

HIGH FREQUENCY REGENERATION FROM SCUTELLUM DERIVED CALLI OF BASMATI RICE cv. BASMATI 385 AND SUPER BASMATI

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Abstract

This study was conducted to obtain a high frequency regeneration from Basmati 385 and Super Basmati, which is a pre-requisite for transformation protocol. Seed was the explant source used in this study. Callus induction was obtained from N₆ media with 2 mg l⁻¹ 2,4-D. Super Basmati exhibited high callus induction efficiency (93.3%) followed by Basmati 385 (90.2 %). Twenty one days old maintained calli when transferred on MS medium with different combinations of auxin-cytokinin for regeneration showed 90% frequency of plant regeneration for Super Basmati with NAA 1mg l⁻¹ and BAP 2.5 mg l⁻¹ and 83% for Basmati 385 on MS medium supplemented with NAA 1 mg l⁻¹ and BAP 5 mg l⁻¹.

Introduction

Aside from *Arabidopsis* in dicot, rice (*Oryza sativa* L.) has been widely used as a plant material serving as a model system for plant genomics and *in vitro* studies in monocots. Due to its economic importance as a staple food for 2.7 billion people, rice has been a more attractive target for developing transgenic rice (Cho *et al.*, 2004). Almost 90% of total rice is grown in Asia. Besides being an important food of people of Pakistan, rice has also gained a respectable position as a foreign exchange earning commodity (Salim *et al.*, 2003).

In Pakistan, rice is cultivated over an area of 2225.2 million ha with a production of 4478.5 million tons giving an average yield of 2013 kgs per hectare. Total area under basmati production is 1377.3 hectare in which 1316.8 hectare is for Punjab. Total production of basmati rice is 2175.5 million tons; Punjab contributes 2304.2 million tons with a 1673 kgs per hectare yield of basmati rice (Anon., 2002-2003).

Pakistani rice especially basmati rice is a major commercial variety grown in rice growing areas of Pakistan and is famous in the world for its particular aroma. Basmati rice, the best quality scented rice produced in Pakistan commands the international market have four times greater price than in the domestic market. Pakistan exports 7% of the total world market (Rashid *et al.*, 2001). Despite all these, yield per unit area of rice in Pakistan is far below from the world average and low from many neighboring countries. The production of basmati rice is severely affected by various stresses, including fungal, bacterial and viral diseases which is one of the reasons for the low yields of rice. Hence, there is a need to improve these commercial cultivars by biotechnological applications.

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Whatever transformation system would be employed, efficient systems for embryogenic callus induction and shoot regeneration have been considered the basic matter in obtaining fertile transgenic rice. Successful callus induction from rice seeds has been reported by several researchers (Gonzalez, 2000., Navraj *et al.*, 1999., Marassi 1996., Valdez *et al.*, 1997., Xie *et al.*, 1995). Among various tissues of rice, scutella-derived callus of mature seeds have been frequently employed as target materials for genetic transformation by *Agrobacterium* mediated and biolistics methods because of the manipulative effectiveness (Chen *et al.*, 1998; Hashizume *et al.*, 1999; Hiei *et al.*, 1994; Rashid *et al.*, 1996., Cho *et al.*, 2004). Plant regeneration via somatic embryogenesis has many advantages in theoretical and applied tissue culture (Vasil *et al.*, 1982). The application of this technique has made rapid progress in dicotyledons, but significantly low in gramineae (Yamada, 1977). Cereals in general, are difficult to manipulate *in vitro* and regeneration is drastically reduced in successive subcultures. Regeneration of plants was achieved from callus of various origins eg., regenerated plants from seed callus (Tamura, 1968), and from embryo callus (Bajaj & Bidani, 1980). Most of the reported work has been done on “Japonica” or “Javanica” varieties and still there is a great deal of work to be done on optimization of media for plant regeneration in “Basmati” varieties, especially those that are most commonly cultivated in Pakistan. Although there are a few reports available on callus induction, regeneration and transgenic plant production in other Basmati cultivars like Basmati 6129, Basmati 370 and Basmati 385 (Rashid *et al.*, 1996). Rashid *et al.*, (2001 and 2004) reported a low regeneration frequency in Super Basmati, but still there is a strong need for more studies on developing high frequency regeneration from these commercially grown varieties

Enthused by the success in *Agrobacterium* mediated transformation, a study was initiated at the Agricultural Biotechnology Programme, National Agricultural Research Center, Islamabad, Pakistan to develop a high frequency callus induction and regeneration system from scutellum derived calli of two rice varieties viz., Basmati-385 and Super basmati for genetic manipulation.

Materials and Methods

Mature seeds of Basmati rice varieties Basmati 385 and Super Basmati were sterilized by protocol as described by Rashid *et al.*, (1996). Treated seeds of Basmati 385 and Super Basmati were dehusked. Seeds were surface sterilized by 70% ethanol for one minute followed by 50% Clorox (*Sodium hypochlorite*) for 20 min. Seeds were continuously shaken during treatment with Clorox and subsequently rinsed with autoclaved distilled water for 3-4 times after a regular interval of 5 min.

