

## GROWTH RESPONSE OF *VIGNA AMBACENSIS* L. SEEDLING TO THE INTERACTION BETWEEN NITROGEN SOURCE AND SALT STRESS

MOHAMMED N. ALYEMENI

*Department of Botany and Microbiology,  
College of Science, King Saud University,  
P.O. Box 2455, Riyadh 11451, Saudi Arabia.*

### Abstract

*Vigna ambacensis* plants grown hydroponically in a Hoagland nutrient solution with either nitrate or ammonia as a source of nitrogen and NaCl at 0, 40, 80 and 160 mM concentrations showed reduction in all growth parameters where ammonia was used as compared to nitrate as a nitrogen source. Salt decreased the growth of plants. Plants grown in nitrate were more salt tolerant as compared to ammonia especially at 80 and 160 mM NaCl level. High salinity conditions reduced calcium and potassium contents in shoots with an increase in sodium which was higher (80  $\mu\text{l}$ ) in  $\text{NH}_4^+$ -N as compared with  $\text{NO}_3^-$ -N source.

### Introduction

Salinization of soils is a serious problem in many areas of the world especially in semi-arid and arid regions where excessive soluble salts decrease the osmotic and water potentials of the root media and therefore impairs water uptake by roots. Reduction in growth of glycophytes under salt stress in terms of fresh weight and dry weight production have been reported (Hue & McCall, 1989; Pessarak *et al.*, 1989). Salt stress is reported to adversely affect plant metabolism and cause nutrient imbalance in *Macedamia* seedlings (Hue & McCall, 1989) and *Phaseolus vulgaris* Seemann & Critchley, 1985). Nitrogen is also reported to affect salt stress tolerance in plants like chickpea (Lauter *et al.*, 1981), corn (Pessarak *et al.*, 1989) or wheat (Lewis *et al.*, 1989).

Nitrogen is a macro nutrient element required in plants through all growth stages of Nitrate and ammonium are the two major forms of nitrogen available for plant uptake. Most plants can absorb and utilize both forms of nitrogen, however a number of plant species have shown that growth on nitrate is more rapid than growth on ammonium as the sole nitrogen source (Smith *et al.*, 1990; Gentry *et al.*, 1989; Alfoldi & Pinter, 1992; Addition of small amounts of  $\text{NH}_4^+$  to  $\text{NO}_3^-$  has shown an increase in growth of tomato (Errebhi & Wilcox, 1990) as compared to nitrate alone Herber & Below, 1989). Nitrogen has a significant influence on numerous physiological mechanisms and on the absorption of other ions (Lips *et al.*, 1990; Magalhaes & Wilcox, 1983) Ammonium nitrogen nutrition is associated with reduced cation uptake as compared to nitrate nutrition (Magalhaes & Wilcox, 1983; Mackalon & Sim, 1980).

*Vigna ambacensis*, an annual herb which grows in the south western region of Saudi Arabia, is cultivated for seeds as a source of food. The present study was con-

ducted to evaluate the response of this species to the interaction of salt stress and nitrogen source.

### Materials and Methods

Seeds of *Vigna ambacensis* were collected from plants in the Gizan district, south western of Saudi Arabia and stored at 10°C until used. Prior to germination studies the seeds were allowed to imbibe water overnight at 25°C and then placed on the surface of two layers of Whatman No.1 filter paper moistured with distilled water petridishes. The dishes were placed in an incubator at 26°C under darkness. After 3 days of germination, four uniform seedlings were transferred to 2-L polyethylene container wrapped with aluminum. The containers were filled with half strength Hoagland nutrient solution containing either  $\text{Ca } 9\text{NO}_3)_2$  or  $\text{KNO}_3$  or  $\text{NH}_4)_2\text{SO}_4$  as a source of nitrogen pH was adjusted to 6 by adding drops of diluted NaOH solution. The solution was continuously aerated. The containers were placed in a growth chamber set at 30/25°C day/night temperature and 14/10h day/night cycle with light intensity of 900 lux. The level of the nutrient solution in each container was maintained at 2 L throughout the experimental period by adding distilled water. Nutrient solution was replaced with a full strength Hoagland nutrient solution one week after transplanting with 40 mM NaCl added to each container except in the control. Concentration of NaCl was increased 40 mM daily till the desired concentration (up to 160 mM) was attained. Four replicates were used per treatment and the nutrient solution was changed weekly.

