H. Dean. 1992. Location of *Bombyx mori* receptor binding region on a *Bacillus thuringiensis* δ -endotoxin. *J. Biol. Chem.*, 267: 3115-3121.
- Malik, K. and S. Riazuddin. 2000. Insecticidal activity of some *Bacillus thuringiensis* strains against *Tribolium castaneum* (Coleoptera; Tenebrionidae). *Punjab Univ. J. Zool.*, 15: 31-34.
- Sanchis, V. and D. Lereclus. 1999. *Bacillus thuringiensis*: A biotechnology model. *J. Soc. Biol.*, 193: 523-530.
- Sato, R., K. Takeuchi, K. Ogiwara, M. Miname, Y. Kaje., M. Suzuki, H. Hori., S. Asano, Morba and H. Inahana. 1994. Cloning, heterologous expression and localization of a novel crystal protein gene from *Bacillus thuringiensis* serovar japonensis strain buibui toxic to scarabaeid insects. *Curr. Microbiol.*, 28: 15-19.
- Shoab, M., A. Anil, M.I. Haque, S.N. Afzal, I. Umer and S. Naz. 2008. Scope of commercial formulation of *Bacillus thuringiensis* Berliner as an alternative to methyl bromide against *Tribolium castaneum* adults. *Pak. J. Bot.*, 40(5): 2149-2156.
- Whiteley, H.R. and H.E. Schnepf. 1986. The molecular biology of parasporal crystal body formation in *Bacillus thuringiensis*. *Ann. Rev. Microbiol.*, 40: 549-576.

(Received for publication 2 January 2009)