The sterilized seeds were aseptically inoculated on N₆ medium (Chu *et al.*, 1975) with 30 g l⁻¹ sugar, 2, 4-D 2 mg l⁻¹ and 6 g l⁻¹ of agar which was used as solidifying agent. The pH of the medium was adjusted to 5.8 before autoclaving at 121°C for 20 min. These inoculated seeds were placed in the growth room for three weeks for callus induction. Only embryogenic calli (white to light yellowish in color, compact and friable) were selected and non-embryogenic calli (mucilaginous and smooth) were discarded. Three weeks old scutellum derived calli were transferred on to maintenance medium which contained N₆ salts and vitamins supplemented with 2 mg l⁻¹ of 2, 4-D, 30 g l⁻¹ sugar and agar 6 g l⁻¹ as a solidifying agent. Calli were placed on maintenance medium for about two weeks.

Media used for regeneration contained basic MS salts and vitamins (Murashige & Skoog, 1962), 3% sucrose and 0.4% gelrite. Regeneration media contained different combinations of BAP and NAA as growth regulators. Maintained calli were placed on regeneration media and were placed in the growth room (light and dark period at $23 \pm 2^\circ\text{C}$ with 16/8h photoperiod with light intensity of 16 m.mol.m.s). After shoot regeneration, plantlets were transferred to MS media without growth regulators for root initiation. In order to determine regeneration frequency, 40-60 cultures were raised for each treatment and each experiment was performed thrice. Subculture period was maintained at 2-3 weeks intervals. Observations were taken every week on the basis of % age of regeneration for the development of shoot/culture and shoot development.

Statistical analysis

All resulting scales of total callus browning, callus differentiation, production of green tissue or spots and plantlet formation among the two rice cultivars with 9 treatments along with three replicates on the MS medium were statistically analyzed using Analysis of Variance technique with a computer program MSTAT-C. Treatment means were compared using Duncan's Multiple Range Test (DMRT).

Establishment of plantlets in the glass house

Well-rooted plantlets were washed with running tap water to remove agar from the surface of roots. They remained suspended in water in test tubes for a few days before transfer to pots in the glass house. The soil mixture used for hardening was ordinary soil taken from rice fields and a high percentage 70-90% of humidity was maintained in the glass house for the first few days. Under these conditions it was to recover almost hundred percent plantlets.

Results and Discussion

The choice of suitable explant source as starting material for transgenic plant production is one of the most important factor. Although some reports used immature derived calli (Dong *et al.*, 1996), but others found that calli initiated from scutellum of mature seeds were excellent starting material for transformation of rice by *Agrobacterium* due to their compactness (Hiei *et al.*, 1994., Rashid *et al.*, 1996., Toki *et al.*, 1997., Lee *et al.*, 1999, Rashid *et al.*, 2001, 2003, 2004., Cho *et al.*, 2004). In the present study calli derived from mature seeds of Basmati rice cv. 385 and Super basmati were compact.

Sterilization is the major factor, which affects the tissue culture. Clorox (Sodium hypo chlorite) was used as a surface sterilization agent, played very important role in the germination of seeds. Seeds of both varieties, Basmati-385 and Super basmati treated with different concentrations of Clorox for 20 minutes for sterilization showed that 100% Clorox when applied for 20 minutes gave 97-98% sterilization but no healthy callus was formed and it affected the growth of callus, Whereas 50% Clorox gave 78.5% sterilization in Super Basmati and 70% sterilization in Basmati 385 with the formation of large, granular, healthy callus. With the increased concentration of Clorox, the growth of callus decreased because at high Clorox concentration there are more chances of the tissue to be destroyed. Clorox 50% was found more suitable for sterilization without browning and inhibition of germination. Our results are nearly similar to Li *et al.*, (1992) who reported that sterilization of seeds with 45%(v/v) Clorox for 30 minutes were effective (Table 1).

Table 1. Effect of Clorox on sterilization in rice seeds for 20 minutes for callus Induction.

| Clorox | Super Basmati | | Basmati 385 | |
|--------|---------------|--------|-------------|--------|
| | Frequency % | Growth | Frequency% | Growth |
| 100% | 98 | - | 97 | - |
| 80% | 95 | + | 88.7 | + |
| 70% | 89 | + | 77 | + |
| 60% | 80 | + | 75.0 | + |
| 50% | 78.5 | ++++ | 70 | +++ |
| 40% | 50 | +++ | 34 | ++ |

- = No growth, + = Poor, ++ = Average, +++ = Good, ++++ = Excellent

Selected utilization of basal medium, growth regulators, carbon sources, and amino acids are important considerations for improving efficiencies of callus induction and shoot regeneration. Especially, shoot regeneration from the callus, the final stage for producing transformants. The N₆ basal media supplemented with 2-6 mg l⁻¹ 2,4-dichlorophenoxyacetic acid (2,4-D) for embryogenic callus induction and 0.5-10 mg l⁻¹ 6-benzylaminopurine (BAP) combined with 0.02-1 mg l⁻¹ α-naphtalenacetic acid (NAA) for shoot regeneration have been selectively used depending on rice cultivars (Jiang *et al.*, 2000; Cho *et al.*, 2004).