Seven weeks after completion of salt treatment, the plants were harvested and separated in to roots stems and leaves. The length of roots and stems were recorded and the total leaf area of each plant was measured. Fresh weight of each plant part was recould and dry weight was obtained after oven drying at 85°C to a constant weight. Dry samples of leaves, and stems were ground into a fine powder separately and digested using sulphuric acid and a mixture of hydrogen peroxide and perchloric acid 1:1 v/v and assayed for minerals ions using atomic absorption spectrophotometer (Model Pye Unicam-Sp. 6560 UV/VIS). Phosphorus was determined calorimetrically suing Ascorbic acid method (Watanabe & Olsem, 1965). Total nitrogen was also determined by the microkjeldahl technique (Jackson, 1958).

### Results

Low NaCl concentration (40 mM) had no effect on plant survival in  $\text{NO}_3^-$  - grown plants which reduced to 63% in  $\text{NH}_4^+$  grown plants (Fig.1). At 160 mM, 50% of  $\text{NO}_3^-$  grown plants survived till the end of the experimental period, while all the  $\text{NH}_4^+$ -grown plants died few days after the completion of salt treatment.

Nitrogen form had a significant effect on fresh weight and dry weight production of all plants parts organs (root, stem and leaves) (Table 1). Plant growth in terms of fresh weight and dry weight was lower in  $\text{NH}_4^+$  grown plants as compared to  $\text{NO}_3^-$  grown plants. Total fresh and dry weight of  $\text{NH}_4^+$ -grown plants was 69.7% and 76.8% of that of  $\text{NO}_3^-$  grown plants, respectively. In both  $\text{NO}_3^-$  - and  $\text{NH}_4^+$  - grown

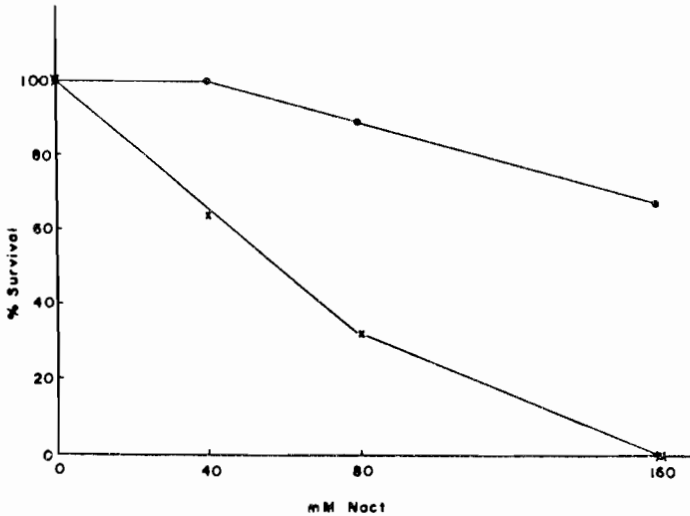


Fig. 1. Effect of interaction between salt stress and nitrogen source on plant survival.

o ----- o nitrate  
x ----- x ammonium

plants, the addition of NaCl reduced total plant fresh weight where salinity of 40 mM slightly increased the fresh weight of  $\text{NH}_4^+$ -grown plants. At 40 mM NaCl, total fresh weight of  $\text{NO}_3^-$ -grown plants was slightly lower (-5%) than that of  $\text{NH}_4^+$ -grown plants whereas at 80 mM NaCl, total fresh weight of  $\text{NH}_4^+$ -grown plants was only 77% of that of  $\text{NO}_3^-$ -grown plants. In  $\text{NO}_3^-$  grown plants, total dry weight declined to 56.8, 49.6 and 25.2% at 40, 80 and 160 mM salt concentration, respectively, whereas, in  $\text{NH}_4^+$ -grown plants, total dry weight reduced to 80.4 and 46.3% at 40 and 80 mM NaCl, respectively.

