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ROLE OF GA3 AND KNO3 IN IMPROVING THE FREQUENCY OF SEED GERMINATION IN *PLANTAGO LANCEOLATA* **L.**

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Abstract

Seeds of *Plantago lanceolata* L., obtained from crops of 1992- 2001 were treated with 100, 200 and 400 ppm of GA₃ and 1000, 2000 and 4000 ppm of KNO₃ by incubating them at 20 °C under 16 h light photoperiod, along with control. Highest germination of 97% was obtained from the seed of the crop of 2001, which decreased to 78.8% in the seeds of the crop of 1992 in untreated control. It was found that age plays a significant role in the decline of frequency of seed germination, which was effectively improved with the use of GA₃ and KNO₃.

Introduction

Plantago lanceolata L., of the family *Plantaginaceae* is a perennial herb that grows upto 70 cm, has lanceolate leaves, deeply venated whitish flowers and has egg shaped fruit capsules. The plant dies to the ground each winter and sprouts anew from its fibrous tap root around mid-spring. The plant prefers saline and wet habitiats upto 2200 meters. The leaves may grow up to about 15 cm long and 10 cm wide. (McCloud & Berenbaum, 2000). It is widely found in Turkey and most of the European countries (Davis, 1975). In Germany, France and Belgium it is cultivated on limited areas for forage (Karl, 1982). Recently, domesticated cultivars have been selected for pasture production in New Zealand (Sanderson & Elwinger, 2000a).

Medicinally, *P. lanceolata* is astringent, demulcent, emollient, cooling, vulnerary, expectorant, antimicrobial, antiviral, antitoxin and diuretic (Leung & Foster 1996) and is effectively used in treatment of tumors (Fracoise *et al.*, 1998). Traditionally, its leaf tea is used for diarrhoea, coughs and dysentery. Leaves are also applied to blisters, sores, swelling and insect stings. The mucilage from any plantain seeds may decrease cholesterol level in blood (Foster & Duke, 1990).

In practice, seed germination rate is very important for successful growing of any crop. Most crop seeds are probably dead after 25 years, even under favorable storage conditions (Martin & Leonard, 1967). However, there is a correlation between longevity of seed and storage conditions (Hughes & Metcalfe, 1972). The old seeds lose their germination ability, with the passage of time. However, it is possible to extend seed germination ability by using some seed germination promoters. Gibberellins and potassium nitrate (KNO₃) are used for breaking seed dormancy and promoting seed germination. Gibberellins are most directly implicated in the control and promotion of germination. Gibberellins promote growth by increasing plasticity of the cell wall followed by the hydrolysis of starch to sugar which reduces the potential in the cell, ^{*}Institute of Biotechnology, Department of Field Crops, Faculty of Agriculture, University of Ankara, Dışkapı, Ankara, Turkey (Corr. author Tel: +90-312-5961286, +90 312 5961467-1467, Fax: +90-312-3179815, Email: kmkhawar@gmail.com, gurbuz@agri.ankara.edu.tr

resulting in the entry of water into the cell causing elongation (Arteca, 1996). There is evidence that natural gibberellin like substances appear during successive stages of afterripening and germination. Gibberellins play an important role in the regulation of seed germination. KNO₃ is most widely used chemical for promoting germination. Solutions of 0.1 to 0.2 % KNO₃ are common in routine germination testing and are recommended by the Association of Official Seed Analysts and the International Seed Testing Association for germination tests of many species (Copeland & McDonald, 1995; Basra, 1994). The objective of the study was to eastablish a relationship between seed age and germination percentage in *Plantago lanceolata* with emphasis on simplicity and practicability. The protocol determines and evaluates reduction in germination due to metabolic changes affecting germination and offers effective, cheap and simple procedure using Gibberellic acid and KNO₃ to improve germination of long stored *P. lanceolata* seeds stored at room temperature.

Materials and Methods

Seed collection and storage

Seeds of *P. lanceolata* from the crops of 1991 to 1992, were collected from the nursery of perennial medicinal and aromatic plants of the department of Field Crops, Faculty of Agriculture, University of Ankara, Turkey and stored at room temperature.

GA₃ and KNO₃ treatment and germination

Batches of 100 seeds each contained in Petri dishes (100 x 10 mm) replicated four times, were treated with three doses each of 100, 200 and 400 ppm of GA₃ for 6 hours and 1000, 2000 and 4000 ppm of KNO₃ for 6 hours and were germinated in between moistened filter papers. The filter papers were humidified regularly with demineralized water. Seeds without any treatment were used as the control. They were incubated in a growth chamber (Sanyo MLR 350 H) at 20 °C under 16 h light photoperiod. The seed were scored for final germination after 21 days according to ISTA, (Anon., 1993).

Significance was determined by analysis of variance (ANOVA) using SPSS for windows (v. 11. SPSS Inc USA) and differences between the means of the seed age and the treatments were compared by Duncan's multiple range test (P<0.05). Data given in percentages were subjected to arcsine (\sqrt{X}) transformation before statistical analysis (Snedecor & Cochran 1967).

Results and Discussion

Frequency of seed germination of untreated seeds

The results indicated no seed dormancy but a sharp reduction in seed germination frequency as influenced by the seed age (Table 1). The seed germination reduced gradually with age with range of 78.8 to 97%. The data further indicated statistically similar frequency of seed germination from the seeds of 1999 to 2001; but different from those of 1995 to 1998, 1993-94 and 1992 (p<0.05). The statistical data showed a sharp decline of germination within each group such that the highest seed germination of 97% from the seeds of 2001 reduced to 78.8%; the lowest from the seeds of 1992.