For obtaining transgenic plants of rice with scutella derived embryogenic callus (Fig. 1A-C), N₆ media (Hiei *et al.*, 1994; Rashid *et al.*, 1996) have been selectively used depending on genotype of rice. A number of reports have emphasized the importance of the type of calli for regeneration efficiency in many monocot plants such as maize (Tomes & Smith, 1995) and rice (Lee *et al.*, 2002). In the present study N₆ media was used for callus induction and callus growth. Sterilized seeds were placed on these media for three weeks. Both varieties showed callus induction and growth with 2 mg l⁻¹ 2, 4-D. Embryogenic callus was selected on the basis of their physical properties as described by Cho *et al.*, 2004. Morphology of the embryogenic callus was slick, yellowish-compact type with globular shape, soft and friable. Both the varieties were good for callus induction. However, Super basmati has more potential for callogenesis. The significant difference between two varieties for callogenesis under the same nutritional condition indicate that callus induction efficiency is genotype specific. These findings are in accordance with the reports of Abassi *et al.*, (2000) and Khanna & Raina (1998), Nasreen & Mohammad (2000). The callus of Super basmati was found more embryogenic than Basmati-385. Callus induction and growth frequency was observed as 93.3% in Super Basmati and 90.2 % in Basmati 385. Our results are contradictory to Rashid *et al.*, (2001) who reported that callus induction frequency of Super Basmati on N₆ medium was 47.7%. Comparing callus induction and growth frequency of two varieties on N₆ media proved that Super Basmati was good as compared to Basmati 385 for callus induction (Table 2). Further these calli were placed on N₆ media with 2 mg l⁻¹ 2, 4-D for growth and proliferation. After two weeks, it was observed that 76% calli showed growth and increased 2-3 times in size (Table 3, Fig. 1-D)

On regeneration media, embryogenic callus tended to show positive responses related with regeneration for green tissue formation or green spotting and shoot regeneration or plantlet formation and a negative response for callus browning (Cho *et al.*, 2004). Additionally, the types and concentrations of cytokinins combined with NAA have been considered to be important factors on shoot regeneration of both indica (Xue & Earle, 1995) and Japonica rice (Lee *et al.*, 2002). Supplementation of higher level of

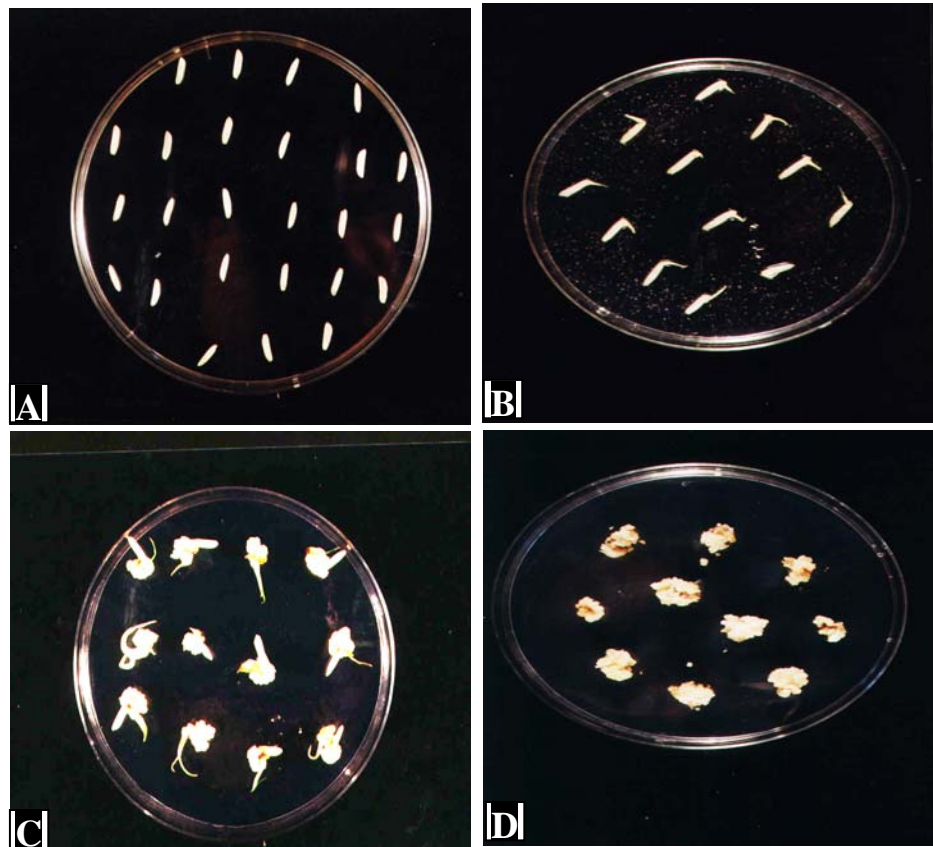


Fig. 1. Callus induction from seed as a explant source

- A. Seeds of rice cv. Bas 385 on N_6 media
- B. Arising of calli from scutellum
- C. Three week old scutellum derived calli
- D. Calli on maintenance media.