Fresh and dry weights of root were not affected by salt concentration up to 80 mM and in some cases the weights slightly increased as compared to control plants. At 160 mM NaCl, root fresh and dry weight reduced significantly in  $\text{NO}_3^-$ -grown plants to 50.9 and 45.6% respectively as compared to control plants. The shoot: root ratio in both ammonium and nitrate fed-plant, decreased respectively in response to salinity, indicating that shoot growth was more severely affected by salinity than root growth. Nitrogen form growth media also influenced the length of root and stem and total leaf area. Root/Stem length and total leaf area were lower in  $\text{NH}_4^+$ -grown plants as compared to  $\text{NO}_3^-$  grown plants. Root length, stem length and total leaf area were respectively reduced by 76.7%, 68.9 and 78.8% in  $\text{NH}_4^+$ -grown plants (Table 2).

Salinity also had some effects on the mineral compositions as influenced by nitrogen source. A decrease in shoot mineral contents, especially for calcium potassium phosphorus and nitrogen of both nitrogen treatment as a result of the increase in salinity level, was observed (Table 3). Sodium shoot content increased from 144 ppm at 0.0 NaCl level to 606 at 80 mM in  $\text{NH}_4^+$ -N treatment, and from 81.4 ppm at 0.0

**Table 1. Effect of nitrogen source and NaCl concentration on fresh and dry weight of *Vigna ambacensis*.**

Nitrogen Source	NaCl Conc. (mM)	root $x \pm S.D$	stem $x \pm S.D$	leaves $x \pm S.D$	shoot	Total	shoot root ratio
<b>Fresh weight (g/plant)</b>							
Nitrate	0.0	0.55 0.014	1.81 0.02	1.73 0.05	3.54	4.09	6.4
	40.0	0.57 0.01	1.20 0.01	0.96 0.04	2.16	2.73	3.8
	80.0	0.53 0.01	0.90 0.01	0.81 0.04	1.17	2.24	3.2
	160.0	0.28 0.005	0.41 0.05	0.04 0.02	0.81	1.09	2.8
Ammonia	0.0	0.38 0.002	0.14 0.01	1.17 0.03	2.31	2.85	6.1
	40.0	0.46 0.002	0.14 0.01	1.30 0.03	2.44	2.90	5.3
	80.0	0.39 0.002	0.50 0.05	0.78 0.02	1.34	1.73	3.4
	160.0						
<b>Dry weight (mg/plant)</b>							
Nitrate	0.0	29.4 1.0	182.1 2.0	16.0 5	242.1	371	8.23
	40.0	29.3 1.0	93.3 1.0	88.2 3	181.5	210.8	6.19
	80.0	29.3 1.0	74.9 1.0	79.8 3	153.7	184	5.25
	160.0	13.4 0.6	81.0 0.5	49.0 2	80.0	93.4	5.9
Ammonia	0.0	22.5	12.9 1.0	134 3	26.3	285	11.9
	40.0	28.5 0.8	79.3 0.5	121 3	200.3	229	7.03
	80.0	21.2 0.8	38.8 0.05	72 2	110.8	132	5.23
	160.0						
<b>Fresh weight (%)      Dry weight (%)</b>							
Nitrate	0.0		---		---		
	40.0		66.75		66.82		
	80.0		54.77		19.60		
	160.0		26.65		25.18		
Ammonia	0.0		-		-		
	40.0		101.75		80.35		
	80.0		60.70		46.32		
	160.0						

in NaCl level to 487 pp at 80 mM  $\text{NO}_3\text{-N}$  indicating more (25% high) accumulation of sodium cation in  $\text{NH}_4\text{-N}$  treated plants. A slight difference in potassium and phosphorus ions was observed with both nitrogen sources (Table 3). However, shoot calcium content at  $\text{NO}_3\text{-N}$  source was twice more than that of  $\text{NH}_4\text{-N}$  source.