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		Treatments					
Years	Control	GA ₃	GA ₃	GA3	KNO ₃	KNO ₃	KNO ₃
		100 ppm	200 ppm	400 ppm	1000 ppm	2000 ppm	4000 ppm
2001	97.0 a [*] 1 [*]	92.8 ab2	92.8 a2	92.0ab2	92.5 ab2	93.3 abc2	94.8 a12
	± 0.408	± 1.887	±1.377	± 0.408	± 0.866	± 1.377	± 1.315
2000	95.5 a1	94.8 a1	93.3 a1	93.0ab1	93.5 ab1	93.0 abc1	92.8 ab1
	± 0.289	± 0.479	± 0.250	± 0.707	± 0.289	± 0.408	± 0.479
1999	94.3 a1	94.3 ab1	92.8 a1	94.0 a1	91.3 bc12	92.3 bc12	90.5 bc 2
	± 0.750	± 0.957	± 0.250	± 0.577	± 1.109	± 1.315	± 0.866
1998	88.5 b2	92.8 ab1	92.5 a1	90.3 ab12	93.5 ab1	94.3 abc1	91.3 abc12
	± 1.041	± 1.377	± 0.957	± 0.854	± 1.041	± 0.479	± 0.854
1997	85.3 bc 2	93.0 ab1	93.0 a1	91.3 ab1	92.8 ab1	92.0 bc1	93.8 ab1
	± 0.629	± 1.155	± 1.080	± 1.315	± 1.797	± 1.080	± 1.315
1996	86.0 bc2	94.3 ab1	92.0 a1	93.3 ab1	95.3 al	96.0 al	94.0 ab1
	± 1.291	± 0.479	± 0.408	± 0.479	± 1.377	± 0.408	± 1.291
1995	85.0 bc2	93.5 ab1	92.0 a1	91.0 ab1	93.8 ab1	94.8 ab1	93.5 ab1
	± 1.225	± 1.893	± 1.225	± 1.354	± 1.493	± 1.109	± 1.190
1994	84.6 c2	90.8 b1	91.3 al	92.3 ab1	88.8 c1	91.0 cl	91.8 ab1
	± 0.479	± 0.854	± 0.750	± 0.854	± 2.097	± 1.472	± 1.250
1993	83.5 c2	90.5 b1	92.0 al	93.8 a1	91.0 bc1	90.8 c1	93.0 ab1
	± 0.866	± 1.555	± 1.080	± 0.629	± 1.080	± 1.315	± 1.080
1992	78.8 d2	86.3 c1	87.5 b1	89.8 b1	84.3 d2	85.0 d2	88.3 c1
	± 0.629	± 1.931	± 2.723	± 1.652	± 1.315	± 1.225	± 1.185
*Letters show vertical groups, figures show horizontal groups in each treatment.							

Table 1. Effects of GA₃ and KNO₃ treatments on seed germination of *Plantago lanceolata* belonging to the crops of 1991–2001.

**Mean value followed by different letters and figures are significant at the 0.05 level

Effects of GA₃ treatment on germination

A significant improvement in GA₃ treated seeds over untreated seeds was observed with a range of 86.3-94.8 % with 100 ppm, 87.5-93.3 % with 200 ppm and 89.8-94.0 % with 400 ppm treatment (Table 1). A visible improvement in frequency of seed germination of GA₃ treated seeds over control was observed on seeds belonging to the crops of 1992-1998. However, the GA₃ treatment in any concentration resulted in decline in the seed germination from the crops of 1999 to 2001. A 100 ppm GA₃ treatment improved seed germination of the seeds of the crops of 1995- 1998 and 400 ppm GA₃ treatment effectively improved seed germination of the crops of 1992 to 1994.

Effect of KNO₃ treatments on germination

The treated seeds showed germination frequency range of 84.3-95.3, 85.0-96.0 and 88.3-94.8 % from 1000 ppm, 2000 ppm and 4000 ppm KNO₃ treatment, respectively. Like GA₃, any of the KNO₃ treatment also was ineffective to improve germination in the seeds of crops belonging to 1999 to 2001 over control (Table 1). However, all doses of KNO₃ improved germination frequency over control, on the seeds belonging to the crops of 1992-1998 periods. Although improvement in the germination of KNO₃ treated seeds was not consistent yet 2000 ppm KNO₃ treated seeds showed the highest germination percentage over control in general.

 GA_3 and KNO₃ treatments showing decrease in germination in seeds of 1999-2001 may be due to endogenous or exogenous plant growth substances based primarily on corresponding shifts in threshold water potential for radicle emergence in line with the results of Ni & Bradford (1993). The progressing seed age resulted in corresponding decrease in the rate of germination especially from seeds belonging to the crops of 1991 to 1998, where less than 90 percent germination was very evident in line with the results of Tosserams *et al.*, (1997) and Davis & Robinson (2001), who observed reduction in seed germination with age. However, a decline in germination in aged seeds reported here was improved using either GA_3 or KNO₃ supporting the results of Saruhan *et al.*, (2002), who found that GA_3 application promoted the seed germination in *P. major*.

Sanderson & Elwinger (2000b), reported that the depth of planting was effective on seedling emergence and recorded 62 % of emergence value in this species. Beside the planting depth, the seed with high germination rate is necessary for good seedling emergence under field conditions. In conclusion, GA_3 treatment seems preferable over KNO₃ to obtain high germination in *P. lanceolata*.

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