BAP was more effective on regeneration of rice (Lee *et al.*, 2002, Xue & Earle, 1995). In the present study, maintained calli were transferred to regeneration medium containing basic MS salts and vitamins with different concentrations of NAA and BAP i.e. NAA 1 mg l^{-1} and BAP .5,1, 1.25,2, 2.5 and 5 mg l^{-1} . Generally, the callus having high regeneration potential formed green tissue or green spots on the surface of callus with fast growth for 4-7 days of cultivation (Fig. 2-B) and, consequently, 0.2-1.0 cm size shoot emerged at 10-14 days (Fig.2-C). The callus regenerated on all the concentrations of growth hormones, but the highest frequency 90% was observed on NAA 1 mg l^{-1} and BAP 2.5 mg l^{-1} in Super Basmati (Fig. 2- E & F) and 83 % in Basmati 385 in combination of NAA 1 mg l^{-1} and BAP 5 mg l^{-1} . (Fig. 2-D).

Table 2. Comparison of callus induction, callus growth frequency of Basmati rice varieties i.e., Basmati 385 and Super basmati on N₆ medium supplemented with N₆ salts and vitamins, 30g/l sucrose, 2mg/l 2,4-D and 4g/l gelrite.

| # of Exp. | # of seeds | Callus induction | | | Callus growth | | | Percentage of callus growth | | |
|----------------|------------|------------------|--------------|--------------|---------------|------------|------------|-----------------------------|------------|------------|
| | | Bas 385 | Super Bas. | Super Bas. | Bas. 385 | Super Bas. | Super Bas. | Bas. 385 | Super Bas. | Super Bas. |
| 1 | 240 | 218 | 199 | 203 | 153 | 84% | 63.7% | | | |
| 2 | 240 | 215 | 222 | 210 | 216 | 87.5% | 90% | | | |
| 3 | 240 | 233 | 208 | 225 | 183 | 90.8% | 76.2% | | | |
| 4 | 240 | 189 | 150 | 167 | 123 | 69.5% | 51.2% | | | |
| 5 | 240 | 236 | 232 | 196 | 224 | 81.6% | 93.3% | | | |
| Average | 240 | 218.2 | 202.2 | 198.8 | 179.8 | 82% | 74% | | | |

Table 3. Maintenance of embryogenic calli of Basmati 385 on N₆ media supplemented with 2 mg l⁻¹ 2, 4-D.

| # of calli | Contamination | | Browning | | Growth and proliferation | | | Percentage of Growth | |
|----------------|---------------|----------|----------|----------|--------------------------|-----------|-------------|----------------------|--|
| | Bas. 385 | Sup. Bas | Bas. 385 | Sup. Bas | Bas 385 | Sup. Bas | Bas. 385 | Sup. Bas. | |
| 25 | 5 | 4 | 3 | 5 | 12 | 21 | 60 | 70 | |
| 30 | 4 | 5 | 6 | 6 | 20 | 24 | 66 | 68.5 | |
| 40 | 5 | 7 | 10 | 4 | 25 | 29 | 62.5 | 72.5 | |
| 40 | 6 | 4 | 5 | 7 | 30 | 30 | 75 | 75 | |
| Average | 5 | 5 | 6 | 5 | 21 | 26 | 65.8 | 71.5 | |