**Table 2. Effect of nitrogen source on root and stem length and total leaf area.**

Nitrogen source	NaCl Conc. (mM)	Root Length (cm) $\bar{x} \pm S.D.$	Stem Length (cm) $\bar{x} \pm S.D.$	Total Leaf Area (cm) $\bar{x} \pm S.D.$
Nitrate	0.0	22.80 1.5	62.92 2	132.27 3
	40.0	18.25 0.7	31.5 2	69.02 2
	80.0	20.79 0.7	18.33 0.5	52.89 1.5
	160.0	17.5 0.6	11.81 0.4	---
Ammonia	0.0	17.54 0.5	18.17 3	104.19 4
	40.0	19.9 0.8	18.43 0.6	85.8 2.5
	80.0	19.3 0.6	12.2 0.5	10.97 1.0
	160.0			

**Discussion**

The present study showed that both the nitrogen forms (nitrate and ammonium) caused differences in all growth parameters of *Vigna ambacensis*. Growth greatly was more reduced in  $NH_4^+$  - grown plants as compared to  $NO_3^-$  - grown plants which is similar to in pearl millet (Smith *et al.*, 1990) wheat Gentry *et al.*, 1989); tomato (Magalhaes & Wilcox, 1983) and maize (Alfoldi & Pinter, 1992). Reduction of plant growth by  $NH_4^+$  as compared to  $NO_3^-$  could be due to an inadequate accumulation of mineral cations and organic anions (Salsac *et al.*, 1987) or might due to a reduction in water uptake which may involve the effect of  $NH_4^+$  on membrane permeability

**Table 3. The effect of salinity levels and nitrogen sources on mineral composition in shoot of *Vigna ambacensis*.**

Nitrogen Source	Salt level (mM)	Element concentration (ppm)				N(%)
		Ca	K	Na	P	
$(NH_4)_2SO_4$	0.0	131.0	612.0	144.0	676.0	24.2
	40.0	126.7	712.0	416.0	656.0	28.1
	80.0	87.6	520.0	606.0	608.0	27.5
	160.0					
$KNO_3$	0.0	313.0	761.0	81.4	680.0	16.1
	40.0	311.8	639.0	267.7	644.0	17.3
	80.0	270.0	586.7	487.8	548.0	15.6
	160.0	163.3	602.9	517.0	481.0	16.2

(Macklon & Sim, 1980). Water stress has also been observed in plants supplied with  $\text{NH}_4^+$  (Quebedaux & Ozburn, 1973;