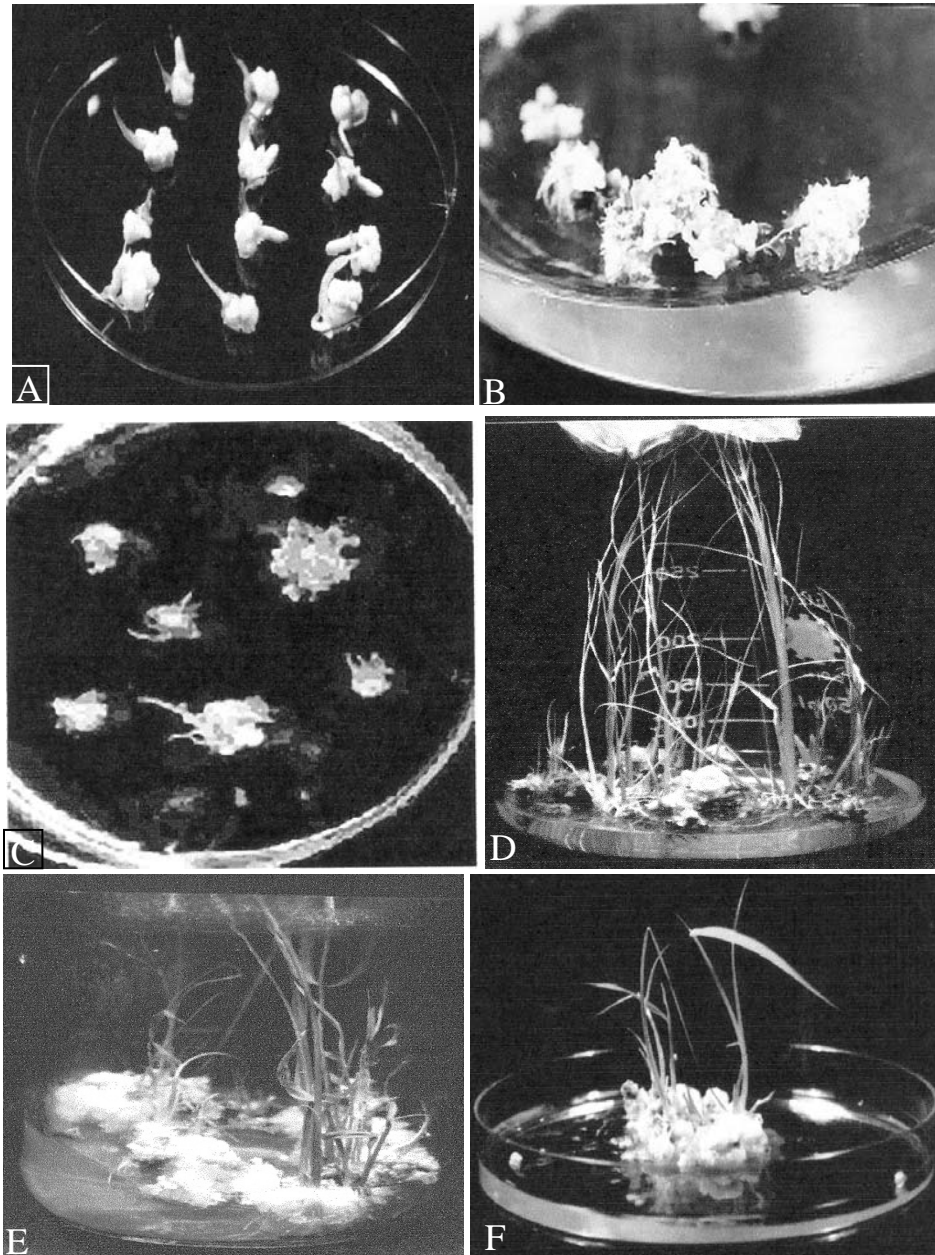


Fig. 2. Regeneration from scutellum derived calli.

A. Three week old scutellum derived calli

B. Calli become green on regeneration media

C. Calli showing further growth on regeneration medium

D. Calli showing regeneration on MS media with NAA 1 mg^{-1} and BAP 5 mg^{-1} in Bas. 385

E & F. Calli showing regeneration on MS media with NAA 1 mg^{-1} and BAP 2.5 mg^{-1} in Super Basmati

Our results are different from the reports of Rashid *et al.*, (2001) in which regeneration frequency was 45.3%. Brar *et al.*, (1985) also reported that the presence of a cytokinin either BAP or Kinetin is essential in promoting plant regeneration from cultured cells (Table 4). Both NAA and BAP was not required for shoot initiation (Azria *et al.*, 2000). Pons *et al.*, (2000) reported that BA yielded more shoots than kinetin in all varieties while in case of using auxin NAA and IAA, it depends on the varieties. Analysis of variance indicates that there are significant differences of regeneration efficiencies of two rice cultivars observed between the supplementation of BAP. As carbon source and osmotic regulator, sucrose has been most acceptable for *in vitro* culture of rice. However maltose (Zhang, 1995) and sorbitol (Rashid *et al.*, 1996) were substituted in the media for callus induction and shoot regeneration. Even though sorbitol is a major factor for embryogenic callus formation in the monocot plant such as maize (Swedlund & Locy, 1993). Cho *et al.*, (2004) reported that growth of callus is stimulated and multiple shoots of regenerated plants could be obtained in higher frequency by supplementation of sorbitol in combination with sucrose or maltose under 5 mg l⁻¹ kinetin. The supplementation of sorbitol and prolin in the medium was attributed to be more effective to obtain regenerated plants of rice rather than utilization of the medium containing sucrose or maltose alone. Yang *et al.*, (1999) reported that proliferation and regeneration of rice callus were suppressed by sorbitol, in contrast controversy to that of Kishor & Reddy (1986) who reported that sorbitol has been used for restoring and enhancing the plant regeneration ability of rice callus. Swedlund & Lucy (1993) also reported that sorbitol, as osmoticum imparted beneficial effect on embryogenesis in cereal crops. In this study sorbitol supplementation in combination with sucrose critically stimulated the growth rate of rice calli. The green spots appeared after 3-4 days of transferring the rice calli on regeneration medium.

Sorbitol as the osmoticum was found not necessary for the induction and growth of rice callus. When seeds of both varieties were inoculated on the N₆ Media (CIM) supplemented with 2,4-D and sorbitol, there was no induction and growth of callus formation. Calli showed necrosis after one week of inoculation (Table 5). Our results are similar to the reports of Alkhayri *et al.*, (1996) who used MS medium with 2,4-D and Kinetin either with sucrose alone or combined with sorbitol. It was reported that medium without sorbitol enhanced callus growth and callus proliferation as compared to sucrose-sorbitol combination. However, the sucrose-sorbitol combination improved regeneration of rice cultures.

Twenty two plantlets of Super basmati and 18 of Basmati 385 have been established on ordinary soil contained in pots by maintaining a high percentage of humidity in the glass house at 25°C and were assessed for their growth by using two parameters i.e., height and tiller. As compared to control, they showed less growth due to sensitivity and lack of development of efficient root system for water and nutrient absorption from the soil (Table 6). The somaclones were short in height as it was reported by Su *et al.*, (1992).

The present study showed that now it is possible to obtain high regeneration frequency of up to 90% from Super Basmati and 83% from Basmati 385, as compared to other varieties. This high regeneration frequency will lead to improve these varieties by genetic transformation technology.

Table 4. Comparison of regeneration efficiencies of rice calli of the two rice cv. Basmati 385 and Super basmati on MS media supplemented with 30g⁻¹ Sucrose, 2 g⁻¹ Casine hydrolysate, 30 g⁻¹ Sorbitol with different treatments of NAA and BAP.

| Number of Treatment | Callus browning | | Callus differentiation | | # of green spots | | Plant formation | | % frequency of plant formation | |
|--|--|----------|------------------------|----------|------------------|----------|-----------------|----------|--------------------------------|----------|
| | Bas. 385 | Sup. Bas | Bas. 385 | Sup. Bas | Bas. 385 | Sup. Bas | Bas. 385 | Sup. Bas | Bas. 385 | Sup. Bas |
| | NAA* 0mg ⁻¹ + BAP** 0mg ⁻¹ | 0.00 h | 0.00 h | 0.00f | 0.00f | 0.00e | 0.00e | 0.00f | 0.00f | 0.00 |
| NAA 1mg ⁻¹ + BAP 0mg ⁻¹ | 16.6 a | 14.00 ab | 0.00f | 0.00f | 0.00e | 0.00e | 0.00f | 0.00f | 0.00 | 0.00 |
| NAA 1mg ⁻¹ + BAP 0.5mg ⁻¹ | 3.33 e-h | 6.6 de | 5.00def | 6.67cde | 6.33bc | 7.67bc | 3.33e-f | 2.00def | 16.6 | 10 |
| NAA 1mg ⁻¹ + BAP 1.0mg ⁻¹ | 5.00 d-g | 5.00 d-g | 7.33cde | 7.33cde | 9.67b | 3.333cde | 2.67 e-f | 2.667c-f | 13.3 | 13.3 |
| NAA 1mg ⁻¹ + BAP 1.25mg ⁻¹ | 6.66 de | 1.67fgh | 11.6bc | 9.33cd | 7.67bc | 9.67b | 5.33cd | 5.00cde | 26.6 | 25 |
| NAA 1mg ⁻¹ + BAP 2.0mg ⁻¹ | 11.6 bc | 4.00 e-h | 12.00bc | 16.00ab | 6.67bc | 15.33a | 5.00 cd | 13.33b | 25.0 | 66.65 |
| NAA 1mg ⁻¹ + BAP 2.5mg ⁻¹ | 1.33 gh | 3.33 e-h | 80.6a | 12.00bc | 19.33a | 9.667b | 16.67 ab | 6.667c | 83.3 | 33.31 |
| NAA 1mg ⁻¹ + BAP 5.0mg ⁻¹ | 3.00 e-h | 0.00h | 16.67 ab | 19.00a | 18.67a | 19.33a | 16.00ab | 18.00a | 80 | 90.0 |
| NAA 2mg ⁻¹ + BAP 1.0mg ⁻¹ | 6.00 def | 8.33cd | 2.00ef | 4.00def | 1.67de | 5.00bcd | 1.00ef | 3.33d-f | 5 | 16.66 |
| Mean= | 5.963 | 4.778 | 8.148 | 8.259 | 7.778 | 7.778 | 5.556 | 5.667 | | |
| Coefficient of variance= | 42.33% | | 34.87% | | 32.40% | | 38.65% | | | |

Coefficient of variance (ANOVA) indicate that, here is no significant difference between two varieties tested for regeneration in our studies.
* α -Naphthalenacetic acid (NAA), **6-benzylaminopurine (BAP)

Table 5. Comparison of effect of sorbitol on the induction of callus on N₆ media supplemented with 2,4-D 2mg l⁻¹, sucrose 3%, 0.6% agar and 0.3% sorbitol and without sorbitol.

| Media with Sorbitol (0.3 %) | | | | | | | | |
|--------------------------------|-------------|-----|----------|---------|---------------|-----|----------|---------|
| No. of Seeds | Basmati 385 | | | | Super Basmati | | | |
| | CI | CG | Browning | % freq. | CI | CG | Browning | % freq. |
| 192 | 150 | 40 | 42 | 20 | 142 | 25 | 48 | 17.6 |
| Media without Sorbitol (0.3 %) | | | | | | | | |
| 192 | 185 | 180 | 3 | 93 | 175 | 170 | 11 | 88.5 |

CI = Callus Induction, CG = Callus Growth

Table 6. Glasshouse assessment of the somaclones of Basmati rice.