Addition of NaCl to the nutrient solution decreased the growth of *V. ambaceusis* either grown in  $\text{NO}_3^-$  or  $\text{NH}_4^+$  as a source of nitrogen. Reduction of plant growth under salt stress is usually attributed to osmotic stress due to external water potential, specific ion effect on metabolic process, or due to nutrient deficiency Seemann & Critchley, 1985; Hue & McCall, 1989). Reduction in growth of *Vigna ambacensis* by salt stress may not be due to  $\text{Na}^+$  toxicity only since growth was more adversely affected in  $\text{NH}_4^+$  - grown plants than  $\text{NO}_3^-$  - grown plants and  $\text{NH}_4^+$  - grown plants should accumulate more  $\text{Na}^+$  than  $\text{NO}_3^-$  - grown plants to help in regulating the cytoplasmic pH because the  $\text{NO}_3^-$  - grown plants grow faster than  $\text{NH}_4^+$  - grown plants. than  $\text{NO}_3^-$  grown plants and  $\text{NH}_4^+$  - grown plants should accumulate more  $\text{Na}^+$  than  $\text{NO}_3^-$  - grown plants to help in regulating the cytoplasmic pH because the  $\text{NO}_3^-$  - grown plants grow faster than  $\text{NH}_4^+$  - grown plants.  $\text{NH}_4^+$  - grown plants were more sensitive to NaCl salinity than  $\text{NO}_3^-$  - grown plants especially at 80 and 160 mM, because  $\text{NH}_4^+$  ions compete with the absorption of other cations. This preassembly leads to a reduction in the leaf area of the plants and consequently in sufficient carbohydrates production to support growth. Similar results have been reported for rice (Youngdahl *et al.*, 1982). Where the toxic effect of the  $\text{NH}_4^+$ -N on plant growth attributed to factors such as interference of NaCl ions with  $\text{NH}_4^+$  assimilation in the root, deficiencies in the plant and acidification of the root environment. The result of the present study is in agreement with those of Lewis *et al.*, (1989) and Leidi *et al.*, (1992) that  $\text{NH}_4^+$  - N grown plants were highly susceptible to salinity. Higher growth rate of  $\text{NO}_3^-$  - plants might increase salt stress tolerance due to the dilution and distribution of absorbed toxic ions to various plant parts by faster growth. Better growth of  $\text{NH}_4^+$  grown plants at 4-0 mM NaCl as compared to  $\text{NO}_3^-$  - plants at the same salt level might be part due the less competition between plants for nutrients since 30% of the plants died whereas all  $\text{NO}_3^-$  - grown plants survived till the end of the experimental period.

Under high salinity conditions, plants generally have reduced root is Okusnya, 1979, 1980). Poor root growth may result from the reduction in the length of the root tip elongation zone, the length of the epidermal cells and a decline in the rate of cell production (Zidan *et al.*, 1990). The reduction in root growth lead to a decrease in its surface area, and therefore, nutrient uptake, higher water potential which restrict better root and shoot growth and thus *Vigna* seedling may not be able to adjust osmotically at high salinity level which is similar to the observations of Greenway (1968) for glycophytes.

## References

- Afoldi, Z and L. Pinter. 1992. Accumulation and partition of biomass and soluble carbohydrates in maize seedlings as affected by source of nitrogen, nitrogen concentration and cultivar. *Journal of Plant Nutrition*, 15: 2567-2583.
- Errebhi, M and G.E. Wilcox. 1990. Tomato growth and nutrient uptake pattern as influenced by nitrogen form. *Journal of Plant Nutrition*, 13: 1031-1043.