| No. of plants | Basmati 385 | | | | Super Basmati | | | |
|----------------|--------------|---------------|-------------|----------------|---------------|---------------|-------------|----------------|
| | Control | | Somaclone | | Control | | Somaclone | |
| | Height (cm) | No. of tiller | Height (cm) | No. of tillers | Height (cm) | No. of tiller | Height (cm) | No. of tillers |
| 1 | 105.5 | 13 | 32.5 | 4 | 65 | 5 | 20 | 1 |
| 2 | 99.9 | 16 | 44.9 | 7 | 76.5 | 9 | 43 | 6 |
| 3 | 150 | 19 | 89.5 | 10 | 88.3 | 8 | 50.6 | 4 |
| Average | 118.4 | 16 | 55.6 | 7 | 76.6 | 7 | 40.8 | 3.6 |

References

- Abbasi, F.M., H. Rashid and A. Qurashi. 2000. Regeneration efficiency and embryogenic callus production of three cultivars of rice. *Pak. J. Agric. Res.*, 16(2): 97-99.
- Al-Khayri, J.M., C.E. Shamblin, R.W. McNew and E.J. Anderson. 1996. Callus Induction and plant regeneration of US rice genotypes as affected by medium constituents. *In Vitro Plant*; 32(4): 227-32, Department of Plant Pathology, University of Arkansas, Fayetteville, AR. 72701, USA
- Anonymous. 2003. Economic survey 2002-2003. Economic Advisor's Wing, Finance Division, Government of Pakistan, Islamabad.
- Azria, D. and P.L. Bhalla. 2000. Plant regeneration from mature embryo derived calli of Australian rice (*Oryza sativa L.*) varieties. *Aus. J. Agri.*, 51: 305-312.
- Bajaj, Y.P.S. and M. Bidani. 1980. Differentiation of genetically variable plants from embryo-derived callus cultures of rice. *Phytomorphology*, 30(2-3): 290-294
- Brar, D.S., D.H. Ling and S. Yoshida. 1985. Plant regeneration from somatic cell culture of some IR varieties of rice. *In: Biotechnology in International Agricultural Research*, International Rice Research Institute, Los Banos, Philippines, p. 3.
- Chen, L., S. Zhang, R.N. Beachy and C.M. Fauquet. 1998. A protocol for consistent large-scale production of fertile transgenic rice plants. *Plant Cell Rep.*, 18: 25-31.
- Cho, J.H., J.Y. Lee and Y.W. Kim. 2004. Improvement of shoot regeneration from scutellum - derived callus in rice. *Korean J. Crop. Sci.*, 49(1): 52-60.
- Chu, C.C., C.S. Wang, C.C. Sun, C. Hsu, K.C. Yin and C.Y. Chu and F.Y. Bi. 1975. Establishment of an efficient medium for anther culture of rice through comparative experiments on the nitrogen sources. *Sci. Sinica.*, 18: 659-668.
- Dong, J., W. Teng, W.G. Buchholz and T.C. Hall. 1996. *Agrobacterium* mediated transformation of Javanica rice. *Mol. Breeding*, 2: 267-276.
- Gonzalez, M.C. 2000. Effects of different growth regulators on *in vitro* culture of rice cultivar. *Tropicales*, 21(1): 23-27.
- Hashizume, F., T. Tsuchiya, M. Ugaki, Y. Niwa, N. Tachibana and Y. Koyama. 1999. Efficient *Agrobacterium*-mediated transformation and the useless of a synthetic GFP reporter gene in leading varieties of japonica rice. *Plant Biotech.*, 16: 397-401.

- Hiei, Y., S. Otha, T. Komari and T. Kumanshiro. 1994. Efficient transformation of rice (*Oryza sativa* L.) mediated by *Agrobacterium* and sequence analysis of the boundaries of the T-DNA. *The Plant Journal*, 6(2): 271-282.
- Jiang, J., S.F. Linscobe, J. Wang and J.H. Oard. 2000. High efficiency transformation of U.S. rice lines from mature seed derived calli and segregation of Glufosinate resistance under field condition. *Crop Sci.*, 40: 1729-1741.
- Khanna, H.K. and S.K. Raina. 1998. Genotype x culture media interaction effects on regeneration response of three rice cultivars. *Plant Cell Tissue & Organ Cult.*, 52(3): 145-153.
- Kishore, P.B.K. and G.M. Reddy. 1986. Retention and revival of regenerating ability by osmotic adjustment in long-term cultures of four varieties of rice. *J. of Plant Physiol.*, 126: 49-54.
- Lee, K., H. Jeon and M. Kim. 2002. Optimization of a mature embryo-based *in vitro* culture system for high frequency somatic embryogenic callus induction and plant regeneration from japonica rice cultivars. *Plant Cell Tiss. Org. Cult.*, 71: 237-244.
- Lee, S.H., Y.G. Shon., S.I. Lee., C.N. Kim., J.C. Koo., C.O. Lim., Y.J. Choi., C.D. Han., C.h. Chung., Z.R. Choe and M.J. Cho. 1999. Cultivars variability in the *Agrobacterium* – rice cell interaction and plant regeneration. *Physiologia Plantarum*, 107: 338-345.
- Li, X.Q., C.N. Lin, S.W. Ritchie, J. Peng, S.B. Gelvin and T.K. Hodges. 1992. Factor influencing *Agrobacterium*-mediated transient expression of GUS in rice. *Plant Mol. Biol.*, 20: 1037-1048.
- Marassi, M.A., O.A. Bovo, A. Socchi and L.A. Mroginski. 1996. Cytokining in the callus induction medium for plant regeneration of rice. *Phyton International J. of Exp. Bot.*, 59(1-2): 155-160.
- Murashige, T. and F. Skoog. 1962. A revised medium for rapid growth and bioassay with tobacco tissue culture. *Physiol. Plant*, 15: 473-497.
- Nasreen, S. and Mohammad. 2000. Effect of growth hormones on callogenesis in Basmati rice. *Pak. J. Biol. Sci.*, 3(12): 2213-2215.
- Navraj, K., M.S. Gill, G. Raman, T.S. Bharaj, S.S. Gosal, N. Kaur and R. Gill. 1999. Factors enhancing somatic embryogenesis and high frequency plant regeneration in rice. *Crop Improvement*, 26(1): 23-27.
- Pons, M.J., V. Marfa, E. Mele and J. Massager. 2000. Regeneration and genetic transformation of Spanish rice cultivars using mature embryo. *Euphytica*, 114: 117-122.
- Rashid, H., S. Yokoi, K. Toriyama and K. Hinata. 1996. Transgenic plant production mediated by *Agrobacterium* in Indica rice. *Plant Cell Rep.*, 15: 727-730.
- Rashid, H., S.Y.A. Bokhari and A. Quraishi. 2001. Callus induction, regeneration and hygromycin selection of rice (Super Basmati). *Online J. Biol. Sci.*, 1: 1145-1146.
- Rashid, H., S.N. R. Bokhari, Z. Chauhry and S.M.S. Naqvi. 2003. Studies on genotype response to callus induction from three basmati cultivars of rice (*Oryza sativa* L.). *Pak. J. Biol. Sci.*, 6(5): 445-447.
- Rashid, H., M. Saleem, Z. Chauhry, S.T. Gilani and A.S. Qureshi. 2004. Studies on developing a high regeneration from seed derived calli of rice (*Oryza sativa* L.). *Pak. J. Biol. Sci.*, 7(2): 273-276.
- Salim, M., M. Akram, M.E. Akhtar and M. Ashraf. 2003. *Rice, A production Handbook*, Balanced Fertilization for Maximizing Economic crop yields. Pakistan Agricultural Research Council, Islamabad. p.1
- Su, R., Rudert. M.L and T.K. Hodges. 1992. Fertile indica and japonica rice plants regenerated from embryogenic haploid suspension culture. *Pl. Cell. Rep.* 12: 45-49.
- Swedlund, B. and R.D. Locy. 1993. Sorbitol as the primary source for the growth of embryogenic callus of maize. *Plant Physiol.*, 103: 1339 -1346.
- Tamura, S. 1968. Shoot formation in calli originated from rice embryo. *Proc. Jpn. Acad.* 44: 543-548.
- Toki, S. 1997. Rapid and efficient *Agrobacterium*-mediated transformation in rice. *Plant Mol. Biol. Reporter*, 15(1): 16-21.
- Tomes, D., T. and O.S. Smith. 1985. The effect of parental genotype on initiation of embryogenic callus form elite maize (*Zea mays* L.) germplasm. *Theor. Appl. Genet.*, 70: 505-509.

- Valdez, M., M. Monoz, J.R. Vega and A.M. Espinoza. 1997. Plant regeneration of indica rice from mature embryo derived calli. *Revistade Biologia Tropical.*, 44(3): 13-21.
- Vasil, I.K., V. Vasil, V.C. Lu, P. Atkins, Z. Haydu and D. Wang. 1982. Somatic embryogenesis in cereals and grasses, In: *Variability in plants regenerated from tissue culture*, (Eds.): E. Earle and Y. Demarly. Praeger Press, New York, p. 3.
- Xie, J.H., M.W. Gao, Q.H. Cai, X.Y. Chens, Y.W. Shen and Z.Q. Liang. 1995. Optimized growth regulator combination in Japonica rice. *Plant Cell Tissue and Organ Culture*, 42(3): 245-250.
- Xue, Q. and E.D. Earle. 1995. Plant regeneration from protoplasts of cytoplasmic male sterile lines of rice (*Oryza sativa* L.). *Plant Cell Rep.*, 15: 76-81.
- Yamada.Y. 1977. Tissue culture studies on cereals. In: *Improvement of plant regeneration from long term cultured calluses of Taipei 309, a model rice variety in vitro studies*. (Eds.): L. Reinert and Y.P.S. Ng, Y.S., Y.D. Zheng, Y. I. Chen, and Y.Y. Jian. 1999. *Plant Cell Tiss. Org. Cult.*, 32: 61-206.
- Yang, Y.S., Y.D. Zheng, Y.L.Chen and Y.Y. Jian. 1999. Improvement of plant regeneration from long-term cultured calluses of Taipei 309, a model rice variety *in vitro* studies. *Plant Cell Tiss. Org. Cult.*, 57: 199-206.
- Zhang, S. 1995. Efficient plant regeneration from indica (group 1) rice protoplasts of one advanced breeding line and three varieties. *Plant Cell Rep.*, 15: 68-71.

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