- Gashaw, L and L.M. Mugwiva. 1981. Ammonium-N and nitrate-N effects on the growth and mineral composition of triticale, wheat and rye, *Agronomy Journal*, 73: 47-51.
- Gentry, L.E., X.T. Wang and F.F. Below. 1989. Nutrient uptake by wheat seedlings that differ in response to mixed nitrogen nutrition. *Journal of Plant Nutrition*, 12: 363-373.
- Greenway, H and R. Munns. 1980. Mechanisms of salt tolerance in non halophytes. *Annual Review of Plant Physiology*, 31: 149-190.
- Herber, J.A. and F.E. Below. 1989. Mixed nitrogen nutrition and productivity of wheat grown in hydroponics. *Annals of Botany*, 63: 643-649.
- Hue, N.V. and W.W. McCall. 1989. Soil salinity and the growth of Mucedamia seedlings. *Journal of Plant Nutrition*, 12: 449-464.
- Jackson, M.L. 1958. Soil chemical analysis, Prentice-Hall International Inc. London. 534 p.
- Kirkby, E.A. and K. Mengel. 1967. Ionic balance in different tissues of the tomato plant in relation to nitrate, urea or ammonium nutrition. *Plant Physiology*, 42: 6-14.
- Lauter, D.J., D.N. Mengel. 1967. Ionic balance in different tissues of the tomato plant in relation to nitrate, urea or ammonium nutrition. *Plant Physiology*, 42: 6-14.
- Lauter, D.J., D.N. Munns and K.L. Clarkin. 1981. Salt response of chickpea as influenced by N supply. *Agronomy Journal*, 73: 961-966.
- Leidei, E.O., M. Silberbush, MIM Soares and S.H. Lips. 1992. Salinity and nitrogen nutrition studies on peanut and cotton plants. *Journal of Plant Nutrition*, 15: 591-604.
- Lewis, O.A.M., E.O. Leidi and S.H. Lips. 1989. Effect of nitrogen source on growth response to salinity in maize and wheat. *New Phytologist* 111: 155-160.
- Lips, S.H., E.O. Leidi, M. Silberbush, M.I. Soares and O.E.M. Lewis. 1990. Physiological aspects of ammonium and nitrate fertilization. *Journal of Plant Nutrition*, 13: 1271-1239.
- Mackalon, A.E. and A. Sim. 1980. Electrophysiological factors in the influence of nitrate and ammonium ions on calcium uptake and translocation in tomato plants. *Physiologia Plantarum*, 49: 449-454.
- Magalhaes, J.R. and G.E. Wilcox. 1983. Tomato growth and mineral composition as influenced by nitrogen and light intensity. *Journal of Plant Nutrition*, 6: 347-862.
- Mass, E.V and G.J. Hoffman. 1977. Crop salt tolerance-current assessment. *ASCE Journal of irrigation and Drain Division* 103: 115-134.
- Pessarak, M., J.T. Huber and T.C. Tucker. 1989. Dry matter yield, nitrogen absorption and water uptake by sweet corn under salt stress *Journal of Plant Nutrition*, 12: 279-290.
- Quebedaux, B. and J.L. Ozburn. 1973. Effect of ammonium nutrition on water stress, water uptake and root pressure in *Lycopersicon esculentum* Mill. *Plant Physiology*, 52: 677-679.
- Salsac, L., S. Chaillou, J.F. Morot-Gau-dry, C. Lesaint and E. Jolivet. 1987. Nitrate and ammonium nutrition in plants. *Plant Physiological Biochemistry*, 25: 805-812.
- Seemann, J.R. and C. Critchley. 1985. Effects of salt stress on the growth, ion content, stomatal behaviour and photosynthetic capacity of salt-sensitive species, *Phaseolus vulgaris* Plants. 164: 151-162.
- Smith, R.L., H.A. Mills, C.S. Hoveland and W.W. Hanna. 1990. Influence of ammonium: nitrate ratios on the growth and nitrogen uptake of pearl millet. *Journal of Plant Nutrition*, 13: 541-553.
- Wilcox, G.E., C.A. Mitchell, and J.E. Hoff. 1977. Influence of nitrogen form on exudation rate and ammonium amide and cation composition of xylem exudate in tomato. *Journal of America Society Hort Science*, 102; 192-196.
- Woolhouse, H.W. and C.K.K. Hardw. 1966. The growth of tomato seedlings in relation to the form of nitrogen supply. *New Phytologist*, 65: 518-526.

- Yan, F., S. Schubert and K. Mengel. 1992. Effect of low root medium pH on net proton release, root respiration and root growth of corn (*Zea mays* L.) and broad bean (*Vicia faba* L.). *Plant Physiology* 99: 415-421.
- Youngdahl, L.J., R. Pacheco. I.J. Street. P.L.G. Vlek, 1982. The kinetics of ammonium and nitrate uptake by young rice plants. *Plant and Soil*, 69: 225-232.
- Zidan, I., H. Azaizeh and P.M. Neumann. 1990. Does salinity reduce growth in maize root epidermal cells by inhibiting their capacity for cell wall acidification? *Plant Physiology*, 93: 7-11.

(Received for publication 27 November, 1996)